GENETIC IDENTITY DETERMINES RISK OF POST-SETTLEMENT MORTALITY OF A MARINE FISH

LAURENT VIGLIOLA,1,4 PETER J. DOHERTY,2 MARK G. MEEKAN,1 DEVIN M. DROWN,3 M. ELIZABETH JONES,3
AND PAUL H. BARBER3

1Australian Institute of Marine Science, P.O. Box 40197, Casuarina MC Darwin NT 0811 Australia
2Australian Institute of Marine Science, PMB 3, Townsville MC Qld 4810 Australia
3Boston University Marine Program, 7 MBL Street, Woods Hole, Massachusetts 02543 USA

Abstract. Longitudinal sampling of four cohorts of Neopomacentrus filamentosus, a common tropical damselfish from Dampier Archipelago, Western Australia, revealed the evolution of size structure after settlement. Light traps collected premetamorphic individuals from the water column ("settlers") to establish a baseline for each cohort. Subsequently, divers collected benthic juveniles ("recruits") at 1–3-month intervals to determine the relative impacts of post-settlement mortality during the first three months. Growth trajectories for individual fish were back-calculated from otolith records and compared with nonlinear mixed-effects models. Size-selective mortality was detected in all cohorts with the loss of smaller, slower growing individuals. Three months after settlement, recruits showed significantly faster growth as juveniles, faster growth as larvae, and larger sizes as hatchlings. The timing and intensity of post-settlement selection differed among cohorts and was correlated with density at settlement. The cohort with the greatest initial abundance experienced the strongest selective mortality, with most of this mortality occurring between one and two months after settlement when juveniles began foraging at higher positions in the water column. Significant genetic structure was found between settlers and three-month-old recruits in this cohort as a result of natural selection that changed the frequency of mtDNA haplotypes measured at the control region. The extent of this genetic difference was enlarged or reduced by artificially manipulating the intensity of size-based selection, thus establishing a link between phenotype and haplotype. Sequence variation in the control region of the mitochondrial genome has been linked to mitochondrial efficiency and weight gain in other studies, which provides a plausible explanation for the patterns observed here.

Key words: growth rates; haplotypes; larval and juvenile fish; maternal and genetic inheritance; mortality; mtDNA; otoliths; predation; recruitment; settlement; size selection.

INTRODUCTION

In the last 100 years, while industrialized fishing has reduced fish populations on a global scale (Pauly and MacClean 2003), vast effort has gone into explaining the variable and uncertain replenishment of marine populations (Sissenwine 1984). Highly variable recruitment dynamics (Roughgarden et al. 1988), spatially structured populations, high fecundity, and almost total but variable mortality of the immature (larvae and juveniles) stages (Bailey and Houde 1989) are typical of marine organisms with complex life cycles. These characteristics are common among marine invertebrates and fishes (Booth and Brosnan 1995), find analogues in the complex life cycles of macroalgae (Reed et al. 2004), and challenge our ability to understand the processes governing the stability of marine populations (Legget and Deblois 1994).

The challenges facing marine larvae are great. Planktotrophic larvae must find nourishment and avoid predators in the ocean, then locate and colonize suitable benthos for metamorphosis into the next generation of juveniles. These requirements impose substantial risks for larval (Bailey and Houde 1989) and juvenile (Sogard 1997) stages that interact to compound uncertainties surrounding individual survival to maturity. While large-scale external factors (weather, ocean productivity, water circulation, and climate) have long been recognized as broad correlates of larval success (Bakun et al. 1982, Sinclair 1988, Cushing 1996), the recent application of individual-based models in ecology to deal with highly variable and nonlinear phenomena (Judson 1994) has improved our ability to examine the ways in which factors interact at local scales and with intrinsic attributes of the propagules to affect larval mortality.

While some sources of mortality (e.g., extreme temperatures) kill randomly, other sources (e.g., starvation, predation) produce effects that are spread unequally among individuals within a cohort. A very
Untangling the effects of size and growth rate on larval mortality is commonly done by longitudinal resampling of the same cohort (individuals spawned during a narrow temporal window) to compare the characteristics of survivors and non-survivors in natural populations (Sogard 1997). For teleosts, this analysis is facilitated by the calcareous otoliths found in the heads of all bony fishes. These inclusions, used in balance and hearing, grow by daily accretions that deposit internal microstructures (Campana and Neilson 1985). When otolith and somatic growth are coupled, otoliths encode a permanent ontogenetic record of size-at-age that can be recovered and converted to prior sizes and/or historical growth rates by simple back calculations (Thorrold and Hare 2002), providing valuable insights to the seldom observed larval stages.

