

# **Ecosystem biogeochemistry considered as a distributed metabolic network ordered by maximum entropy production**

Joseph J Vallino

Marine Biological Laboratory  
Ecosystems Center  
Woods Hole, MA 02543  
jvallino@mbl.edu

18 Sep 2009

**Keywords:** Biogeochemistry; Entropy, Free energy, Maximum entropy production; Ecosystem; Metabolic networks; Self organization; Living systems; Dissipative structures, Life, Optimal control.

**Short title:** MEP-based biogeochemistry modeling

**Submitted:** *Phil. Trans. R. Soc. B*, (Jan, 2009)

**Revised:** 18 Sep 2009

## **Abstract**

We examine the application of the maximum entropy production principle for describing ecosystem biogeochemistry. Since ecosystems can be functionally stable despite changes in species composition, we utilize a distributed metabolic network for describing biogeochemistry, which synthesizes generic biological structures that catalyze reaction pathways, but is otherwise organism independent. Allocation of biological structure and regulation of biogeochemical reactions is determined via solution of an optimal control problem in which entropy production is maximized. However, because synthesis of biological structures cannot occur if entropy production is maximized instantaneously, we propose that information stored within the metagenome allows biological systems to maximize entropy production when averaged over time. This differs from abiotic systems that maximize entropy production at a point in space-time, which we refer to as the steepest descent pathway. It is the spatiotemporal averaging that allows biological systems to outperform abiotic processes in entropy production, at least in many situations. A simulation of a methanotrophic system is used to demonstrate the approach. We conclude with a brief discussion on the implications of viewing ecosystems as self organizing molecular machines that function to maximize entropy production at the ecosystem level of organization.

# 1. Introduction

Many of the chemical transformations that occur on Earth are catalyzed by biological systems, and in most cases these biological machines enhance reaction rates by many orders of magnitude over abiotic processes (Falkowski *et al.* 2008). Now that human society can match or exceed many of the natural biogeochemical reaction rates, it is critical that we understand how living systems organize to process energy and mass on local, regional and global scales. Given the importance of biogeochemical cycles on climate and ecological processes, it is surprising that we lack any agreed upon theoretical basis for biogeochemistry. Current biogeochemical models largely take an organismal perspective, where emphasis is placed on understanding and modeling the growth and interaction of the various organisms that comprise a given ecosystem. The biogeochemistry is then a secondary consequence of the complex food web (for aquatic systems) or physiological (for terrestrial systems) dynamics. Unfortunately, other than conservation of mass, and implicit conservation of energy, no fundamental rules or theories govern the equations used to model the organisms and their interactions. While the theory of evolution by natural selection provides a mechanism for self-organization of complex biological structures, the theory is indeterminate in regards to the emergent properties biological systems follow, if any (Peters 1976, Murray, Jr. 2001). Consequently, our biogeochemical models are based purely on imperial observations, where each system is modeled largely as a case study that depends on the organisms present in the particular system under study. As natural or human induced environmental conditions change, the population of the dominate organisms in an ecosystem shift in response, but we have little understanding or predictive capabilities regarding such system reorganization. Consequently, our biogeochemical models are brittle and of dubious

value when predictions are extrapolated beyond observations used to calibrate them. Yet, ironically, extrapolation is often the primary rationale for quantitative modeling.

Dating back to at least Lotka (1922), who proposed that ecosystems organize toward a state of maximum power, there has been a significant amount of work in theoretical ecology that focuses on understanding the governing principles that organize ecosystems. Biological systems are far from equilibrium and contain low entropy ordered structures that are maintained by external energy dissipation (Schrödinger 1944, Morowitz 1968). Accordingly, there has been a great deal of interest in nonequilibrium thermodynamic (or thermodynamically inspired) applications to living systems involving: power (Lotka 1922, Odum and Pinkerton 1955), biomass to maintenance (Margalef 1968), minimum entropy (Prigogine and Nicolis 1971), exergy (Mejer and Jorgensen 1979), ascendancy (Ulanowicz 1986), emergy (Odum 1988), energy dissipation (Schneider and Kay 1994), respiration (Washida 1995, Choi *et al.* 1999), thermodynamic efficiency (Nielsen and Ulanowicz 2000), constructal theory (Bejan 2007), as well as others (Ulanowicz and Platt 1985, Weber *et al.* 1988, Toussaint and Schneider 1998, Jorgensen *et al.* 2000). However, the theories have not gained wide acceptance and are seldom employed to model biogeochemistry. While many of the theories have a similar basis (Jorgensen 1994, Fath *et al.* 2001), they differ sufficiently to cause confusion.

As evident by this special journal issue, there is a renewed interest in the principle of maximum entropy production (MEP) for nonequilibrium systems due to both theoretical and observational research. The original application of MEP dates to Paltridge (1975), who demonstrated that if global heat transport between the tropics and the poles operates at MEP, one can accurately predict meridional heat flux, latitudinal temperatures, and fractional cloud cover; a similar analysis has also been applied to describe the climatology of Mars and Titan (Lorenz *et al.*

2001). Others speculated that ecosystems follow MEP (Ulanowicz and Hannon 1987, Swenson 1989), but it garnered little attention without theoretical and experimental support. Dewar's (2003, 2005) analysis now provides a theoretical basis for the MEP principle, as he derives a provisional proof for MEP for nonequilibrium steady state systems with sufficient degrees of freedom. The general implication of the MEP principle is that systems will organize, within biophysiochemical constraints, so as to maximize the rate of entropy production. MEP is an appealing extension to classic, equilibrium thermodynamics that dictates systems will move to a state of maximum entropy at equilibrium. In essence, MEP indicates systems will attempt to achieve equilibrium via the fastest allowable pathways. Equally desirable, the MEP principle does not distinguish between biotic and abiotic systems so can be applied generally.

In this manuscript we will examine the application of the maximum entropy production principle to describe biogeochemistry. To facilitate this, we will view ecosystems from the perspective of a distributed metabolic network, which will replace the traditional organismal focus. With the new perspective, we will develop, via example with a methanotrophic community, a mathematical framework to describe ecosystem biogeochemistry. We conclude with a discussion on the advantages and limits of the MEP approach.

## **2. Theoretical Framework**

**2.1 Entropy Context.** In this manuscript, entropy will refer to Clausius' (1850) original definition; the energy dissipated when converting internal energy into work. Under constant temperature and pressure, the energy that can be used for work (or drive other reactions) is Gibb's free energy, which accounts for losses due to entropy. Most importantly, if an exergonic chemical reaction occurs but all the energy liberated is dissipated as heat to the surroundings

(i.e., energy is not stored), then all the free energy is converted to entropy at the given temperature. Unlike energy, free energy is not conserved. In fact, free energy is more accurately a measure of entropy that can be produced at a given temperature (Lineweaver and Egan 2008). For processes of interest here, entropy production will be equated to chemical or radiative energy dissipation at the temperature of the surroundings, and all discussions refer to internal system entropy production (Meysman and Bruers 2007).

**2.2 Macrostates, Microstates and Ecosystems.** The MEP principle requires a system to have many degrees of freedom; that is, many different microstate configurations (technically, micropaths) that produce the same entropy producing macrostate (Dewar 2003). In this paper, we extend the concept of microstates to biological systems. Here, a microstate will refer to the organisms present and their connectivity in an ecosystem. Based on MEP principle, there should exist many different species configurations and trophic connections that give rise to the same entropy producing macrostate. Over appropriate timescales, species composition and their trophic relationships dramatically change, as has been observed in methanogenic (Fernandez *et al.* 1999), nitrifying (Graham *et al.* 2007) and planktonic communities (Beninca *et al.* 2008) that exhibit dynamics primarily at the species level but maintain similar biogeochemistry (i.e., functional stability). Indeed, multiplicity of biological microstates appears consistent with neutral theory (Pueyo *et al.* 2007), provided the different configurations are functionally complementary. Due to the large number of microbial species and their high abundances (Gans *et al.* 2005, Sogin *et al.* 2006), the microstate analogy is most applicable to microbial systems, which will be our primary focus.

**2.3 Biophysiochemical Constraints.** In any implementation of MEP principle, system constraints are critical in determining the relevant solution and the rate of entropy production at MEP (Crooks 2007). As an example, consider a burning mixture of methane and air. If no energy is stored, then all the free energy of combustion is ultimately dissipated as entropy at the temperature of the surroundings. In this case, the process is operating at the MEP state, where the magnitude of entropy production is constrained by the kinetic theory for gasses, which we could calculate *a priori*. We consider this type of abiotic process operating in a steepest descent mode, where the rate of entropy production is maximized at any instance in time and space, subject to kinetic constraints.

