

Virioplankton abundance from multiple cruises at the Bermuda Atlantic Time Series Station (BATS), Western Sargasso Sea from 2000-2011 (Ocean Microbial Observatory project)

Website: <https://www.bco-dmo.org/dataset/543808>

Data Type: Cruise Results

Version: 1

Version Date: 2014-12-16

Project

» [Transitions in the Surface Layer and the Role of Vertically Stratified Microbial Communities in the Carbon Cycle - An Oceanic Microbial Observatory](#) (Ocean Microbial Observatory)

Contributors	Affiliation	Role
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Abstract

Virioplankton abundances were measured from samples collected from January 2000 to December 2011 at the Bermuda Atlantic Time Series Station (BATS), Western Sargasso Sea, as part of the larger BATS program aboard the R/V *Weatherbird II* or the R/V *Atlantic Explorer*. Supporting data provided by the BATS time-series program and are available at (<http://bats.bios.edu/>).

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Coverage

Spatial Extent: Lat:31.67 Lon:-64.17

Temporal Extent: 2000-01-28 - 2009-12-09

Dataset Description

Virioplankton abundances were measured from samples collected from January 2000 to December 2011 as part of the larger BATS program aboard the R/V *Weatherbird II* or the R/V *Atlantic Explorer*. Supporting data provided by the BATS time-series program and are available at (<http://bats.bios.edu/>).

Acquisition Description

Methodology: from Parsons et al (2011):

Study site and sample collection:

Samples were collected aboard the RV Weatherbird II or the RV Atlantic Explorer at the BATS site (31° 40' N, 64°10' W). All cruises were conducted as part of the larger BATS program and sampled at least monthly with biweekly sampling between February and April. This sampling strategy has been successful in revealing the major temporal microbial and biogeochemical patterns at this site (Carlson and Ducklow, 1996; Steinberg et al., 2001; Morris et al., 2005; Carlson et al., 2009; Treusch et al., 2009; Lomas et al., 2010). A broader assessment of the BATS biogeochemical data is presented in Deep Sea Research II in 1996 (volume 43, issues 2–3) and 2001 (volume 48, issues 8–9).

Samples for virioplankton (0, 20, 40, 60, 80, 100, 140, 160, 200, 250 and 300 m) and bacterioplankton (0, 10, 20, 40, 60, 80, 100, 120, 140, 160, 200, 250 and 300 m) were collected at the BATS site from January 2000 to December 2009 via conductivity, temperature, depth profiling rosette equipped with 12 l Niskin bottles. The 120 m virioplankton sample was added after October 2007. Throughout the entire time-series, all virioplankton samples were fixed with 0.02 μm filtered formalin (1% final concentration), placed in 5 ml cryovials and flash frozen in liquid nitrogen (Wen et al., 2004) until processing (within 12 weeks of collection). Samples for bacterioplankton abundance were fixed with 0.2 μm filtered glutaraldehyde (1% final concentration) and stored at either 4 °C for 72 h or flash frozen and subsequently stored at –80 °C for up to 6 months until processing as described in Steinberg et al (2001). Storage tests demonstrated no appreciable loss of virioplankton or bacterioplankton abundance when stored in liquid nitrogen for periods up to 6 months (unpublished data). Picophytoplankton samples were collected at the same depths through 250 m from October 2001 to December 2009 (Casey et al., 2007). Samples for fluorescence in situ hybridization (FISH) of specific heterotrophic bacterioplankton lineages were collected from the upper 300 m from January 2003 to December 2005 (Carlson et al., 2009).

Biogeochemical and physical data collected at the BATS site are available at <http://bats.bios.edu>. The MLD was determined as the depth where potential density ($\sigma\text{-t}$) of the water was equal to sea surface $\sigma\text{-t}$ plus the equivalent in $\sigma\text{-t}$ to a 0.2 °C decrease in temperature (Sprintall and Tomczak, 1992). Contour plots were created in Ocean Data View (R Schlitzer, <http://odv.awi.de/>) with VG Gridding and linear mapping adjusted to the median of each data set. Statistics (Pearson's correlation and two-tailed Student's t-test for unequal variances), ratios and percent contributions were determined using Microsoft Excel.