Otolith records have shown correlations between larval growth rates and cohort strength at settlement (Searcy and Sponaugle 2001, Bergenis et al. 2002, Wilson and Meekan 2002), correlations between larval growth rates and juvenile growth rates (Vigliola and Meekan 2002), and correlations between juvenile growth rates and post-settlement mortality (Searcy and Sponaugle 2001, Vigliola and Meekan 2002, McCormick and Hoey 2004). Similar effects have been observed in temperate reef fishes (e.g., Shima and Findlay 2002, Ravenotos and Macpherson 2005). However, it remains unclear if size-selective mortality during larval dispersal and recruitment is a consistent feature affecting most cohorts within a fish population. Furthermore, it is unclear whether selective mortality arises from density effects upon competitive interactions (Webster 2004), from food and temperature effects upon larval growth (Meekan et al. 2003), from maternal effects such as egg size and provisioning (Jones and McCormick 2002), or from genotypes that influence physiological performance (Planes and Romans 2004). These issues are not inconsequential; consistent selective mortality acting upon heritable characters may generate long lasting effects on both the demographic and the genetic compositions of surviving populations (Conover and Munch 2002, Heino and Godo 2002).

Vigliola and Meekan (2002) demonstrated positive selection in one cohort of the coral reef damselfish, Neopomacentrus filamentosus, with respect to larval size. Here, we build on these results to further extend our understanding of the consequences of size-selective mortality. First, we explore whether size-selection is a consistent feature of post-settlement mortality in this species through the examination of multiple larval cohorts with respect to both size and growth rate. Second, we examine the role of competitive interactions, environmental factors, and potential maternal effects through correlations between fish densities, growth rates and size at hatching with observed selective mortality. Finally, because of the extreme nature of the observed selective mortality, we explore the genetic consequences within the cohort that experienced the strongest natural selection to determine if and how strong size-selection can alter the genetic composition of a cohort.

**METHODS**

**Study species**

Neopomacentrus filamentosus is a small (<8 cm) damselfish that is widely distributed in the Indo-Australian Archipelago (Allen 1991). Adults of this planktivorous fish are abundant in the Dampier Archipelago, Western Australia (20°30' S, 116°40' E), where they occupy coral heads in shallow (4–10 m) water. Males guard nests of benthic eggs during Austral summer months (October to March). After four to five days of development, eggs hatch into pelagic larvae that spend an average of 18 days in the plankton (Vigliola and Meekan 2002). Settlement occurs on new moons and young recruits initially school near the bottom of large coral colonies. As they grow, recruits move up in the water column to join older juveniles and adults schooling above the coral heads. This ontogenetic habitat shift occurs at 20–25 mm, which represents one to two months of growth after settlement (Vigliola and Meekan 2002).

**Collection methods**

Light traps (Doherty 1987) were used to collect live presettlement reef fishes from the Dampier Archipelago over two summers. Ten traps (see Fig. 1 in Meekan et al. [2001] for trap design) were moored near the surface and cleared daily for two weeks around consecutive new moons from November 1998 to February 1999 and October 1999 to February 2000 (Vigliola and Meekan 2002). On retrieval, the content of each trap was preserved immediately in 98% methanol, which is harmless to otoliths, but essential to preserve the DNA of the incoming cohorts.

Although our sampling encompassed two entire spawning seasons for Neopomacentrus filamentosus, and eight potential lunar cohorts, presettlement fish were only abundant in catches from four months...
 Trait

A. Hansen and A. Retzel, in this species has been validated by a tag-recapture study. Nearest 1 light with immersion oil at 1000 magnification. Daily increments within each otolith were measured to the nearest 1 \( \mu m \) along the longest axis of each section using an image analysis system. Daily deposition of increments in this species has been validated by a tag-recapture study (A. Hansen and A. Retzel, unpublished data). However, we assumed that the increment closest to the core of each otolith was formed on the day of hatching (e.g., Campagna and Neilson 1985).