Outside the flammability limits, CH<sub>4</sub> and O<sub>2</sub> still react in a steepest descent mode, but the reaction proceeds extremely slowly. The reaction rate can be accelerated considerably by the addition of a catalyst, but for self organizing systems, the catalyst must be created from CH<sub>4</sub> and O<sub>2</sub> and/or materials contained within the system. The reaction rate, and entropy produced, depends on the effectiveness and quantity of the catalyst. An insufficient amount of catalyst limits the reaction rate, but too much catalyst wastes resources. If the catalyst contains internal energy, which it is likely to, then over-synthesis of catalyst is not consistent with MEP, because entropy could have been produced instead of catalyst. The catalysts effectiveness (i.e., reaction rate per unit mass) depends on its molecular structure, which in turn depends on the resources available to construct it. Given the available elements, what is the most effective catalyst that can be constructed? Obviously, this is an extremely difficult question to answer given the tremendous number of degrees of freedom. We know the upper bound is the complete removal of the reaction's activation energy, but how much the activation energy can be decreased is constrained by the elemental resources used to build the catalyst. Unlike the relatively simple

kinetics of combustion, we cannot determine *a priori* what the maximum reaction rate is nor what the maximum entropy production rate could be. Experiments are necessary.

**2.4 MEP from Individuals or Ecosystems?** Of course, the catalysts of interest here are enzymes, but we include the organisms that synthesize enzymes as part of the catalyst.

Continuing our example, we know from experiments that methanotrophs catalyze methane oxidation as follows,



where methanotrophs, M, both catalyze the reaction and are produced by it if their growth efficiency,  $\varepsilon_{\text{M}}$ , is  $> 0$ . The question we address here is how might evolution by natural selection lead to a system that maximizes entropy production? The competitive exclusion principle (Armstrong and McGehee 1980) tells us that the organism with the fastest growth will dominate at the exclusion of all others. To achieve fast growth an organism must balance efficiency versus speed. In (1) above, if  $\varepsilon_{\text{M}}$  is close to unity (high efficiency), the reaction will proceed slowly due to thermodynamic constraints (Gnaiger 1990, Pfeiffer and Bonhoeffer 2002, but also see Section 3.1 below) and methanotroph productivity will be low. As  $\varepsilon_{\text{M}}$  approaches 0 (low efficiency), the reaction can proceed rapidly, but once again with low methanotroph productivity due to low efficiency. Between 0 and 1 there is an optimum  $\varepsilon_{\text{M}}$  that maximizes methanotroph productivity that is selected for by evolution (Ibarra *et al.* 2002), but this value does not maximize entropy production because the methane that contributed to methanotroph biomass could have been oxidized. We conclude that growth of individual organisms as selected for by evolution does *not* follow the MEP principle. But ecosystems are not composed of a single species, as grazers, G, are always present that consume prey, such as methanotrophs, as given by,





which is also an autocatalytic reaction that produces exponential growth. Of course, grazers of the methanotroph-grazers are often present, as well as detritivores. Extending this logic naturally leads to food webs observed in nature. A net result of all the predation and recycling is that the total biomass in the system is constantly turned over. It is possible to have all organisms growing near their maximum rates without any biomass accumulation, which can lead to an MEP state. Hence, it is necessary to have a closed food web of prey, predators and detritivores in order to achieve MEP. The predator-prey interactions also insure biological structures are continuously and dynamically reallocated to those reactions that can assure MEP under changing conditions.

**2.5 MEP, Information and Timescales.** Interestingly, the effectiveness of methanotrophs to catalyze (1) depends on information stored in their genome, which specifies how to construct astoundingly complex organic structures from resources available in the environment. Over evolutionary time, enzymes associated with methanotrophy would presumably improve (Adami 2002), so that less protein is needed by modern methanotrophs than ancient methanotrophs to attain the same reaction rate. Hence, the amount of entropy produced for a given amount of protein changes with evolution, as well as the introduction of new pathways all together. But information embedded in the metagenome makes it difficult to predict maximum entropy production in biological systems from first principles as can be done with physical systems and demonstrated by Paltridge (1975). For this paper we will rely on simple metabolic networks as constraints. Genomic information also permits biological systems to integrate entropy production over time and circumvent the steepest descent pathway of abiotic systems.

As discussed above, maximum entropy production rate for a flammable mixture of CH<sub>4</sub> and air is dictated by gas kinetics. While combustion is the MEP solution, it tends to destroy ordered structures, so has only short persistence. If a perturbation extinguishes the flame, the CH<sub>4</sub> and air mixture will accumulate until a serendipitous spark is reintroduced. Consequently, there can be periods of massive entropy production followed by long periods of no entropy production, and combustion never occurs if the mixture falls outside its flammability limits. If a catalyst is introduced, such as methanotrophs, then entropy can be continuously produced over substantial transients. Even though the instantaneous entropy production will be lower with methanotrophs, the average rate of entropy production can exceed that of the sporadic steepest descent route.

If MEP follows only steepest descent pathways, then not only should CH<sub>4</sub> be oxidized, but all biomass as well, as this would produce the greatest instantaneous entropy production. However, if time-averaged entropy production is maximized (Moroz 2008), then allocating some CH<sub>4</sub> to methanotrophs and grazers increases entropy production over the integration interval. Unlike physical systems, biological systems have the capability to predict the future, where the information to do so is contained within the system metagenome. For instance, deciduous forests store some resources during the growing season that allows them to maintain dormancy over the winter or dry period, which requires expectations of future conditions. Microbes also exhibit temporal strategies such as spore formation and luxury uptake mechanisms (Berman-Frank and Dubinsky 1999, Khoshmanesh *et al.* 2002). When considering biological systems, strategies are not instantaneous, but are integrated over time based on a prediction of the future that has been selected for by evolution. By avoiding the steepest descent pathway, information can allow a system to produce entropy even when confronted with perturbations. Of course, perturbations of sufficient magnitude will disrupt biological systems as well. Nevertheless, we postulate *the*

*difference between abiotic and biotic processes is that the former always follows a pathway of steepest descent, while the later follows a pathway dictated by information that leads to greater entropy production when averaged over time.* Of course, the two pathways are always competing, such as occurs when fire consumes a forest. Pathways of averaged entropy production may be flanked by pathways of steepest descent.

**2.6 Distributed Metabolic Networks** Because of the multitude and redundancy of microstates, understanding flow of energy and mass through ecosystems at the organismal level is problematic (Graham *et al.* 2007, Beninca *et al.* 2008). Instead of focusing at the organismal level, we will pursue a functional representation using a metabolic network abstraction. When considering metabolic capabilities of microbial systems, we often find that metabolic function is distributed amongst all three domains of life: Bacteria, Archaea and Eukaryote. Since the species that comprise metabolic networks can undergo substantial substitution with only minor impact on functional characteristics (Fernandez *et al.* 2000, Wittebolle *et al.* 2008), we will view microbial systems as metabolic networks that can be distributed in space and time, but resemble multicellular organisms (Wilson and Sober 1989, Shapiro 1998).

### **3. Mathematical Framework**

As defined above, our basic model framework uses the metabolic network perspective that is applied at the ecosystem level. The formulation is similar to how single cells control metabolism by regulating the synthesis and degradation of enzymes associated with pathways necessary for growth. We assume that communities organize such that their combined metabolic expression maximizes entropy production. A methanotrophic microbial community in a batch reactor that is sparged with methane and air will serve as an example to demonstrate the approach.

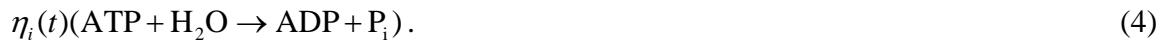
**3.1 Optimized Metabolic Ecosystem Network (OMEN) Model.** In our approach the microbial food web is replaced by a metabolic network that synthesizes generic biological structure,  $\mathcal{S}$ , which consists largely of enzymatic protein, but also represents other macromolecules expressed by microbial communities for growth and form. The biological structure can be allocated to any reaction in the network and serves as the reaction's catalyst. The metabolic network also orchestrates energy and mass acquisition necessary to construct biological structure itself, where the MEP principle governs how biological structure is allocated to any metabolic reaction. The mathematical framework is general and can be extended to any microbial system by expanding the distributed metabolic network based on knowledge from classic microbiology and metagenomics. One can also examine how the introduction or evolution of new metabolic functions alters resource allocation and system dynamics as well as examine dynamics when metabolic functions are removed.