Virioplankton abundance:

Virioplankton abundance was enumerated according to the methods of Noble and Fuhrman (1998). Briefly, water samples were filtered on to 0.02 μm Anodisc aluminum oxide filters (Whatman, Kent, UK), stained with SYBR Green I (Molecular Probes Inc., Eugene, OR, USA), and enumerated via epifluorescence microscopy using an Olympus AX70 microscope (Olympus, Tokyo, Japan) equipped with a Toshiba CCD video camera (Irvine, CA, USA) and Pro-series Capture Kit version 4.5 (I-CUBE, Crofton, MD, USA). Ten images from each sample were processed with scripts written in Image Pro Plus (Media Cybernetics, Bethesda, MD, USA) for particles sized 0.01–0.27 μm^2 , using the clean borders function (cells touching the edge of the image or grid were omitted). We consider these estimates of viral abundance conservative because it is possible that some viral particles less than one pixel were omitted from the final count. We performed pairwise comparisons of automated versus manual enumeration of virioplankton abundance to determine any discrepancies between the two approaches. Samples collected along a gradient from offshore (BATS; n=92) to onshore waters of Bermuda (n=32) were highly correlated with automated counts being slightly greater than manual counts (slope=1.07, r=0.99, n=134). The lower estimates of viral abundance from manual counts may have resulted from image fading during enumeration and/or operator fatigue. We argue that for this study, the automated image analysis was the most reliable approach for viral particle enumeration. The coefficient of variation for the automated counts averaged 11% (n=1517).

Processing Description

BCO-DMO Processing:

- added conventional header with dataset name, PI name, version date, reference information
- renamed parameters to BCO-DMO standard

- converted longitudes to negative values to represent degrees West
- converted -999 to nd for 'no data'

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Parameters

Parameter	Description	Units
station	BATS cruise number during which sample was collected	unitless
cruise_ID	BATS cruise ID for the sample that matches the BATS sample collected from the same niskin	unitless
date_in	date of collection at the time of CTD entry year month day	unitless
decyear	decimal year	unitless
lat_in	Latitude at the time of CTD entry in degrees N	decimal degrees
lon_in	Longitude at the time of CTD entry in degrees W	decimal degrees
depth	the actual depth in meters	meters
depth_nom	bottle target depths in meters	meters
depth_mixed	mixed layer depth in meters; MLD was determined as the depth where potential density (σ_t) of the water was equal to sea surface σ_t plus the equivalent in σ_t to a 0.2 °C decrease in temperature (Sprintall and Tomczak 1992).	meters
abund_vir	Virioplankton abundance 10^9 cells per liter	10^9 cells per liter
abund_vir_sd	standard deviation for Virioplankton Abundance in 10^9 cells per liter	10^9 cells per liter

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Instruments

Dataset-specific Instrument Name	Niskin bottle
Generic Instrument Name	Niskin bottle
Dataset-specific Description	12 liter Niskin bottles
Generic Instrument Description	A Niskin bottle (a next generation water sampler based on the Nansen bottle) is a cylindrical, non-metallic water collection device with stoppers at both ends. The bottles can be attached individually on a hydrowire or deployed in 12, 24 or 36 bottle Rosette systems mounted on a frame and combined with a CTD. Niskin bottles are used to collect discrete water samples for a range of measurements including pigments, nutrients, plankton, etc.