The SL of each fish on every day between hatching and capture was back calculated from its otolith using the linear biological intercept method (Campagna 1990), which assumes that otoliths grow in proportion to body length (Vigliola et al. 2000). Evidence for this assumption is provided by the strong linear relationship between otolith radius and fish length for this species (Vigliola and Meekan 2002). The biological intercept was set at hatching where mean otolith radius and fish size were estimated at 9.4 \( \mu m \) and 2.3 mm SL, respectively (Vigliola and Meekan 2002).

Two-factor (cohort and time) nonlinear mixed-effects (NLME) models (Pinheiro and Bates 2000) were used to compare the estimates of growth trajectories back calculated from the otoliths of fish surviving to different ages. These models have recently been developed to examine longitudinal, unbalanced, and autocorrelated data sets, characteristics that are typical of data extracted from fish otoliths (Chambers and Miller 1995). To investigate differences in growth rates before and after settlement (\( t_s \)), we fitted a parabolic model (after Vigliola 1998) to larvae,

\[ L(t) = L_0 + K_1 t^2 \quad \text{for } t \leq t_s \]  
and a standard von Bertalanffy function (Quinn and Deriso 1999) to juveniles,

\[ L(t) = L_{\text{asym}} \left[ 1 - e^{-K_2 (t-t_0)} \right] \quad \text{for } t > t_s \]

where \( L(t) \) is standard length (mm) at age \( t \) (in days), \( L_0 \) is length at hatching, \( K_1 \) is growth acceleration (mm/d\(^2\)), \( L_{\text{asym}} \) is average length at the end of asymptotic growth period, \( K_2 \) is instantaneous growth (percent per day), \( t_0 \) is the predicted age when individuals have a length of zero, and \( t_s \) is the age at transition between larval and juvenile stages such that

\[ L_s = L_0 + K_1 t_s^2 = L_{\text{asym}} \left[ 1 - e^{-K_2 (t_s-t_0)} \right] \]

for the two curves to intersect at the point \((L_s, t_s)\). Larval and juvenile growth parameters were estimated setting \( t_s = 12 \text{ d} \) (the age of the youngest settler) as first approximation in Eqs. 1 and 2. The exact point of intersection between the two growth curves was then determined a posteriori by solving Eq. 3 using numerical optimisation (Nocedal and Wright 1999). The parabolic growth model (Vigliola 1998) is based on the assumption of proportionality between somatic growth and fish age. Theoretical justification of this model is based on feeding behavior during the planktonic larval stage. After hatching fish larvae need to learn which prey they can successfully capture and how to attack them, so that the rate of feeding success and hence growth rate will increase with age (Kentouri and Divanach 1982).

Correlation analyses were used to examine the strength and timing of size-selective mortality in relation to body size, growth rates, and fish densities. The intensity of natural selection (SI) was measured as the difference in trait means before and after selection standardized by the standard deviation before selection (Brodie et al. 1995)

\[ SI = \frac{(\text{Trait})_{\text{after}} - (\text{Trait})_{\text{before}}}{SD_{\text{before}}} \]
Standard length at 12 d was the trait on which the strength of selection was estimated.

**Molecular analysis**

Otoliths revealed the strongest phenotypic selection in the November 1998 cohort. Consequently, this cohort was selected for molecular analysis. DNA from individuals used in the otolith analyses was extracted from muscle tissue using a 10% Chelex (BioRad Laboratories, Hercules, California, USA) solution (Walsh et al. 1991). A fragment of the putative mitochondrial control region was then amplified using primers CR-A and CR-E (Lee et al. 1995). Hotstart thermocycling parameters using Amplitaq DNA polymerase consisted of an initial hold at 80°C followed by 39 cycles of 94°C, 30 s, 50°C, 30 s, 72°C, 45 s, and a final 5-min extension. PCR products were visualized on a 1% agarose gel and enzymatically prepared for sequencing through incubation with ExonucleaseI (5 units; Amersham Biosciences Corporation, Arlington Heights, Illinois, USA) and Shrimp Alkaline Phosphatase (0.5 units; Amersham) at 37°C for 30 min followed by denaturation at 80°C for 15 min. Cycle sequencing was done with ABI BigDye 3.0 chemistry (Applied Biosystems, Foster City, California, USA) using the above PCR primers. Sequencing products were analyzed on an ABI 377 automated sequencer (Applied Biosystems); forward and reverse sequences were proofread and compiled in Sequencher 4.0 (GeneCodes Corporation, Ann Arbor, Michigan, USA). Alignments were performed using ClustalW (Thompson et al. 1994) then modified by eye. MacClade 4.05 (Maddison and Maddison 2002) was used to format the sequences and merge identical haplotypes for use by Arlequin (Schneider et al. 2000).