For our aerobic methanotrophic metabolic network example (Fig. 1), eight half reactions account for methane oxidation ( $r_1, r_2, r_3$ ), including sugar biosynthesis ( $r_1$ ), nitrate reduction ( $r_4$ ) for biological structure synthesis from  $\text{NH}_4^+$  and sugars ( $r_5$ ), and biological structure and detritus degradation ( $r_6, r_7, r_8$ ) (Table S1 in Supplementary Materials). Each of the eight reactions has associated biological structure ( $\mathcal{S}_{1...7}$ ), except for reactions 7 and 8, which are both catalyzed by the same structure,  $\mathcal{S}_7$ , to minimize model degrees of freedom. The use of half reactions (Table S1) increases the flexibility of the network to utilize available electron acceptors or donors, but to insure electron conservation all half reactions are coupled to an electron shuttle reaction as follows,



where  $\varepsilon_i$  is the number of electron pairs produced by reaction  $i$ . Gibbs free energies are calculated for each reaction (Table S1) after coupling to (3). We use the approach of Alberty (2003, 2006) to calculate the standard Gibbs free energy of reaction, which accounts for proton dissociation equilibria between chemical species ( $\text{H}_2\text{CO}_3 \Leftrightarrow \text{H}^+ + \text{HCO}_3^-$ , etc.) at a specified pH and temperature. The overall free energy of reaction, denoted as  $\Delta G_r^\dagger(r_i)$ , accounts for the concentration of reactants and products ( $\mathbf{c}(t)$ ), where ionic strength,  $I_S$ , is used to estimate activity coefficients (Alberty 2003). For the methanotrophic network simulation, we use the following conditions: pH 7,  $I_S$  1.92  $\mu\text{M}$ , at 20°C. Free energy, enthalpy and entropy of formation for biological structure is based on Battley (1999, 2003) for bacteria; however, this is not critical as the free energy of living organisms is similar in value to the substrates they are constructed from.

Of course,  $\Delta G_r^\dagger(r_i)$  represents the available or needed energy only if the reaction is run reversibly, which is clearly not the case for biological systems. To account for inefficient energy transfer or energy production, all reactions are coupled to an energy source (or sink) reaction in the form of ATP hydrolysis (or synthesis) of the form,



Even though we know most ATP reaction couplings from biochemistry, cells have the ability to dissipate ATP (Russell and Cook 1995), and different organisms in a community can have different ATP couplings for the same pathways (Helling 2002). Consequently, we treat energy coupling,  $\eta_i(t)$ , as a control variable to be determined by optimization. The combined whole

reaction  $i$  consists of (3) plus (4) and half reaction  $i$  (Fig. 1, Table S1). The Gibbs free energy of the combined reaction  $i$ ,  $\Delta G_c^\dagger(r_i)$ , is then given by

$$\Delta G_c^\dagger(r_i) = \Delta G_r^\dagger(r_i) + \eta_i(t)\Delta G_r^\dagger(r_{ATP}), \quad (5)$$

where  $\Delta G_r^\dagger(r_{ATP})$  is the Gibbs free energy of (4). By altering the magnitude and sign of  $\eta_i(t)$ , an endergonic reaction can be driven forward, and by inefficient coupling of (4) with exergonic reactions, energy can be extracted at less than 100% efficiency, allowing the reaction to proceed at higher rates. This is the standard tradeoff between power and efficiency; operating reactions at high throughput necessitates low efficiency, while attaining high efficiency limits reaction rate, which all biotic and abiotic system must contend with (Gnaiger 1990, Pfeiffer and Bonhoeffer 2002).

Metabolic network reaction rates are given by,

$$r_i = v_i \mathfrak{S}_i(t) F_i^K(\mathbf{c}) F_i^T(\mathbf{c}), \quad (6)$$

where  $v_i$  is an experimentally determined rate constant per unit of biological structure allocated,  $\mathfrak{S}_i(t)$  is biological structure allocated to reaction  $i$ , and  $F_i^K(\mathbf{c})$  and  $F_i^T(\mathbf{c})$  are kinetic and thermodynamic forces, respectively. The kinetic force is given by the general form,

$$F_i^K(\mathbf{c}) = f_{NADH}(\zeta_{NADH}, \varepsilon_i) f_{ATP}(\zeta_{ATP}, \eta_i) \prod_j \left( \frac{c_j}{c_j + K_{i,j}} \right)^{\Phi_{i,j}}, \quad (7)$$

where  $c_j$  is the concentration of substrate  $j$ , and the matrix element,  $\Phi_{i,j}$ , equals 1 if reaction  $i$  consumes substrate  $j$ ; otherwise  $\Phi_{i,j}$  is zero (Table S1). To ensure electron and energy conservation, we assume biological structure has a fixed amount of NAD+NADH and

ADP+ATP storage per unit of total biological structure,  $\mathfrak{S}_T = \sum_i \mathfrak{S}_i$ , as specified by the constants  $\zeta_{NADH}^T$  and  $\zeta_{ATP}^T$ , respectively ( $\mu\text{mol} (\text{mmol } \mathfrak{S}_T)^{-1}$ ). Consequently, the amounts of NAD and NADH as well as ATP and ADP per biological structure are constrained by,

$$\zeta_{NADH}^T = \zeta_{NADH}(t) + \zeta_{NAD}(t) \text{ and } \zeta_{ATP}^T = \zeta_{ATP}(t) + \zeta_{ADP}(t). \quad (8)$$

We define the functions  $f_{NADH}$  and  $f_{ATP}$  based on  $\zeta_{NADH}^T$  and  $\zeta_{ATP}^T$  as,

$$f_{NADH}(\zeta_{NADH}, \varepsilon_i) = \begin{cases} \zeta_{NADH} / (\zeta_{NADH} + K_{NADH}) & \varepsilon_i < 0 \\ \zeta_{NAD} / (\zeta_{NAD} + K_{NADH}) & \varepsilon_i > 0 \\ 1 & \varepsilon_i = 0 \end{cases} \quad (9)$$

$$f_{ATP}(\zeta_{ATP}, \eta_i) = \begin{cases} \zeta_{ATP} / (\zeta_{ATP} + K_{ATP}) & \eta_i > 0 \\ \zeta_{ADP} / (\zeta_{ADP} + K_{ATP}) & \eta_i < 0 \\ 1 & \eta_i = 0 \end{cases} \quad (10)$$

Functions (9) and (10) account for the redox and energy state of the system, analogous to cellular metabolism.

Chemical reaction rates are often limited by kinetics, so the thermodynamic force is often ignored because it is usually close to unity. However, as the reaction approaches equilibrium,  $F_i^T(\mathbf{c})$  in (6) approaches zero and constrains the net reaction rate no matter how favorable the reaction kinetics. It can be shown (Boudart 1976, Jin and Bethke 2003) that the thermodynamic force is related to the Gibbs free energy of the reaction,  $\Delta G_c^\ddagger(r_i)$ , as follows,

$$F_i^T(\mathbf{c}) = 1 - \exp\left(\frac{\Delta G_c^\ddagger(r_i)}{RT\chi_i}\right) \text{ for } \Delta G_c^\ddagger(r_i) \leq 0, \quad (11)$$

where  $R$  is the gas constant,  $T$  is temperature (°K) and  $\chi_i$  is the average stoichiometric number for net reaction  $i$  (Boudart 1976, Vuddagiri *et al.* 2000). By adjusting  $\eta_i(t)$ ,  $F_i^T(\mathbf{c})$  can be driven from 0 to 1 via relationship to (5). Because of the complexity of community metabolic networks,  $\chi_i$  is treated as a tunable parameter for each reaction.

