

1 Title: Response of dark respiration to temperature in *Eriophorum vaginatum* from a 30 year old  
2 transplant experiment in Alaska

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14

15 **Abstract**

16 Background: In the Arctic region, temperature increases are expected to be greater under  
17 anticipated climate change than the global average. Understanding how dark respiration ( $R_d$ ) of  
18 common Arctic plant species acclimates to changes in the environment is therefore important for  
19 predicting changes to the Arctic carbon balance.

20 Aims: To investigate the influence of genotype and growing environment on  $R_d$ , the temperature  
21 response ( $Q_{10}$ ) of  $R_d$ , and foliar N ( $N_{\text{leaf}}$ ) of the Arctic sedge *Eriophorum vaginatum*.

22 Method: We measured  $R_d$ , its  $Q_{10}$  and  $N_{\text{leaf}}$  of *E. vaginatum* populations that were reciprocally  
23 transplanted 30 years previously along a latitudinal transect of 370 km in northern Alaska.

24 Results:  $R_d$  and  $Q_{10}$  did not differ among populations (ecotypes) of *E. vaginatum*, but the local  
25 environment had a significant effect on both variables.  $R_d$  as well as  $N_{\text{leaf}}$  was higher in northern,  
26 colder sites, while  $Q_{10}$  was lower there.

27 Conclusions:  $R_d$  in the different populations of *E. vaginatum* is a very plastic trait and controlled  
28 by growing environment, as is  $N_{\text{leaf}}$ . The lower  $Q_{10}$  values in the northern sites were most likely a  
29 consequence of substrate inhibition of  $R_d$  at higher temperatures.

30 Keywords:  $Q_{10}$ , common garden experiment, leaf respiration, *Eriophorum vaginatum*, Arctic leaf  
31 nitrogen, reciprocal transplant

32

### 33 **Introduction**

34 Autotrophic respiration is a key component of the carbon budget of an ecosystem, contributing  
35 30-65% of the total CO<sub>2</sub> released into the atmosphere (e.g. Janssen et al 2001; Luo et al. 2007). It  
36 is also widely acknowledged that understanding the long-term acclimation of autotrophic  
37 respiration is important in light of the anticipated global increase in temperature (e.g. Atkin and  
38 Tjoelker 2003; Luo et al. 2007; Atkin et al. 2008). Especially in the Arctic, temperature increases  
39 are expected to be greater than the global average (Solomon et al. 2007). Observations have  
40 already shown Arctic-wide warming trends since 1958 (0.11°C per decade (Kaufman et al.  
41 2009), while Chapin et al. (2005) found that summer warming in arctic Alaska and western  
42 Canada accelerated to about 0.3° to 0.4°C per decade between 1961–2004. Currently, the Arctic  
43 acts as a modest carbon sink (McGuire et al 2009), but with changing temperatures it is  
44 important to understand how climate change can affect different components of carbon fluxes,  
45 such as autotrophic respiration.

46  
47 Previous studies have shown that plant respiration acclimates to altered growth temperatures. For  
48 example, when measured at the same temperature, plants that have acclimated to lower growing  
49 temperatures had a higher foliar dark respiration ( $R_d$ ) and a higher short-term temperature  
50 responsiveness of  $R_d$  compared with plants that were acclimated to higher growing temperatures  
51 (e.g. Strain and Chase 1966; Bolstad et al. 2003; Loveys et al. 2003; Bruhn et al. 2007; Campbell  
52 et al. 2007; Tjoelker et al. 2008). The short-term temperature responsiveness of  $R_d$  can be  
53 expressed by using  $Q_{10}$ , which represents the factor of change of  $R_d$  per 10 °C increase in  
54 temperature, usually measured between 10°C and 20°C. Atkin and Tjoelker (2003) described  
55 two different kinds of temperature acclimation of plant respiration: (1) altered temperature

56 sensitivity (i.e. a change in  $Q_{10}$ ), and (2) a shift up or down in the overall temperature response  
57 curve (with no change in  $Q_{10}$ , but with a change in the intercept of the response curve). The latter  
58 form of acclimation is likely to be more common in newly developed tissue, while the former  
59 occurs at a shorter time scale, in tissue that is already fully developed (e.g. Atkin et al. 2000; Ow  
60 et al. 2008). Long-term cold acclimation is also often accompanied by an increase in leaf  
61 nitrogen ( $N_{\text{leaf}}$ ) (e.g. Tjoelker et al. 1999; Lee et al. 2005; Tjoelker 2008), which is associated  
62 with increased investment in glycolytic and mitochondrial proteins. With more of these proteins,  
63 higher  $R_d$  rates at the same temperature are possible and consequently cold acclimation of  $R_d$  in  
64 the long-term (Atkin et al. 2005; Tjoelker et al. 2008).