Dataset-specific Instrument Name	CTD
Generic Instrument Name	CTD profiler
Generic Instrument Description	The Conductivity, Temperature, Depth (CTD) unit is an integrated instrument package designed to measure the conductivity, temperature, and pressure (depth) of the water column. The instrument is lowered via cable through the water column and permits scientists observe the physical properties in real time via a conducting cable connecting the CTD to a deck unit and computer on the ship. The CTD is often configured with additional optional sensors including fluorometers, transmissometers and/or radiometers. It is often combined with a Rosette of water sampling bottles (e.g. Niskin, GO-FLO) for collecting discrete water samples during the cast. This instrument designation is used when specific make and model are not known.

Dataset-specific Instrument Name	Epifluorescence Microscope
Generic Instrument Name	Fluorescence Microscope Image Analysis System
Dataset-specific Description	Olympus AX70 microscope (Olympus, Tokyo, Japan) equipped with a Toshiba CCD video camera (Irvine, CA, USA)
Generic Instrument Description	A Fluorescence (or Epifluorescence) Microscope Image Analysis System uses semi-automated color image analysis to determine cell abundance, dimensions and biovolumes from an Epifluorescence Microscope. An Epifluorescence Microscope (conventional and inverted) includes a camera system that generates enlarged images of prepared samples. The microscope uses excitation ultraviolet light and the phenomena of fluorescence and phosphorescence instead of, or in addition to, reflection and absorption of visible light.

Dataset-specific Instrument Name	Flow Cytometer
Generic Instrument Name	Flow Cytometer
Dataset-specific Description	Becton Dickenson (Franklin Lakes, NJ, USA; formerly Cytopeia) high speed jet-in-air InFlux flow cytometer, using a 488 nm blue excitation laser, appropriate Chl-a (692±20 nm) and phycoerythrin (580±15 nm) bandpass filters.
Generic Instrument Description	Flow cytometers (FC or FCM) are automated instruments that quantitate properties of single cells, one cell at a time. They can measure cell size, cell granularity, the amounts of cell components such as total DNA, newly synthesized DNA, gene expression as the amount messenger RNA for a particular gene, amounts of specific surface receptors, amounts of intracellular proteins, or transient signalling events in living cells. (from: http://www.bio.umass.edu/micro/immunology/facs542/facswhat.htm)

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Deployments

OC449-10

Website	https://www.bco-dmo.org/deployment/59029
Platform	R/V Oceanus
Start Date	2008-12-14
End Date	2008-12-18
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0810

Website	https://www.bco-dmo.org/deployment/58062
Platform	R/V Atlantic Explorer
Start Date	2008-05-03
End Date	2008-05-25
Description	One in a series of transect cruises to study the biological and biogeochemical aspects of the marine phosphorus cycle. Note the cruise identifiers for the Atlantic Explorer were originally formatted as XYY## (e.g. X0806 was the 6th cruise in 2008). The data files include cruise IDs of this type. The vessel operator changed the cruise ID syntax several years after the cruise and the official cruise ID syntax was changed to AEYY##. For example, AE0810 should be the same cruise as X0810. One exception for this dataset is that X0804 is cruise ID AE0810 (unclear how the cruise numbering scheme got so confused). Database validation showed that AE0804 was not the correct cruiseid based on information at R2R. The cruiseid was then updated to reflect the corrected information (the May 2008 cruise was AE0810. Additional Information from R2R Site

AE0811

Website	https://www.bco-dmo.org/deployment/58921
Platform	R/V Atlantic Explorer
Start Date	2008-05-28
End Date	2008-06-01
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0813

Website	https://www.bco-dmo.org/deployment/58922
Platform	R/V Atlantic Explorer
Start Date	2008-06-21
End Date	2008-06-30
Description	Sampling was conducted monthly from May 2008-April 2010 at the Bermuda Atlantic Time-Series (BATS). Samples were collected for DNA analysis from four depths in the upper water column and from 150 m particle traps. Cruise information and original data are available from the NSF R2R data catalog.