Reactions  $r_5(t)$  and  $r_6(t)$  represent the synthesis and degradation of all biological structures, respectively. However, which of the seven biological structures to produce or degrade at time  $t$  has not been specified. Consequently, we introduce an additional set of control variables,  $\sigma_i(t)$ , that specify the partitioning of biological structure synthesis, which is analogous to transcription plus translation (Fig. 2). While we could introduce another set of control variables to determine which biological structures to degrade, we make the assumption that degradation of biological structure is nonspecific and only depends on the relative concentration of  $\mathfrak{S}_i$  to total biological structure,  $\mathfrak{S}_7$ , and the concentration of  $\mathfrak{S}_6$ . In addition, biological structure is not perfectly degraded to building block materials (i.e.,  $\text{CH}_2\text{O}$  and  $\text{NH}_4^+$ ), but produces some detrital carbon (dC) and nitrogen (dN) (Fig. 1), as specified by the assimilation parameters  $f_{aC}$  and  $f_{aN}$ , respectively (Table S1). Breakdown of detrital C and N is controlled by reactions  $r_7(t)$  and  $r_8(t)$ , but only one biological structure,  $\mathfrak{S}_7$ , catalyzes both reactions.

Based on the metabolic network and reaction rates (Fig. 1), a mass balance model can be constructed for the state variables,  $\mathbf{x}^T(t) = [\mathbf{c}(t), \mathfrak{S}(t), \boldsymbol{\zeta}(t)]^T$ , which represent the concentrations of chemical species,  $\mathbf{c}(t)$ , biological structures,  $\mathfrak{S}(t)$ , as well as the system's redox and energy states ( $\zeta_{NADH}$ ,  $\zeta_{ATP}$ ), and has the general form,



$$\frac{d\mathbf{x}(t)}{dt} = \mathbf{f}(\mathbf{x}(t); \boldsymbol{\eta}(t), \boldsymbol{\sigma}(t)) \quad (12)$$

Once the control variables  $\boldsymbol{\sigma}(t)$  and  $\boldsymbol{\eta}(t)$  are specified over time, the state equation (12) can be solved for the state variables (see Eqs. S1-S14 in the supplemental material for the expansion of Eq. 12 in scalar form).

The control variables,  $[\boldsymbol{\eta}(t), \boldsymbol{\sigma}(t)]^T$ , are determined by formulating and solving an interval optimization problem in which average entropy production rate,  $\left\langle \frac{dS}{dt} \right\rangle_{t_j}$ , is maximized over a specified interval of time,  $\delta_t$  for each  $t_j$  interval. Entropy production is given by the negative of a reaction rate times the Gibbs free energy of the reaction divided by temperature, summed over all reactions (Eu 1992 pp. 131-141), which is readily calculated. The general form of the model is as follows:

$$\begin{aligned} \max \quad & \left\langle \frac{dS}{dt} \right\rangle_{t_j} = \frac{-1}{T\delta_t} \int_{t_j}^{t_j+\delta_t} \sum_i r_i(\tau) \Delta G_c^\dagger(r_i(\tau)) d\tau & \text{Subject to :} \\ \text{wrt } & \boldsymbol{\eta}(t), \boldsymbol{\sigma}(t) & \boldsymbol{\eta}_L \leq \boldsymbol{\eta}(t) \leq \boldsymbol{\eta}_U \\ & & 0 \leq \sigma_i(t) \leq 1 \\ & & \sum_i \sigma_i(t) = 1 \end{aligned} \quad (13)$$

which is a class of optimal control problems (Kirk 1970); however, since no constraints are imposed on the state variables,  $\mathbf{x}(t)$ , it is not necessary to include the differential equations (12) as part of the constraints, as they are explicitly satisfied. To obtain a solution over a specified time domain,  $[t_0, t_f]$ , the optimal control problem (13) is solved repetitively over a sufficient number of  $j$  intervals,  $[t_j, t_j + \delta_t]$ , to cover the entire domain. There are several ways of solving

(13), including linear programming following linearization at time,  $t$  (Vallino *et al.* 1996, Vallino 2003). However, we have implemented an interval optimization method using SNOPT (Gill *et al.* 2005) that solves the nonlinear programming problem over the interval,  $\delta_t$  via sequential quadratic programming coupled with block implicit methods to solve the associated differential equations (12) (Brugnano and Magherini 2004). In the current implementation the optimal interval,  $\delta_t$ , is subdivided into  $n_g$  grid points, and the control functions  $[\boldsymbol{\eta}(t), \boldsymbol{\sigma}(t)]^T$  are discretized over the interval as linear piecewise continuous functions. The length of the optimization interval,  $\delta_t$ , is an interesting aspect of the model, as it reflects the characteristic time scale of environmental variability that a living system has evolved to cope with over a given spatial scale (O'Neill *et al.* 1986). We expect that microbial systems can be described by short optimization intervals ( $\sim$ days), while for forest ecosystems  $\delta_t$  would require longer intervals ( $\sim$ year). It is also possible to cast the problem as a type of infinite horizon (Carlson *et al.* 1991) or receding horizon (Ito and Kunisch 2002) optimal control problem, but we have yet to fully investigate that possibility.

## 4. Simulation Results

Figures 3-5 show results from a standard simulation based on the above equations for a 18 L methanotrophic microbial community that is sparged with 2.9% methane in air at 20 mL min<sup>-1</sup> (STP) starting with initial nutrient concentrations of 700  $\mu$ M NO<sub>3</sub><sup>-</sup>, 70  $\mu$ M PO<sub>4</sub><sup>3-</sup>, plus salts and trace elements. The simulation conditions are based on current methanotrophic microcosm experiments that are being used to guide model development and test the MEP principle; however, the model has not been calibrated to the observations yet, but we do provide some preliminary experimental data in the Supplementary Materials for qualitative comparison. For

these simulations, an eight day optimization interval,  $\delta_t$ , was employed, which results in  $\text{NO}_3^-$  depletion by day 75 (Fig. 3a), which closely matches observations (Fig. S2a). While not evident on the linear scale (Fig. 3), constant exponential growth begins at time zero but does not result in significant changes in state variables until day 40, which is similarly observed experimentally (Fig. S2). Because entropy production is maximized over an 8 day interval instead of instantaneously, entropy production can be increase if some  $\text{CH}_4$  is allocated to  $\mathcal{S}$  rather than being completely oxidized to  $\text{CO}_2$ . During the first 75 days, biological structure is allocated to respiration ( $\mathcal{S}_3$ ),  $\text{CH}_4$  oxidation ( $\mathcal{S}_1$ ),  $\text{CH}_2\text{O}$  oxidation ( $\mathcal{S}_2$ ), biological structure synthesis ( $\mathcal{S}_5$ ) and nitrate reduction ( $\mathcal{S}_4$ ) in decreasing magnitude (Fig. 3b). The majority of  $\text{NO}_3^-$  is converted to  $\text{NH}_4^+$  for biological structure synthesis; however, some  $\text{NH}_4^+$  accumulates, but is later consumed after  $\text{NO}_3^-$  depletion (Fig. 3a). Ammonium accumulation during the first 75 days occurs to balance excess electrons resulting from  $\text{CH}_4$  oxidation that is not completely balanced by  $\text{O}_2$  reduction to  $\text{H}_2\text{O}$ . Ammonium accumulation and depletion also occurs in the experimental microcosms (Fig. S2a), both before and after  $\text{NO}_3^-$  depletion. Entropy production rate increases rapidly during the first 75 d, while both the “intracellular” redox ( $\zeta_{\text{NADH}}$ ) and energy ( $\zeta_{\text{ATP}}$ ) states oscillate around their balance state of  $0.5 \mu\text{mol mmol}^{-1}$  (Fig. 4). Because “intracellular”  $\zeta_{\text{NADH}}$  and  $\zeta_{\text{ATP}}$  have faster characteristic timescales than other state variables (see Eqns S10 and S11), balancing internal NADH/NAD and ATP/ADP ratios severely constrains reaction rates and may account for energy spilling observed in bacterial cultures (Dauner *et al.* 2001).

Near the 75 d transition point, control variables change so that  $\sigma_4$  decreases to zero while  $\sigma_6$  and  $\sigma_7$  turn on for the first time (Fig. 5a). The latter two control variables allocate biological structure to decomposition of biological structure and detritus, respectively, while  $\sigma_4$  controls

$\text{NO}_3^-$  uptake, which is exhausted after 75 d, so is no longer needed. Control variables also alter reaction energy coupling via changes in  $\eta_i$  around the transition point (Fig. 5b). Prior to  $\text{NO}_3^-$  depletion, the system runs at high efficiency, in that ATP synthesis is high ( $\eta_3$  more negative), while ATP requirements for biological structure synthesis,  $r_5$ , are maintained low ( $\eta_5$  less positive) (Fig. 5b). After  $\text{NO}_3^-$  depletion, control variables switch to favor low ATP synthesis ( $\eta_3$  less negative) and high ATP requirements ( $\eta_5$  more positive); a low efficiency mode, which is necessary to maintain energy balance as dictated by  $\zeta_{ATP}$  via (10) (Fig. 4). Laboratory cultures have also been shown to spill ATP under nutrient limited growth (Russell and Cook 1995), so also exhibit similar low efficiency modes.