Genetic differentiation resulting from size-selective mortality was examined by treating settlers and recruits as distinct temporal populations, and comparing their genetic structure using $\theta_{ST}$ as calculated in Arlequin (Schneider et al. 2000) with statistical significance determined through 3000 random permutations. To explore the effects of the different sizes of the settler and recruit samples, 68 fish (the size of the recruit sample) were selected randomly from the 91 settlers 500 times and the $\theta_{ST}$ values between groups calculated in Arlequin. To further explore the effects of genetic drift caused by high mortality, 10, 20, 30, 40, 50, 60, or 70 fish were removed at random from the settler sample, representing up to 77% mortality. This sample of remaining “survivors” was then compared to the original settler sample to determine the range of $\theta_{ST}$ values resulting from high, but random, mortality. For each removal class, a distribution was created from 1000 iterations of random removals to determine the probability of genetic drift producing the observed results.

The relationship between genetic structure and size-selective mortality was further explored by manipulating selection intensity a posteriori such that the settler and recruit samples were made more similar or more different in (1) size at hatching, (2) size at settlement (12 d), and (3) larval growth. Size selective mortality was imposed to widen the phenotypic difference between temporal samples by removing the 10, 20, or 30 smallest/slowest fish from the recruit sample and/or removing the 10, 20, or 30 largest/fastest fish from the settler sample. Size selective mortality was then imposed to minimize the phenotypic differences between samples through the opposite treatments: i.e., removing the 10, 20, or 30 smallest/slowest fish from the settler sample, and/or removing the 10, 20, or 30 of the largest/fastest fish from the recruit sample. To control for potential sample size effects, the same number of randomly selected individuals was removed simultaneously from the non-treatment group. In total, 12 a posteriori experimental treatments were performed. Because Arlequin only indicates $\theta_{ST}$ values significantly larger than zero, a simulation was performed for each trait in which 10, 20, or 30 individuals were removed at random from both data sets. The resulting $\theta_{ST}$ values from 1000 iterations formed a distribution from which values significantly larger than the $\theta_{ST}$ obtained from natural selection alone could be determined and represent a further test for genetic drift within the cohort.

**RESULTS**

**Light trap catches**

A total of 694 Neopomacentrus filamentosus larvae were collected during the 1998/1999 and 1999/2000 spawning seasons with most fish collected from late spring to early summer (October to December) in both years. The average size of potential settlers was 11.16 mm SL (SD = 0.89 mm, range = 7.70–13.80 mm) and the average age of a representative sample of 345 fish was 18 days (SD = 3 d, range = 12–27 d). Larval supply was highly variable among cohorts with total catches ranging between 0 and 381 potential settlers (Appendix A: Table A1).

**Comparison of larval growth trajectories among cohorts**

Parabolic models provided equally good fits to the first 12 days of larval growth for all four cohorts, whether back calculated from settlers or three-month recruits (Fig. 1). Comparisons of the coefficients of the NLME model indicated that cohorts at three months after settlement were composed of fish that were significantly bigger at hatching (Fig. 2A; two-way ANOVA, $F_{1,624} > 12.79, P < 0.001$), had grown significantly faster as larvae (Fig. 2B; F tests for linear combinations, $F_{1,7560} > 19.23, P < 0.001$), and were larger at 12 d (Fig. 2C; two-way ANOVA, $F_{1,624} > 20.76, P < 0.001$) than their initial composition when sampled at settlement, albeit with significant interactions for starting size and growth rate (Appendix B: Table B1). Qualitatively, all four cohorts were modified in the same way, with three months of post-settlement mortality selectively eliminating individuals with the smallest starting sizes and the slowest growth rates.
Effect size varied among cohorts. For example, settlers from the December 1999 cohort had smaller hatch sizes than settlers from the previous month (Fig. 2A; two-way ANOVA, $F_{1,624} = 14.37, P < 0.001$) but no differences in hatch sizes were detected between the recruit samples from these same cohorts (two-way ANOVA, $F_{1,624} = 1.75, P = 0.18$). Larval growth rates from the same cohorts showed the opposite pattern. Settlers from the December 1999 cohort recorded faster growth than settlers from the November 1999 cohort (tests for linear combinations, $F_{1,7569} = 23.42, P < 0.0001$), but this difference was erased by the time both cohorts were three months old (Fig. 2B, tests for linear combinations, $F_{1,7569} = 0.91, P = 0.34$). In contrast, settlers from the December 1999 cohort also had faster growth than settlers from the November 1998 cohort ($F_{1,7569} = 6.36, P < 0.05$) but recruit samples from the same cohorts showed a bigger difference in the opposite direction ($F_{1,7569} = 56.92, P < 0.001$).