65  
66 One drawback of many studies of acclimation of respiration to temperature has been that the  
67 acclimation period at a lower or higher temperature has often encompassed periods ranging from  
68 one week or one growing season, which might not be a suitable time scale for studying effects of  
69 climate change. In this study, we present results from a 30-year-old reciprocal transplant  
70 experiment. In August 1980 and 1982, Shaver et al. (1986) established a latitudinal transect with  
71 six common gardens along the Dalton Highway in Alaska, in which whole tussocks of the sedge  
72 *Eriophorum vaginatum* L. were reciprocally transplanted over a distance of 370 km. Each of the  
73 gardens was located more than 200 m from the road in order to avoid artefacts of the road traffic  
74 (e.g., dust deposition). Each common garden included locally transplanted tussocks as well as  
75 tussocks that originated from the other garden sites. *E. vaginatum* is one of the most common  
76 and abundant species in northern Alaska (Britton 1966). It is a clonal species and individual  
77 tillers typically live less than 8 years (Fetcher and Shaver 1983; Mark et al. 1985), meaning that  
78 the entire biomass of the transplanted plants had been replaced at least 4 times before we

79 sampled them. The common gardens in the transect, which spans 3.30 degrees in latitude (~ 370  
80 km), provided a unique opportunity to study the long-term acclimation of foliar respiration of  
81 plants from one location. In addition, they allowed us to determine whether different populations  
82 (ecotypes) of the same species had different physiological responses to changes in the growing  
83 environment. For the populations in the transect, differences in morphology and growth between  
84 ecotypes have been established previously (Shaver et al. 1986; Fetcher and Shaver 1990).  
85 Bennington et al. (2012) showed that tussocks that were retransplanted into their sites of origin  
86 had higher survival than tussocks from elsewhere on the transect, thus demonstrating home-site  
87 advantage. Likewise, we were interested whether differences in  $R_d$  rates between ‘home and  
88 away’ populations could be detected after 30 years of growth in the different sites. Additionally,  
89 we wanted to investigate if observed differences in rates or the temperature response of  $R_d$  were  
90 related to any changes in the values of  $N_{leaf}$ . The hypotheses tested in this study were: (1) There  
91 is no difference within a site amongst the transplanted tussocks (local vs. non-local origin) in  
92 their  $R_d$  rates,  $N_{leaf}$ , or  $Q_{10}$  values (i.e. there is no ecotypic variation). (2) Between sites  
93 (gardens), the  $R_d$  values at a standardised temperature are higher at the northern sites that have  
94 lower average temperatures. (3) The leaves from the colder, more northern sites have higher  $N_{leaf}$   
95 values. (4) The  $Q_{10}$  values remain identical between sites, though the intercept of the temperature  
96 response curve differs).

97

## 98 **Material and methods**

99 Four of the six gardens described in Shaver et al. (1986) (No Name Creek, Coldfoot, Toolik  
100 Lake, and Sagwon) were visited between 16 and 23 July 2011 (Table 1). Two of these gardens  
101 are situated south of the Brooks Range (No Name Creek and Coldfoot), while the other two

102 (Toolik and Sagwon) are north of the Brooks Range. For 2011, the most northerly site (Sagwon)  
103 and the most southerly site (No Name creek) differed over 5 °C in average annual temperature  
104 (Table 1), and even more in the months leading up to the measurements (May-July 2011, Table  
105 1). Additional details about the installation of the common gardens, environmental variation, and  
106 variation in growth and flowering can be found in Shaver et al. (1986), Fetcher and Shaver  
107 (1990), and Bennington et al. (2012). In this study, no individuals from the Coldfoot population  
108 were measured and because not all the transplanted tussocks had survived, a balanced design of  
109 measurements was not possible.