In the latter part of the simulation, detrital C and N (dC, dN) begin to accumulate due to synthesis of  $\mathfrak{S}_6$ , which degrades biological structure with dC and dN formation (Fig. 3a).

Biological structure is ultimately limited by N availability, so that more entropy can be produced if N is maintained within the biological structure pool,  $\mathfrak{S}$ . Consequently, control variables allocate some biological structure to  $\mathfrak{S}_7$  to return N in dN to active catalyst. While  $\mathfrak{S}_7$  also causes conversion of dC to  $\text{CH}_2\text{O}$ , entropy production is not C limited due to the continuous addition of  $\text{CH}_4$ . While complete oxidation of dC would contribute to entropy production, entropy production per unit of biological structure is higher for  $\text{CH}_4$  oxidation. Consequently, biological structure is not squandered to completely utilize dC under the simulated conditions. As a result, dC continuously accumulates (Fig. 3a). Interestingly, we also have observed long-term accumulation and depletion of dissolved organic C and N in the experimental microcosms (Fig. S2b) and large amounts of particulate C has visually accumulated in the microcosms (Fig. S1), but it has not been quantified yet. Synthesis of  $\mathfrak{S}_7$  continues until decomposition of dN matches

its production, at which point  $\sigma_7$  is down regulated and the system reaches pseudo steady state at ~300 d (Fig. 3).

During the period in which N is bound in dN ( $75 < t < 300$ ), entropy production is low, but increases to its steady state value once recycling of N from dN is balanced (Fig. 4). The system reaches pseudo-steady state near day 300, where entropy production rate is  $2.16 \text{ J L}^{-1} \text{ d}^{-1} \text{ }^\circ\text{K}^{-1}$  and the dominate reaction rates are respiration ( $r_3$ :  $1570 \text{ } \mu\text{M d}^{-1}$ ), methane oxidation ( $r_1$ :  $815 \text{ } \mu\text{M d}^{-1}$ ) and  $\text{CH}_2\text{O}$  oxidation ( $r_2$ :  $755 \text{ } \mu\text{M d}^{-1}$ ). Production of biological structure,  $r_5$ , matches its degradation rate,  $r_6$ , at  $120 \text{ } \mu\text{M d}^{-1}$ . The high frequency 8 d oscillations observed throughout the simulation (Figs. 3-5) obviously result from the 8 d optimization interval used. However, if the sub-interval daily data points are not plotted, then the oscillations disappear. The oscillations are analogous to changes in plant metabolism that occur over the course of the daily photic period. Interestingly,  $\text{CH}_2\text{O}$  acts as an “internal” storage mechanism over the sub-interval (Fig. 3a). The magnitude of  $\text{CH}_2\text{O}$  storage increases with longer optimization intervals, or if  $\text{CH}_4$  is periodically introduced into the system instead of continuously (data not shown). While no attempt has been made to calibrate the model to experimental data yet, maximizing entropy production over successive intervals does produce results at least qualitatively similar to our observations (see Supplementary Materials).

## 5. Discussion

The maximum entropy production principle states that systems will tend to follow pathways that maximize entropy production at steady state. If internal self organized structures facilitate entropy production, then those structures are more likely to develop and persist, such as occurs with Rayleigh-Benard convection cells and hurricanes (Schneider and Kay 1994). In the MEP

perspective, living systems can be viewed as molecular machines that catalyze reactions that both synthesize and degrade molecular machines and dissipate energy via redox reactions in the process, but are otherwise undifferentiated from abiotic systems. Because MEP states that there must be many different ways to produce entropy maximally, the form of the molecular machines is unimportant for biogeochemical MEP. Consequently, we have chosen a metabolic perspective of ecosystem biogeochemistry that only predicts functional characteristics in terms of allocation of molecular machinery. The approach strives for long-term predictability at the expense of short-term dynamic details. However, the metabolic model does predict biological structure allocation (Fig. 3b), which can be related to observed functional groups and community level gene expression obtained from experiments, and it does produce reasonable results and qualitatively compares with our experimental mesocosm data prior to calibration. Furthermore, the metabolic MEP perspective leads to some interesting extrapolations, which we briefly explore below.

Current MEP theory applies to steady state systems (Dewar 2003, Niven 2009), but ecosystems seldom operate at steady state and at times can be quite far from it. During major transient events, such as recovery from system collapse, MEP may not be a useful descriptor of system response. Rather, the community composition will likely play the dominant role in system dynamics as it reorganizes and attempts to return to MEP. However, for perturbations that an ecosystem has evolved to cope with, MEP should be an important system attractor, so we expect MEP to be relevant even if a system is not strictly at steady state. In fact, it is the ability of living systems to predict future states in the presence of fluctuations that allows biological systems to produce more entropy than abiotic systems that take the steepest descent approach. To predict future states requires a system to store relevant information. Hence, increases in

information (i.e., physical complexity (Adami *et al.* 2000)) facilitate increases in thermodynamic entropy production—a rather interesting parallel.

Biological systems acquire information through evolution and store it within the metagenome. This information describes how to construct biological structures from the resources available. But to maximize entropy production requires optimal use, and possibly selection, of resources at the whole system level (Traulsen and Nowak 2006, Williams and Lenton 2008, Wilson and Wilson 2008), not at the level of the constituent components. Information stored in any genome correlates with the entire system (Adami 2002), including that stored in other genomes, so all ecosystem organisms must be inherently coupled to achieve MEP. Consider a closed ecosystem similar to our methanotrophic system, which contains finite resources and so a limit to the total amount of biological structure that can be synthesized. How should biological structures be allocated to maximize entropy production? While some biological structure must be allocated to oxidizing methane, other structures must be allocated to recycling nutrients, including nutrients contained in biological structures that are no longer useful (Chesson and Kuang 2008). Of course, excessive or insufficient allocation to any one structure will result in sub optimal entropy production, so system level coordination (e.g., Fig. 3b) is an emergent property of the MEP principle.

While one can image a single organism that could orchestrate all ecosystem functions, it would be at a competitive disadvantage to any specialist organism that only maintained a fraction of the entire metagenome (Mills *et al.* 1967). Distributing genomic information allows a system to retain important information not relevant to the current ecosystem state at very low copy number, such as observed in the rare biosphere (Sogin *et al.* 2006). Accordingly, a distributed metabolic system can function as a single metabolic entity, but is more efficient with system resources

hence is more likely to be selected for at the system level (Traulsen and Nowak 2006, Williams and Lenton 2008, Wilson and Wilson 2008). Consequently, we view an ecosystem composed of autonomous individuals all attempting to maximize their own fecundity in the Darwinian context, but this leads to an MEP state at the system level. If MEP is a true attractor, then ecosystems that recycle nutrients more effectively should become more spatially dominate. Likewise, systems that produce more entropy over longer time periods should also be selected for. We speculate that *systems will evolve towards entropy production that is maximized over infinite time and space*. This contrasts abiotic (information lacking) systems that maximize entropy production at a point in space-time (steepest descent).

If a system organizes toward maximum entropy production over infinite time and space, then steepest descent routes must be inhibited, but they cannot be prevented from occurring. As already mentioned, a forest fire rapidly increases short term entropy production, but at the expense of long term entropy production. Invasive species can produce identical phenomenon if the invading species propagates via oxidation of biological structure. However, if the invading species causes lower entropy production on longer timescales, then the new state will not be stable. On the contrary, if the invasive species increases averaged entropy production, then it should persist. Entropy production serves as a useful measure of system stability then. While changes in species abundances are often associated with instability, if the change does not alter entropy production, then from the MEP perspective, the system is stable. Changes in biodiversity that lead to lower entropy production would be considered detrimental to system stability.