While some of these exact comparisons involve tiny differences in the model parameters that may arise from chance or parameter estimation, the overall pattern is one of consistent modification of cohort composition at, or after, settlement from selective mortality against small and/or slow-growing individuals. The large effect size observed in all three model parameters for the November 1998 cohort (Fig. 2A–C) compared with all three cohorts from 1999 (except that for hatch size in the December 1999 cohort) shows that the strength of size selection is not constant.

**Comparison of larval growth trajectories within cohorts**

Two cohorts (November 1998, November 1999) were resampled more than once yielding information about the evolution of the patterns described above (Fig. 2D–F). As before, interaction terms were significant for both parameters of the parabolic larval growth model (Appendix B: Table B1) fitted to the crossed factors of cohort (November 1998, November 1999) and age (settlers, recruits sampled at one, two, and three months after settlement). Comparisons of model coefficients for the November 1998 cohort showed no modification of hatch sizes between settlers and recruits at one month (two-way ANOVA, $F_{1,681} = 1.09, P > 0.29$), but progressive modification of this cohort over the next two intervals (Fig. 2D). The hatch sizes of recruits two months after settlement were larger than those of recruits at one month (two-way ANOVA, $F_{1,681} < 1.09, P > 0.29$), but progressive modification of this cohort over the next two intervals (Fig. 2D). The hatch sizes of recruits two months after settlement were larger than those of recruits at one month (two-way ANOVA, $F_{1,681} > 32.09, P < 0.001$), and the hatch sizes of recruits three months after settlement were larger than those of recruits at two months (two-way ANOVA, $F_{1,681} > 4.68, P < 0.04$). In contrast, hatch sizes in the November 1999 cohort did not differ between settlers and recruits up to two months after settlement (two-way ANOVA, $F_{1,681} > 2.96, P > 0.08$ for all comparisons). However, hatch sizes in the oldest recruit sample (three months) were significantly larger than in earlier samples of this cohort (two-way ANOVA, $F_{1,681} > 15.24, P < 0.001$).
Average larval growth rates (Fig. 2E) increased with each resampling of the November 1998 cohort (F tests for linear combinations, $F_{1,8253} > 4.06$, $P < 0.05$). In the November 1999 cohort, larval growth rates in the settler sample were slower than those in the first two collections of recruits (F tests for linear combinations, $F_{1,8253} > 10.57$, $P < 0.01$), but did not differ between the latter collections (F tests for linear combinations, $F_{1,8253} < 0.35$, $P > 0.55$). Both were slower, however, than larval growth rates recorded in recruits collected three months after settlement (F tests for linear combinations, $F_{1,8253} > 40.65$, $P < 0.001$).

Quantitative comparisons of larval size at 12 d produced the same outcomes as those described for growth rates (cf. Fig. 2E, F) and size-at-age plots confirm that these differences can be traced back to hatch sizes (Fig. 3A, B). This is clearest in trajectories from the November 1998 cohort, where there was strong selection on recruits between one and two months after settlement.
Comparison of juvenile growth trajectories within cohorts

Interaction terms were significant for both parameters of the von Bertalanffy growth model (Appendix B: Table B1) fitted to the crossed factors of cohort (November 1998, November 1999) and age (recruits at one, two, and three months after settlement), showing that recruits from both cohorts had different back-calculated growth trajectories when sampled at different times after settlement (Fig. 3C, D). The parameters of this model indicated that these differences were largely the propagation of differences present during larval life. A significant interaction term for the intercept $t_0$ of the model was consistent with changing mean size at settlement, albeit with differences between the two cohorts. The stronger selection evident on the November 1998 cohort was reflected in greater changes in post-settlement growth rate (Fig. 4A) and consistent elevation of predicted final size at the end of the study (Fig. 4B). In contrast, the November 1999 cohort showed less change in both parameters with significant change in $L_{\text{asym}}$ detected only at the final sampling. For the six samples for which we could estimate both a parabolic larval and a von Bertalanffy juvenile growth models (November cohorts at one, two, and three months after settlement), numeric optimization of Eq. 3 revealed an exact age at transition close to 12 days ($12.9 \pm 0.7$ d, mean $\pm$ SD; range 12–14 d, $n = 6$).