AE0816

Website	https://www.bco-dmo.org/deployment/58923
Platform	R/V Atlantic Explorer
Start Date	2008-07-15
End Date	2008-07-21
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0820

Website	https://www.bco-dmo.org/deployment/58924
Platform	R/V Atlantic Explorer
Start Date	2008-08-11
End Date	2008-08-17
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0823

Website	https://www.bco-dmo.org/deployment/58925
Platform	R/V Atlantic Explorer
Start Date	2008-09-09
End Date	2008-09-14
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0826

Website	https://www.bco-dmo.org/deployment/58926
Platform	R/V Atlantic Explorer
Start Date	2008-10-06
End Date	2008-10-12
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0901

Website	https://www.bco-dmo.org/deployment/58920
Platform	R/V Atlantic Explorer
Start Date	2009-02-07
End Date	2009-02-11
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0902

Website	https://www.bco-dmo.org/deployment/58919
Platform	R/V Atlantic Explorer
Start Date	2009-02-21
End Date	2009-02-23

AE0903

Website	https://www.bco-dmo.org/deployment/58918
Platform	R/V Atlantic Explorer
Start Date	2009-03-03
End Date	2009-03-06
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0904

Website	https://www.bco-dmo.org/deployment/58917
Platform	R/V Atlantic Explorer
Start Date	2009-03-16
End Date	2009-03-20
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0905

Website	https://www.bco-dmo.org/deployment/58916
Platform	R/V Atlantic Explorer
Start Date	2009-03-31
End Date	2009-04-02
Description	Sampling was conducted monthly from May 2008-April 2010 at the Bermuda Atlantic Time-Series (BATS). Samples were collected for DNA analysis from four depths in the upper water column and from 150 m particle traps. Cruise information and original data are available from the NSF R2R data catalog.

AE0907

Website	https://www.bco-dmo.org/deployment/58915
Platform	R/V Atlantic Explorer
Start Date	2009-04-14
End Date	2009-04-17
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0909

Website	https://www.bco-dmo.org/deployment/58914
Platform	R/V Atlantic Explorer
Start Date	2009-05-14
End Date	2009-05-20
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0911

Website	https://www.bco-dmo.org/deployment/58913
Platform	R/V Atlantic Explorer
Start Date	2009-06-09
End Date	2009-06-13
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0916

Website	https://www.bco-dmo.org/deployment/58912
Platform	R/V Atlantic Explorer
Start Date	2009-07-13
End Date	2009-07-19
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0919

Website	https://www.bco-dmo.org/deployment/58911
Platform	R/V Atlantic Explorer
Start Date	2009-08-13
End Date	2009-08-17
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0921

Website	https://www.bco-dmo.org/deployment/58910
Platform	R/V Atlantic Explorer
Start Date	2009-09-09
End Date	2009-09-15
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0924

Website	https://www.bco-dmo.org/deployment/58909
Platform	R/V Atlantic Explorer
Start Date	2009-10-08
End Date	2009-10-12
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0926

Website	https://www.bco-dmo.org/deployment/58908
Platform	R/V Atlantic Explorer
Start Date	2009-11-06
End Date	2009-11-10
Description	Cruise information and original data are available from the NSF R2R data catalog.

AE0928

Website	https://www.bco-dmo.org/deployment/58907
Platform	R/V Atlantic Explorer
Start Date	2009-12-07
End Date	2009-12-11
Description	Cruise information and original data are available from the NSF R2R data catalog.

BATS_cruises

Website	https://www.bco-dmo.org/deployment/58883
Platform	Unknown Platform
Report	http://bats.bios.edu/index.html
Start Date	1988-10-20
Description	Bermuda Institute of Ocean Science established the Bermuda Atlantic Time-series Study with the objective of acquiring diverse and detailed time-series data . BATS makes monthly measurements of important hydrographic, biological and chemical parameters throughout the water column at the BATS Study Site, located at 31 40N, 64 10W.