If a system is subject to external perturbations, then we may expect the system to store energy internally so that entropy production can be maintain unperturbed, such as observed in CH<sub>2</sub>O



dynamics in our modeled system (Fig. 3a). One way to store energy is to increase the fraction of large biological structures. From the metabolic theory of ecology (MTE), organism respiration increases only as the  $3/4$  power of its mass (West *et al.* 1997). Consequently, a unit mass of bacteria has a much higher specific respiration than an equivalent mass of elephant or whale. While our model above can store “internal” energy as  $\text{CH}_2\text{O}$ , that cannot occur in the environment unless it is protected from microbial consumption, such as in the form of a large animal. If a perturbation occurs, large animals may perish, but they provide energy to prevent hierarchical collapse of the entire system. This is consistent with the observation that biomass specific respiration decreases with increase ecosystem maturity (Odum 1969).

While we have discussed some of the possible implications of the MEP principle in this section, our primary objective of this manuscript has been to demonstrate how MEP can be used quantitatively as a governing principle to understand and model biogeochemical processes. A key aspect of our approach is to replace the organismal focus with a functional one, which should produce more robust predictions, albeit with some loss of details. To further develop this approach, it is necessary to examine ecosystems purely at the functional level. For instance, many higher heterotrophs merely sever to capture and masticate biological structure, while other organisms provide physical structure, such as for trapping sediments on marshes (Mudd *et al.* 2004), facilitating evapotranspiration (Wang *et al.* 2007), or other transport limited phenomena (Meysman *et al.* 2006). We will need to understand the functions that organisms contribute to ecosystems and to quantify functional rates per unit amount of biological structure allocated. Even though examining ecosystems from an MEP perspective will not always be productive for every interest, it does provide an alternative vantage point that can provide great insight. Based

on the MEP principle, living systems exist because they hasten entropy production over greater spatiotemporal scales than abiotic systems.

**Acknowledgements:** I thank Axel Kleidon for organizing the workshop on MEP in the Earth System, as well as editing this special issue of the *Philosophical Transactions of the Royal Society B* along with Yadvinder Malhi. The work presented here was funded by the PIE-LTER program (NSF OCE-0423565), as well as from NSF CBET-0756562, NSF EF-0928742 and NASA Exobiology and Evolutionary Biology (NNG05GN61G).

## 6. References

- Adami,C. (2002) Sequence complexity in Darwinian evolution. *Complexity* **8**, (2) 49-56, doi: 10.1002/cplx.10071.
- Adami,C., Ofria,C., and Collier,T.C. (2000) Evolution of biological complexity. *Proceedings of the National Academy of Sciences of the United States of America* **97**, (9) 4463-4468.
- Alberty,R.A. (2003) *Thermodynamics of biochemical reactions*, Hoboken, NJ, Wiley & Sons, 397 pp.
- Alberty,R.A. (2006) *Biochemical thermodynamics: Applications of Mathematica*, Hoboken, NJ, Wiley & Sons, 464 pp.
- Armstrong,R.A. and McGehee,R. (1980) Competitive Exclusion. *Amer.Nat.* **115**, (2) 151-170.
- Battley,E. (2003) Absorbed heat and heat of formation of dried microbial biomass: Studies on the thermodynamics of microbial growth. *Journal of Thermal Analysis and Calorimetry* **74**, (3) 709-721, doi: 10.1023/B:JTAN.0000011003.43875.0d.
- Battley,E.H. (1999) An empirical method for estimating the entropy of formation and the absolute entropy of dried microbial biomass for use in studies on the thermodynamics of microbial growth. *Thermochimica Acta* **326**, (1-2) 7-15.
- Bejan,A. (2007) Constructal theory of pattern formation. *Hydrology and Earth System Sciences* **11**, (2) 753-768.
- Beninca,E., Huisman,J., Heerkloss,R., Johnk,K.D., Branco,P., Van Nes,E.H., Scheffer,M., and Ellner,S.P. (2008) Chaos in a long-term experiment with a plankton community. *Nature* **451**, (7180) 822-825, doi: 10.1038/nature06512.
- Berman-Frank,I. and Dubinsky,Z. (1999) Balanced growth in aquatic plants: Myth or reality? *BioScience* **49**, (1) 29-37.
- Boudart,M. (1976) Consistency between kinetics and thermodynamics. *J.Phys.Chem.* **80**, (26) 2869-2870.
- Brugnano,L. and Magherini,C. (2004) The BiM code for the numerical solution of ODEs. *Journal of Computational and Applied Mathematics* **164-165**, 145-158, doi: 10.1016/j.cam.2003.09.004.
- Carlson,D.A., Haurie,A.B., and Leizarowitz,A. (1991) *Infinite horizon optimal control: Deterministic and stochastic systems*, 2 Ed., Berlin, Springer-Verlag, 332 pp.
- Chesson,P. and Kuang,J.J. (2008) The interaction between predation and competition. *Nature* **456**, (7219) 235-238, doi: 10.1038/nature07248.

- Choi,J.S., Mazumder,A., and Hansell,R.I.C. (1999) Measuring perturbation in a complicated, thermodynamic world. *Ecol.Modelling* **117**, (1) 143-158, doi: 10.1016/S0304-3800(99)00042-3.
- Clausius,R. (1850) Ueber die bewegende Kraft der Wärme und die Gesetze, welche sich daraus für die Wärmelehre selbst ableiten lassen. *Annalen der Physik und Chemie* **155**, (3) 368-397, doi: 10.1002/andp.18501550306.
- Crooks,G.E. (2007) Beyond Boltzmann-Gibbs statistics: Maximum entropy hyperensembles out of equilibrium. *Physical Review E (Statistical, Nonlinear, and Soft Matter Physics)* **75**, (4 (041119)) 1-5, doi: 10.1103/PhysRevE.75.041119.
- Dauner,M., Storni,T., and Sauer,U. (2001) Bacillus subtilis Metabolism and Energetics in Carbon-Limited and Excess-Carbon Chemostat Culture. *J.Bacteriol.* **183**, (24) 7308-7317, doi: 10.1128/JB.183.24.7308-7317.2001.
- Dewar,R. (2003) Information theory explanation of the fluctuation theorem, maximum entropy production and self-organized criticality in non-equilibrium stationary states. *Journal of Physics A: Mathematical and General* **36**, 631-641.
- Dewar,R.C. (2005) Maximum entropy production and the fluctuation theorem. *Journal of Physics A: Mathematical and General* **38**, (21) L371-L381, doi: 10.1088/0305-4470/38/21/L01.
- Eu,B.C. (1992) *Kinetic theory and irreversible thermodynamics*, Montreal, John Wiley & Sons Canada, Ltd., 752 pp.
- Falkowski,P.G., Fenchel,T., and DeLong,E.F. (2008) The Microbial Engines That Drive Earth's Biogeochemical Cycles. *Science* **320**, (5879) 1034-1039, doi: 10.1126/science.1153213.
- Fath,B.D., Patten,B.C., and Choi,J.S. (2001) Complementarity of Ecological Goal Functions. *Journal of Theoretical Biology* **208**, (4) 493-506.
- Fernandez,A.S., Hashsham,S.A., Dollhopf,S.L., Raskin,L., Glagoleva,O., Dazzo,F.B., Hickey,R.F., Criddle,C.S., and Tiedje,J.M. (2000) Flexible community structure correlates with stable community function in methanogenic bioreactor communities perturbed by glucose. *Appl.Microbiol.Biotechnol.* **66**, (9) 4058-4067.
- Fernandez,A., Huang,S., Seston,S., Xing,J., Hickey,R., Criddle,C., and Tiedje,J. (1999) How Stable Is Stable? Function versus Community Composition. *Applied and Environmental Microbiology* **65**, (8) 3697-3704.
- Gans,J., Wolinsky,M., and Dunbar,J. (2005) Computational Improvements Reveal Great Bacterial Diversity and High Metal Toxicity in Soil. *Science* **309**, (5739) 1387-1390.
- Gill,P.E., Murray,W., and Saunders,M.A. (2005) SNOPT: An SQP Algorithm for Large-Scale Constrained Optimization. *SIAM Review* **47**, (1) 99-131.