**FIG. 3.** Comparison of growth models fitted by nonlinear mixed-effects (NLME) models for two cohorts (November 1998 and 1999) of *Neopomacentrus filamentosus* sampled at four times (0, 1, 2, and 3 months after settlement). (A, B) Presettlement parabolic and (C, D) post-settlement Von Bertalanffy growth models for (A, C) November 1998 and (B, D) November 1999. Error bars indicate 95% CI. Means are only shown every 3, 6, or 10 days for clarity.

**FIG. 4.** Comparison of post-settlement growth parameters of two cohorts (November 1998 and 1999) of *Neopomacentrus filamentosus* sampled at three times (1, 2, and 3 months after settlement). (A) Von Bertalanffy relative growth rates $K_2$ and (B) asymptotic lengths at the end of the study $L_{\text{asym}}$ (based on standard length, SL) are compared by nonlinear mixed-effects (NLME) models. Different lowercase letters indicate significant differences at $P < 0.05$. Error bars indicate 95% CI.
Variability in the timing and strength of size-selective mortality

Analysis using NLME models of growth trajectories showed that size-selective mortality occurred in all cohorts, with smaller individuals disappearing from back calculations as cohorts were sampled at longer intervals since settlement (Fig. 5). Significant cohort-by-age interactions in all analyses (Appendix B: Table B1) indicated that the strength of size selection varied both among cohorts and sampling intervals. Selection intensity (SI) was strongest in the November 1998 cohort (SI = 2), lowest in the December 1999 cohort (SI = 0.6), and intermediate in the other cohorts. Although selection intensity was not correlated with planktonic growth rate \((r = -0.16, P = 0.58)\), it was positively correlated with the magnitude of light trap catches \((r = 0.65, P = 0.17)\) implying a link with the density of settlers, albeit not one arising immediately after settlement. In the November 1998 cohort, the strongest episode of size-selective mortality occurred between one and two months after settlement, at an average size of 23 mm SL, while a similar episode in the November 1999 cohort occurred a month later at an average size of 29 mm SL.

Genetic selection

A total of 365–533 bp of mitochondrial control region DNA sequence was obtained for 91 settlers and 68 recruits collected three months after settlement from the November 1998 cohort. These yielded a total 145 unique haplotypes (82 and 67 for settler and recruits, respectively, with four shared between samples). Genetic structure among the two samples was slight, but significant \((\phi_{ST} = 0.016, P = 0.041)\), indicating temporal genetic structure. In contrast to differences resulting from natural selection, 500 replicate trials comparing a random assortment of 68 settlers to the original 91 settlers resulted in no \(\phi_{ST}\) values statistically different from zero. Removing 10, 20, 30, 40, 50, and 60 larvae at random from the settlers resulted in no significant \(\phi_{ST}\) values when compared to the original settler sample. Only when 70 (77%) of the larvae were randomly removed were two significant \(\phi_{ST}\) values obtained, indicating \(P = 0.004\) of getting a significant \(\phi_{ST}\) by genetic drift.

Augmenting selective pressure by removing the smallest and slowest growing individuals from the recruit sample increased the phenotypic difference among settler and recruit samples as well as the magnitude and significance of \(\phi_{ST}\) values for both size at day 12 and growth rate, but not size at hatching where additional selective pressures had equivocal effects (Fig. 6A–C; Appendices C–E: Tables C1, D1, and E1). Furthermore, these increased \(\phi_{ST}\) values were larger than \(\phi_{ST} = 0.016\) resulting from natural selection alone as determined by the distribution resulting from 1000 random permutations where 10, 20, or 30 individuals were removed at random from both samples. Selective removals of larger, faster growing fish from the settler sample also increased the phenotypic difference between settler and recruits, yielding larger and more significant \(\phi_{ST}\) values that were significantly larger than \(\phi_{ST} = 0.016\) resulting from selection alone (Fig. 6D–F; Appendices C–E: Tables C1, D1, and E1).
Removing the smallest and slowest growing individuals from the settler sample decreased the phenotypic difference among settler and recruit samples, but $\phi_{ST}$ values were still significantly larger than zero for most comparisons. However, with the exception of the treatment that removed 10 of the individuals that were smallest at hatching, none of these values were significantly different than $\phi_{ST} = 0.016$ resulting from selection alone (Fig. 6A–C; Appendices C–E: Tables C1, D1, and E1). Similarly, the removal of the larger, faster growing fish from the recruit sample also decreased the phenotypic difference between settler and recruits. In this case, $\phi_{ST}$ value decreased in magnitude and/or became nonsignificant (Fig. 6D–F; Appendices C–E: Tables C1, D1, and E1).