110 Temperature response curves for  $R_d$  were measured in situ between 9:00 am and 6:00 pm with  
111 portable photosynthesis equipment fitted with an expanded temperature control kit (Li-Cor 6400  
112 and Li-Cor 6400-88, Li-Cor, Inc, Lincoln, USA). For each replicate, a selection of *E. vaginatum*  
113 leaves per tussock (3-9) was used. The mean  $\pm$  SD leaf temperatures ( $T_{leaf}$ ) ranged between  $10 \pm$   
114  $2.0$  °C and  $25 \pm 1.9$  °C for each curve and  $R_d$  measurements were taken at intervals of  $\sim 2.5$  °C.  
115 Each response curve took between 20 and 70 minutes, depending on how quickly the higher leaf  
116 chamber temperatures were reached. After the respiration measurements, the leaf samples were  
117 dried at 60 °C to a constant weight, ground and analysed for CHN with a Perkin-Elmer Series II  
118 2400 CHNS/O Analyzer (LECO Corporation, U.S.A.). The response of  $R_d$  to  $T_{leaf}$  was fitted by  
119 regression using a modified Arrhenius equation (e.g. Lloyd and Taylor 1994; Griffin et al. 2002):

$$120 \quad R_{dark} = a \cdot e^{bT_{leaf}}$$

121 where  $R_d$  is respiration rate,  $a$  and  $b$  are fitted parameters, respectively and  $T_{leaf}$  is leaf  
122 temperature. The  $Q_{10}$  values of the temperature response curve were then derived from:

$$123 \quad Q_{10} = \frac{R_{dT+10}}{R_{dT}}$$

124 where  $R_{dT}$  and  $R_{d(T+10)}$  are respiration rates at the temperature of  $T_{\text{leaf}}$  and  $T_{\text{leaf}} + 10$ . We chose  
125 this relatively simple equation because the range of temperatures at which we could measure  $R_d$   
126 did not include the maximum temperature for leaf respiration, which is about 55 °C for *E.*  
127 *vaginatum* (O. O'Sullivan, pers. comm.). This made fitting the temperature response to other  
128 equations (for example, a polynomial equation) more difficult.

129 Statistical analyses were carried out in R with the *agricolae* package (R Development Core Team  
130 2008). To analyse environmental (i.e. 'site' or 'garden') and genotype effects on  $R_d$ ,  $Q_{10}$ , and  
131  $N_{\text{leaf}}$ , we used an additive main effects multiplicative interaction (AMMI) model. Before the  
132 tests, the  $Q_{10}$  values were ln-transformed to obtain normally distributed data.

133

## 134 **Results**

135 The AMMI tests showed an effect of garden on  $R_d$  ( $P < 0.04$ ), but no effect of population and no  
136 interaction between genotype and garden ( $P < 0.29$  and  $0.62$ , respectively). Hence, within the  
137 gardens, the *E. vaginatum* leaves from different populations did not differ in their  $R_d$ . Between  
138 gardens, leaves from Sagwon had the highest  $R_d$  at 10°C (Figure 1A). A similar result prevailed  
139 for  $Q_{10}$  values, as the AMMI tests showed an effect of the environment (i.e. garden) on  $Q_{10}$  ( $P <$   
140  $0.02$ ), but no effect of population or interaction between population and garden ( $P < 0.63$  and  
141  $0.89$ , respectively). Overall, the northern sites Sagwon and Toolik Lake had significantly lower  
142 values for  $Q_{10}$  than the two southern sites (Figure 1B). In contrast, both garden and garden \*  
143 population significantly influenced the values for  $N_{\text{leaf}}$  ( $P < 0.001$  and  $P < 0.04$ , respectively),  
144 with the northern gardens having larger values (Figure 1C), implying that  $N_{\text{leaf}}$  of the different  
145 populations responded differently to the transplantation.

146

147 **Discussion**

148 The 30-year-old reciprocal transplant study showed that  $R_d$  and  $Q_{10}$  in *E. vaginatum* populations  
149 were quite plastic. We found no significant effect of population within gardens on these traits,  
150 therefore our first hypothesis (H1) was supported, which is also agrees with the general finding  
151 of thermal acclimation of  $R_d$  in higher plants (e.g. Strain and Chase 1966; Bolstad et al. 2003;  
152 Zaragossa-Castells et al. 2007; Tjoelker et al. 2008; Rodriguez-Calcerdera et al. 2010). However,  
153 this result contrasts with the response of life history and morphological variables, such as tussock  
154 survival rate and tiller size from the same experiment. For these traits, Bennington et al. (2012)  
155 found home-site advantage (i.e. advantage for the population that originated in the common  
156 garden) in tussock survival rates, as well as greater plasticity in tiller size in *E. vaginatum* that  
157 originated from the southern sites. Therefore, the lack of difference between populations in  
158 plasticity in  $R_d$  and  $Q_{10}$  (which are measured at the tissue level) cannot be extrapolated to the  
159 functioning of *E. vaginatum* at the whole plant level, especially if some ecotypes produce fewer  
160 tillers in their 'away' environment. In other words, although the physiological parameters on a  
161 tissue scale do acclimate, there are some genetic based population differences (e.g. in survival  
162 rate, tiller length) that can limit the ability of plants to respond to a change in environment.