- Gnaiger,E. (1990) Concepts on efficiency in biological calorimetry and metabolic flux control. *Thermochimica Acta* **172**, 31-52.
- Graham,D.W., Knapp,C.W., Van Vleck,E.S., Bloor,K., Lane,T.B., and Graham,C.E. (2007) Experimental demonstration of chaotic instability in biological nitrification. *ISME J* **1**, (5) 385-393, doi: 10.1038/ismej.2007.45.
- Helling,R.B. (2002) Speed versus Efficiency in Microbial Growth and the Role of Parallel Pathways. *J.Bacteriol.* **184**, (4) 1041-1045, doi: 10.1128/jb.184.4.1041-1045.2002.
- Ibarra,R.U., Edwards,J.S., and Palsson,B.O. (2002) Escherichia coli K-12 undergoes adaptive evolution to achieve in silico predicted optimal growth. *Nature* **420**, (6912) 186-189, doi: 10.1038/nature01149.
- Ito,K. and Kunisch,K. (2002) Receding horizon optimal control for infinite dimensional systems. *ESAIM: Control, Optimisation and Calculus of Variations* **8**, 741-760, doi: 10.1051/cocv:2002032.
- Jin,Q. and Bethke,C.M. (2003) A New Rate Law Describing Microbial Respiration. *Applied and Environmental Microbiology* **69**, (4) 2340-2348.
- Jorgensen,S.E. (1994) Review and comparison of goal functions in system ecology. *Vie Milieu* **44**, (1) 11-20.
- Jorgensen,S.E., Patten,B.C., and Straskraba,M. (2000) Ecosystems emerging: 4. growth. *Ecol.Modelling* **126**, (2-3) 249-284.
- Khoshmanesh,A., Hart,B.T., Duncan,A., and Beckett,R. (2002) Luxury uptake of phosphorus by sediment bacteria. *Water Res.* **36**, (3) 774-778, doi: 10.1016/S0043-1354(01)00272-X.
- Kirk,D.E. (1970) *Optimal control theory: an introduction*, 1 Ed., Englewood Cliffs, Prentice-Hall, Inc., 452 pp.
- Lineweaver,C.H. and Egan,C.A. (2008) Life, gravity and the second law of thermodynamics. *Physics of Life Reviews* **5**, (4) 225-242, doi: 10.1016/j.plrev.2008.08.002.
- Lorenz,R.D., Lunine,J.I., and Withers,P.G. (2001) Titan, Mars and Earth : Entropy production by latitudinal heat transport. *Geophys.Res.Lett.* **28**, (3) 415-418.
- Lotka,A.J. (1922) Contribution to the Energetics of Evolution. *PNAS* **8**, (6) 147-151.
- Margalef,R. (1968) *Perspectives in ecological theory*, Chicago, University of Chicago Press, 112 pp.
- Mejer,H. and Jorgensen,S.E. (1979) Exergy and ecological buffer capacity. In: *State-of-the-art in ecological modelling*, 829-846. Edited by Jorgensen,S.E., London, Pergamon Press.

- Meysman,F.J.R. and Bruers,S. (2007) A thermodynamic perspective on food webs: Quantifying entropy production within detrital-based ecosystems. *Journal of Theoretical Biology* **249**, (1) 124-139, doi: 10.1016/j.jtbi.2007.07.015.
- Meysman,F.J.R., Galaktionov,O.S., Gribsholt,B., and Middelburg,J.J. (2006) Bioirrigation in permeable sediments: Advective pore-water transport induced by burrow ventilation. *Limnol.Oceanogr.* **51**, (1) 142-156.
- Mills,D.R., Peterson,R.L., and Spiegelman,S. (1967) An extracellular Darwinian experiment with a self-duplicating nucleic acid molecule. *Proceedings of the National Academy of Sciences of the United States of America* **58**, (1) 217-224.
- Morowitz,H.J. (1968) *Energy flow in biology: biological organization as a problem in thermal physics*, New York, Academic Press, 179 pp.
- Moroz,A. (2008) On a variational formulation of the maximum energy dissipation principle for non-equilibrium chemical thermodynamics. *Chemical Physics Letters* **457**, (4-6) 448-452, doi: 10.1016/j.cplett.2008.04.050.
- Mudd,S.M., Fagherazzi,S., Morris,J.T., and Furbish,D.J. (2004) Flow, Sedimentation, and Biomass Production on a Vegetated Salt Marsh in South Carolina: Toward a Predictive Model of Marsh Morphologic and Ecologic Evolution. In: *The Ecogeomorphology of Salt Marshes*, 165-187. Edited by Fagherazzi,S., Marani,M., and Blum,L.K., Washington D.C., AGU.
- Murray,B.G., Jr. (2001) Are ecological and evolutionary theories scientific? *Biological Reviews* **76**, (2) 255-289, doi: 10.1017/S146479310100567X.
- Nielsen,S.N. and Ulanowicz,R.E. (2000) On the consistency between thermodynamical and network approaches to ecosystems. *Ecol.Modelling* **132**, (1-2) 23-31.
- Niven,R.K. (2009) Steady state of a dissipative flow-controlled system and the maximum entropy production principle. *Physical Review E (Statistical, Nonlinear, and Soft Matter Physics)* **80**, (2) 021113-021115, doi: 10.1103/PhysRevE.80.021113.
- O'Neill, R. V., DeAngelis, D. L., Waide, J. B., and Allen, T. F. H. (1986) A hierarchical concept of ecosystems. Monographs in population biology, Princeton, Princeton University Press, **23**, 1-253.
- Odum,E.P. (1969) The strategy of ecosystem development. *Science* **164**, (3877) 262-270.
- Odum,H.T. and Pinkerton,R.C. (1955) Time's speed regulator: the optimum efficiency for maximum power output in physical and biological systems. *Am.Sci.* **43**, 321-343.
- Odum,H.T. (1988) Self-Organization, Transformity, and Information. *Science* **242**, (4882) 1132-1139.

- Paltridge, G.W. (1975) Global dynamics and climate—a system of minimum entropy exchange. *Q.J.Roy.Met.Soc.* **104**, 927-945.
- Peters, R.H. (1976) Tautology in evolution and ecology. *Am Nat* **110**, (971) 1-12.
- Pfeiffer, T. and Bonhoeffer, S. (2002) Evolutionary consequences of tradeoffs between yield and rate of ATP production. *Zeitschrift für Physikalische Chemie* **216**, (1) 51-63, doi: 10.1524/zpch.2002.216.1.051.
- Prigogine, I. and Nicolis, G. (1971) Biological order, structure and instabilities. *Q.Rev.Biophys.* **4**, 107-148.
- Pueyo, S., He, F., and Zillio, T. (2007) The maximum entropy formalism and the idiosyncratic theory of biodiversity. *Ecology Letters* **10**, (11) 1017-1028, doi: 10.1111/j.1461-0248.2007.01096.x.
- Russell, J.B. and Cook, G.M. (1995) Energetics of bacterial growth: balance of anabolic and catabolic reactions. *Microbiol.Mol.Biol.Rev.* **59**, (1) 48-62.
- Schneider, E.D. and Kay, J.J. (1994) Complexity and thermodynamics: towards a new ecology. *Futures* **26**, (6) 626-647.
- Schrödinger, E. (1944) *What is life?*, Cambridge, UK, Cambridge University Press
- Shapiro, J.A. (1998) Thinking about bacterial populations as multicellular organisms. *Annu.Rev.Microbiol.* **52**, 81-104.
- Sogin, M.L., Morrison, H.G., Huber, J.A., Welch, D.M., Huse, S.M., Neal, P.R., Arrieta, J.M., and Herndl, G.J. (2006) Microbial diversity in the deep sea and the underexplored "rare biosphere". *PNAS* 0605127103, doi: 10.1073/pnas.0605127103.
- Swenson, R. (1989) Emergent attractors and the law of maximum entropy production: Foundations to a theory of general evolution. *Systems Research* **6**, (3) 187-197.
- Toussaint, O. and Schneider, E.D. (1998) The thermodynamics and evolution of complexity in biological systems. *Comp.Biochem.Physiol.A* **120**, (1) 3-9.
- Traulsen, A. and Nowak, M.A. (2006) Evolution of cooperation by multilevel selection. *PNAS* **103**, (29) 10952-10955, doi: 10.1073/pnas.0602530103.
- Ulanowicz, R.E. (1986) *Growth and development: ecosystems phenomenology*, New York, Springer-Verlag, 203 pp.
- Ulanowicz, R.E. and Hannon, B.M. (1987) Life and the production of entropy. *Proc.R.Soc.London, Ser.B* **232**, (1267) 181-192.
- Ulanowicz, R.E. and Platt, T. (1985) Ecosystem theory for biological oceanography. *Can. Bull. Fish. Aquat. Sci.*, 213, Ottawa, 260 pp.