**DISCUSSION**

**The settlement transition and predation risk**

Settlement is a critical transition in the life history of all benthic marine organisms with complex life cycles and dispersive larvae denoting the shift from planktonic to demersal lifestyles (Roughgarden et al. 1988). For most species, it is an abrupt transition requiring the rapid acquisition of new adaptations and the loss of old ones with the most extreme manifestation being the radical metamorphoses of invertebrates. Conversion to sessile life is an irrevocable choice with profound consequences for subsequent reproduction (Moran and Emlet 2001, Phillips 2002) because it is not unusual for settlement to be followed immediately by very high, even catastrophic,
mortality (Caffey 1985, Gosselin and Qian 1997a, b). Metamorphic invertebrates with small body sizes and little time to grow defensive adaptations are vulnerable to a wide range of mortality sources, including physiological stresses (Schmidt et al. 2000), predation (Hunt and Mullineaux 2002), and even removal by indiscriminate grazing (Sammaroo 1980, Barnes 1999).

While the metamorphosis of benthic fish larvae may be less spectacular than those of invertebrates, settlement is still a period of rapid physiological, behavioral, and ecological change (Kaufman et al. 1992, McCormick and Makey 1997). Many species adopt sedentary lives after settlement (Sale 1978), so there are parallels with sessile forms and habitat selection is no less evident or important (e.g., Wellington 1992). Research on tropical and temperate reef fishes has highlighted the critical importance of predation during (Doherty et al. 2004) and following (Webster 2002) settlement, when naïve metamorphs are still adapting to their benthic environment (McCormick et al. 2002). Consequently, settlement can be a time of intense natural selection upon traits affecting individual fitness.

**Size selective mortality in Neopomacentrus filamentosus**

*Neopomacentrus filamentosus* is a planktivorous damselfish that settles gregariously with conspecifics. At the neighborhood level, prey clumping can attract predators (Hixon 1991) so that per capita mortality of prey in dense clumps is higher than those with more uniform distributions even when total abundance is higher in the latter (Brunton and Booth 2003). While schooling is a defense against predation, it can also reduce individual fitness because bony fish have plastic growth rates (van Rooij et al. 1995) and social interactions in fish schools of mixed age often results in growth suppression of subordinate individuals, especially at higher densities (Doherty 1982, Booth 1995, Webster 2004).

The key ecological result from our study was that post-settlement mortality in four lunar cohorts of *Neopomacentrus filamentosus* was consistently selective against the smallest and slowest-growing individuals in each cohort during the first three months of benthic life. The timing and intensity of size selection varied between the two cohorts that were resampled more than once (November 1998, November 1999) and the strongest size selection was observed in the cohort with highest initial density. These results are consistent with density-dependent mortality selecting against individual variation.

*Neopomacentrus filamentosus* collected from the water column before settlement showed substantial phenotypic variation. Their ages ranged from 12 to 27 days and their sizes from 7.7 to 13.8 mm. Despite this variation, the majority of live individuals had the silver pigmentation typical of metamorphs suggesting that all were developmentally competent to settle. Since we did not measure the biochemistry of these presettlement fish, we cannot exclude linked selection for condition factors other than larval growth (Kerrigan 1996). While this might have been an issue if we had not detected any phenotypic selection, there was a clear and significant disadvantage to small size and/or slow growth rate in newly settled individuals.

The current paradigm for recruiting reef fish is one of greatest mortality immediately after settlement (Hixon and Webster 2002), whereas we detected size selection between one and two months after settlement that coincided with an ontogenetic shift in microhabitat. Settlers shelter initially around the bases of coral “bommies” but later migrate up to the tops of these structures where they join mixed age groups foraging higher in the water column. This is when competitive interactions with older conspecifics have greatest potential for facilitating selective predation (Brunton and Booth 2003, Webster 2004). The micro-habitat segregation evident at settlement may be a strategy to reduce this conflict at the time when new colonists are most vulnerable. Finn and Kingsford (1996) describe similar habitat segregation and ontogenetic movement during the phased colonization of coral reefs by schooling cardinalfish, which are preferred prey of many piscivores (Webster and Almany 2002).