AE0801

Website	https://www.bco-dmo.org/deployment/776209
Platform	R/V Atlantic Explorer
Start Date	2008-01-14
End Date	2008-01-18
Description	Bermuda Atlantic Time-series Study (BATS)/Hydrostation-S

AE0803

Website	https://www.bco-dmo.org/deployment/776212
Platform	R/V Atlantic Explorer
Start Date	2008-02-22
End Date	2008-02-24
Description	Bermuda Atlantic Time-series Study (BATS)/Hydrostation-S

AE0804

Website	https://www.bco-dmo.org/deployment/776215
Platform	R/V Atlantic Explorer
Start Date	2008-03-12
End Date	2008-03-16
Description	Bermuda Atlantic Time-series Study (BATS)/Hydrostation-S

HB0805

Website	https://www.bco-dmo.org/deployment/58451
Platform	R/V Henry B. Bigelow
Start Date	2008-06-19
End Date	2008-07-20
Description	This was a survey cruise collecting oceanographic profile (hydrography) data (as well as other data) and was part of the ongoing oceanographic data collection project.

AE0805

Website	https://www.bco-dmo.org/deployment/776217
Platform	R/V Atlantic Explorer
Start Date	2008-03-24
End Date	2008-03-26
Description	Bermuda Atlantic Time-series Study (BATS)/Hydrostation-S

AE0808

Website	https://www.bco-dmo.org/deployment/776221
Platform	R/V Atlantic Explorer
Start Date	2008-04-14
End Date	2008-04-22
Description	Bermuda Atlantic Time-series Study (BATS)/Hydrostation-S and for the Project: Carbon Flux Through the Twilight Zone/Mercury Species Cycling and Flux in the Oceanic Water Column.

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Project Information**Transitions in the Surface Layer and the Role of Vertically Stratified Microbial Communities in the Carbon Cycle - An Oceanic Microbial Observatory (Ocean Microbial Observatory)**

Website: <http://www.bios.edu/research/projects/oceanic-microbial-observatory/>

Coverage: Bermuda Atlantic Time-Series study site

(Adapted from the NSF award abstract) The premise of this project is that stratified bacterioplankton clades engage in specialized biogeochemical activities that can be identified by integrated oceanographic and microbiological approaches. Specifically, the objective of this project is to assess if the mesopelagic microbial community rely on diagenetically altered organic matter and subcellular fragments that are produced by microbial processes in the euphotic zone and delivered into the upper mesopelagic by sinking or mixing. In past efforts this microbial observatory had greater success cultivating members of the euphotic zone microbial community, and revealed an unanticipated growth requirement for reduced sulfur compounds in alphaproteobacteria of the SAR11 clade. Genomic information showed that intense competition for substrates imposes trade-offs on bacterioplankton - there are regions of N dimensional nutrient space where specialists win. We postulate that specific growth requirements may explain some the regular spatial and temporal patterns that have been observed in upper mesopelagic bacterioplankton communities, and the difficulties of culturing some of these organisms. The specific objectives of this project are: 1) to produce ¹³C and ¹⁵N labeled subcellular (e.g., soluble, cell wall, and membrane) and DOM fractions from photosynthetic plankton cultures and use stable isotope probing to identify specific clades in the surface and upper mesopelagic microbial community that assimilate fractions of varying composition and lability. 2) to use fluorescence in situ hybridization approaches to monitor temporal and spatial variability of specific microbial populations identified from the SIP and HTC experiments. To increase resolution we will use CARD-FISH protocols. 3) to measure the proteomes of bacterioplankton communities to identify highly translated genes in the surface layer and upper mesopelagic, and community responses to seasonal nutrient limitation. 4) and, to cultivate these organisms via high

throughput culturing (HTC) by pursuing the hypothesis that they require specific nutrient factors and/or diagenetically altered organic substrates. Complete genome sequences from key organisms will be sought and used as queries to study patterns of natural variation in genes and populations that have been associated with biogeochemically important functions.

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Funding

Funding Source	Award
NSF Division of Ocean Sciences (NSF OCE)	OCE-0802004

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