- Vallino, J.J., Hopkinson, C.S., and Hobbie, J.E. (1996) Modeling bacterial utilization of dissolved organic matter: Optimization replaces Monod growth kinetics. *Limnol. Oceanogr.* **41**, (8) 1591-1609.
- Vallino, J.J. (2003) Modeling Microbial Consortia as Distributed Metabolic Networks. *Biol Bull* **204**, (2) 174-179.
- Vuddagiri, S.R., Hall, K.R., and Eubank, P.T. (2000) Dynamic Modeling of Reaction Pathways on the Gibbs Energy Surface. *Ind. Eng. Chem. Res.* **39**, (2) 508-517, doi: 10.1021/ie9904091S0888-5885(99)00409-1.
- Wang, J., Bras, R.L., Lerdau, M., and Salvucci, G.D. (2007) A maximum hypothesis of transpiration. *JGR Biogeosciences* **112**, (G03010) 1-11, doi: 10.1029/2006JG000255.
- Washida, T. (1995) Ecosystem configurations consequent on the maximum respiration hypothesis. *Ecol. Modelling* **78**, (3) 173-193.
- Weber, B.H., Depew, D.J., and Smith, J.D. (1988) Entropy, information, and evolution: New perspective on physical and biological evolution. MIT Press, Cambridge, MA, 390 pp.
- West, G., Brown, J.H., and Enquist, B.J. (1997) A General Model for the Origin of Allometric Scaling Laws in Biology. *Science* **276**, (5309) 122-126, doi: 10.1126/science.276.5309.122.
- Williams, H.T.P. and Lenton, T.M. (2008) Environmental regulation in a network of simulated microbial ecosystems. *PNAS* **105**, (30) 10432-10437, doi: 10.1073/pnas.0800244105.
- Wilson, D.S. and Sober, E. (1989) Reviving the superorganism. *Journal of Theoretical Biology* **136**, (3) 337-356.
- Wilson, D.S. and Wilson, E.O. (2008) Evolution "for the Good of the Group". *Am. Sci.* **96**, (5) 380-389, doi: 10.1511/2008.74.1.
- Wittebolle, L., Vervaeren, H., Verstraete, W., and Boon, N. (2008) Quantifying Community Dynamics of Nitrifiers in Functionally Stable Reactors. *Applied and Environmental Microbiology* **74**, (1) 286-293, doi: 10.1128/AEM.01006-07.



## Figure Captions

**Fig. 1.** Simple distributed metabolic network for a methanotrophic-base food web. Seven biological structures,  $\mathcal{S}_i$ , catalyze the reactions, including degradation of detrital N (dN) and C (dC).

**Fig. 2.** Illustration of how the partition function ( $\sigma_i$ ) controls synthesis of biological structures,  $\mathcal{S}_i$ .

**Fig. 3.** Concentration of (a) some state variables and (b) all biological structures in  $\mu\text{M}$ , except for dC, which is in mM.

**Fig. 4.** Redox,  $\zeta_{NADH}$ , and energy,  $\zeta_{ATP}$ , state variables ( $\mu\text{mol mmol}^{-1}$ ), and entropy production rate,  $\langle dS / dt \rangle$ , averaged over eight day intervals for the duration of the simulation.

**Fig. 5.** Control variables for (a) partitioning of biological structure synthesis,  $\sigma_i$ , and (b) ATP coupling to metabolic reactions,  $\eta_i$ . Note, plot (a) only shows control variables about the time of nitrate depletion, from 50 to 100 d.

Fig. 1. Vallino

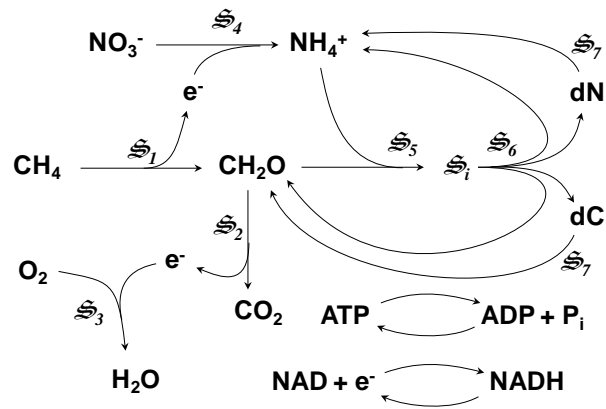


Fig. 2 Vallino

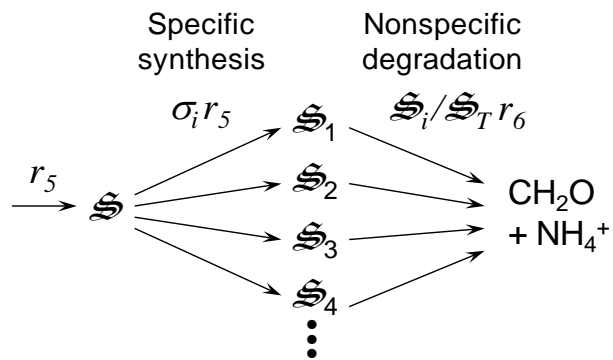


Fig. 3 Vallino

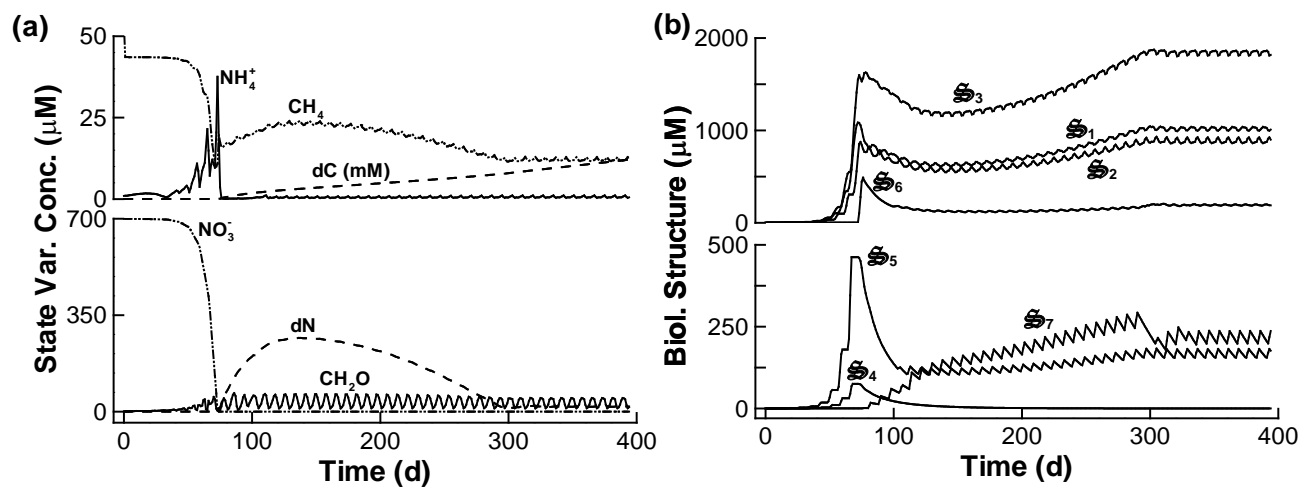


Fig. 4 Vallino

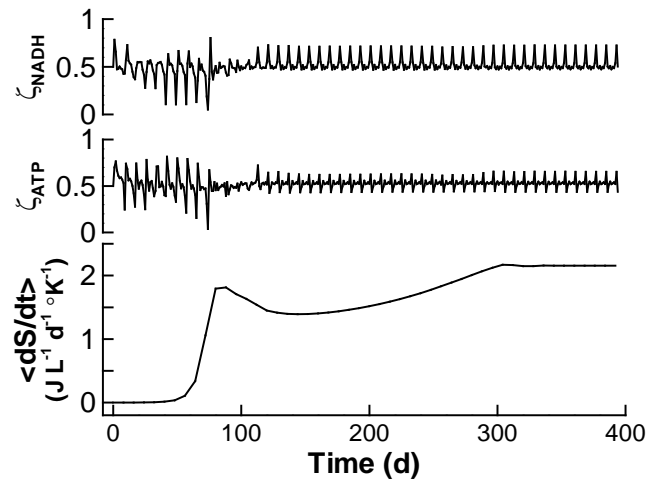


Fig. 5 Vallino