163

164 No difference in  $Q_{10}$  was expected between research sites (gardens), as Atkin and Tjoelker  
165 (2003) suggested that long-term thermal acclimation represents a shift up or down in the overall  
166 temperature response curve (no change in  $Q_{10}$ , but with a change in the intercept of the response  
167 curve). However, the lower  $Q_{10}$  values in the colder, northern Sagwon site do not support our  
168 hypothesis (H4). Additionally, global differences in values of  $Q_{10}$  for  $R_d$  from Atkin and Tjoelker

169 (2003) show that colder, more northerly sites have higher, rather than lower  $Q_{10}$  values. In the  
170 context of these global patterns, it would be expected that a site, such as Sagwon would have  
171 higher, not lower,  $Q_{10}$  values when compared with the warmer, southern sites. It is possible that  
172 the latitudinal range included in this study ( $3.3^\circ$  latitude) is not large enough to reflect patterns  
173 that are observed globally. In addition, the pattern of cold acclimation resulting in higher  $Q_{10}$   
174 values, as described by Atkin and Tjoelker (2003), requires that in colder environments plants  
175 have a higher build-up of substrates (e.g. non-structural carbohydrates resulting from  
176 photosynthesis) due to a changed balance between  $R_d$  and foliar C uptake. The higher amount of  
177 substrate consequently allows for relatively higher  $R_d$  values with short-term warming, such as in  
178 a temperature response curve (Atkin and Tjoelker 2003). In our study, data on the non-structural  
179 carbohydrate content in the leaves is lacking. However, if this higher build-up of substrates did  
180 not occur in the colder, northern sites this could explain why the pattern described by Atkin and  
181 Tjoelker (2003) was not observed.

182 The increased values of  $N_{\text{leaf}}$  and  $R_d$  in the northern gardens (Figures 1A and 1C) supported our  
183 Hypotheses 2 and 3, and suggest an increased investment in the respiratory apparatus. Higher  
184 values for  $N_{\text{leaf}}$  at colder sites have been observed in boreal forest species (Tjoelker et al. 1999;  
185 Tjoelker et al. 2008) while increased protein levels and investment in mitochondrial volume have  
186 been found in cold-acclimated plants (Graham and Patterson 1983; Armstrong et al. 2006).  
187 Therefore, although the values of  $Q_{10}$  in the colder, northern sites do not suggest acclimation of  
188  $R_d$  themselves, the higher values for  $N_{\text{leaf}}$ , together with those for  $R_d$  give indirect support for a  
189 contrary conclusion. Higher values for  $R_d$  associated with lower values for  $Q_{10}$  at low  
190 temperatures have been observed in other studies. Xiong et al. (2000) found a greater  
191 temperature sensitivity of Antarctic species *Colobanthus quitensis* (Kunth) Bartl. and

192 *Deschampsia antarctica* E.Desv. when grown at higher (12 °C and 20 °C) rather than lower (7  
193 °C) temperatures, but higher respiration in the cold-acclimated plants when measured at the same  
194 temperature. Larigauderie and Körner (1995), however, showed that thermal acclimation of a  
195 range of species can differ widely, both within genera, growth forms, and habitats. In sum, it  
196 might be hard to observe the effects of thermal acclimation of  $R_d$  in cold-acclimated  
197 environments through *in situ* measurements without factors, such as substrate limitation being  
198 considered.

199 Overall, under the anticipated warming of the Arctic, the *E. vaginatum* populations in Alaska  
200 will probably acclimate their  $R_d$  and the  $Q_{10}$  with higher temperatures. This implies that changes  
201 in this species' abundance or biomass following Arctic warming are a more important factor to  
202 consider when studying the effects of Arctic warming on the C balance of this ecosystem.

203

## 204 **Conclusions**

205 This study shows that  $R_d$ ,  $Q_{10}$  and  $N_{leaf}$  are plastic traits in Alaskan populations of the species *E.*  
206 *vaginatum*, since the growing environments, rather than the genotypes explained most of the  
207 variation in these parameters. This thermal acclimation of  $R_d$  this species is probably facilitated  
208 through changes in protein levels and mitochondrial volume as indicated by changes in  $N_{leaf}$ . It  
209 has to be noted though that acclimation of  $R_d$  is a physiological response and that for the overall  
210 effects of Arctic warming on *E. vaginatum* other plant traits are important as well.

211

212

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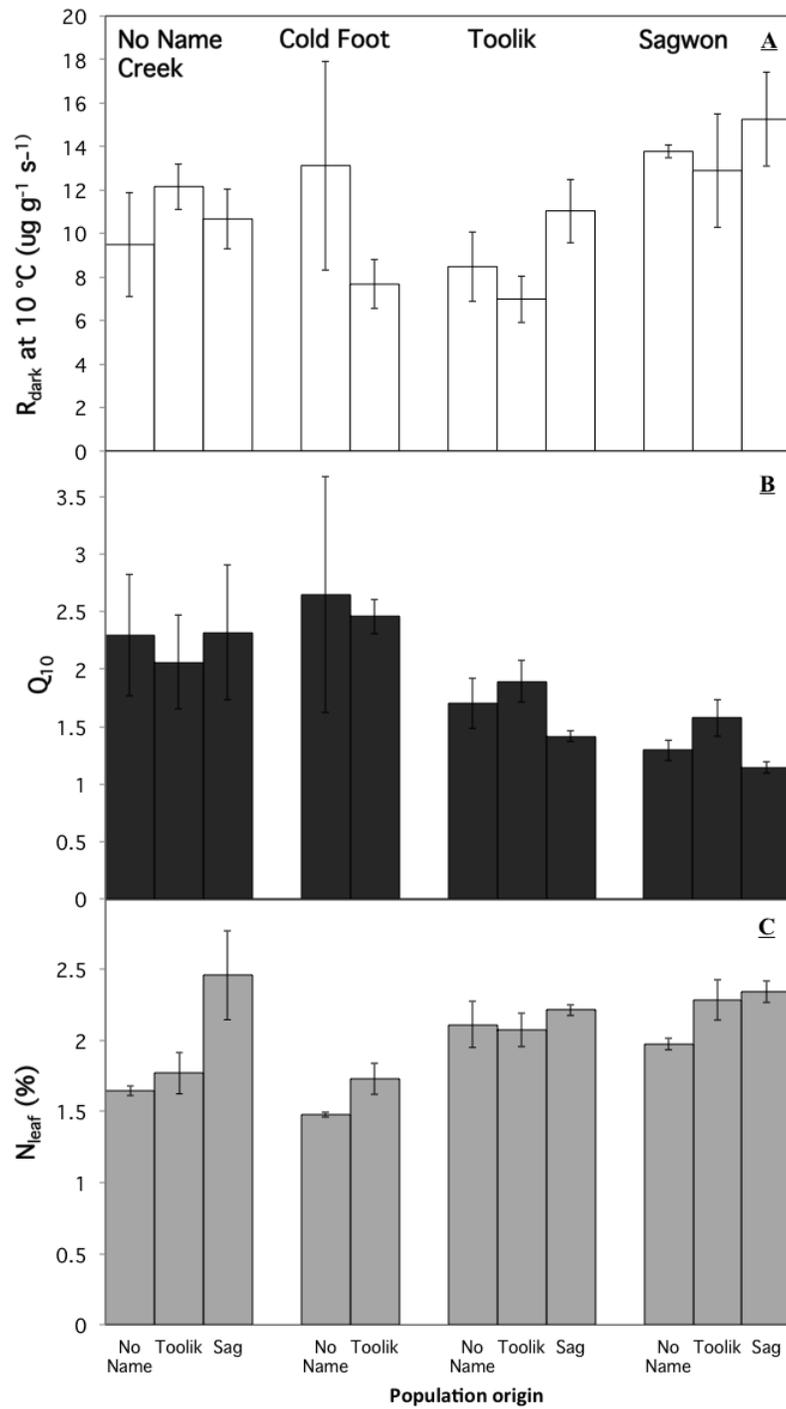
296 **Figure legends**

297

298 Figure 1. Average  $R_d$  at 10 °C (a),  $Q_{10}$  values of  $R_d$  (b) and average  $N_{leaf}$  (c) per population origin  
299 (denoted below) and common garden (denoted above) and their standard errors.

300

301



302 **Figure 1.**

303

304