A lack of detectable size selection in the first month after settlement must not be confused with low mortality. The basis of our test was longitudinal resampling of a cohort to recover the characteristics of survivors (Sogard 1997) and was designed explicitly to capture evidence of size selection. As such, it is based on relative abundance of growth phenotypes and is silent about mortality that does not alter size structure. Doherty et al. (2004) found instantaneous mortality of 60% during settlement of a tropical surgeonfish that was not size selective and would be missed by this analysis. This may be typical of mortality that is brief and/or dependent upon random encounter between predator and prey, unlike the selective mortality observed in dense clumps of size-structured prey (Brunton and Booth 2003).

Sogard (1997) points out that high mortality is one of three necessary conditions for the detection of size selection because low non-random mortality may not detectably modify the size distribution of survivors. This could explain the different pattern of size selection seen in the November 1999 cohort of *Neopomacentrus filamentosus*, which was mostly expressed in fish between two and three months of age. While this cohort settled at 20% of the density of the November 1998 cohort, it was composed of smaller fish with slower growth histories before settlement, which is indicative of cooler water temperatures (Meekan et al. 2003). Unfortunately, we do not have independent environmental data, but a cooler year resulting in slower pre- and post-settlement growth could explain why the ontogenetic migration to higher positions in the water column was delayed by a month until juveniles grew to the size to risk this transition.

**Temporal genetic selection**

In the cohort of highest density (November 1998), size-selective post-settlement mortality resulted in the
generation of subtle but significant genetic structure between settlers and recruits after three months. Previous studies have reported genetic differentiation among samples of recruits (Johnson and Black 1982, Li and Hedgecock 1998, Bernardi et al. 2001) as well as between recruits and adults (e.g., Moberg and Burton 2000). These differences have been variously attributed to larvae originating from different source populations (e.g., Bernardi et al. 2001), unequal parental contribution to larval pools (Li and Hedgecock 1998), and post-settlement selection (Johnson and Black 1982, Moberg and Burton 2000). In our study, we can exclude unequal parental contribution as a possible explanation because >90% of settlers and 98% of recruits had unique haplotypes. While the cohort may have contained larvae from different sources (i.e., a Wahlund effect), the genetic changes we observed must have come either from natural selection preferring individuals with particular genotypes or from genetic drift.

A strong reduction of cohort size could alter the genetic structure of a cohort between two samples by the action of neutral genetic drift. Random sampling of the 91 settlers to produce different groups of 68 settlers for the initial comparison set did not yield significantly different \(\phi_{ST}\) values in 500 trials, suggesting that the observed pattern was not an artefact of reduced sample size. Furthermore, comparing the settler sample to a modified settler sample with 77% of individuals removed at random only produced two significant \(\phi_{ST}\) values in 1000 iterations, indicating a 0.4% probability that genetic drift alone could produce significant genetic structure, assuming similar levels of selective mortality.

In contrast, the link between size selection and genetic differentiation was corroborated by the positive relationship between the intensity of the size selection imposed a posteriori and the strength of the resulting genetic structure. Increasing the phenotypic difference between settlers and recruits, whether achieved by removing smaller/slower growing fish from the recruit sample or by removing larger/faster growing fish from the settler sample, was accompanied by a sharp and significant increase in \(\phi_{ST}\) values for both sizes at day 12 and growth rate, though not for size at hatching. In contrast, the reverse manipulations of the samples, removing larger/faster growing individuals from the recruit sample or small/slower growing fish from the settler sample, decreased the phenotypic difference between temporal samples and resulted in the loss of detectable genetic structure measured by size at day 12 and/or growth rate; with equivocal results for size at hatching. In nearly all cases \(\phi_{ST}\) values resulting from augmented selection significantly exceeded those resulting from random removals. These results combined with 77% mortality only having a 0.4% chance of producing significant genetic structure at random downplay the role of genetic drift in producing the observed results. Instead, it appears that the temporal genetic structure among samples was a direct function of selective mortality.

Temporal genetic selection in marine population is not uncommon (Neigel 1997). Examples include selection of allozyme alleles along salinity (Burton and Feldman 1983, Hilbish and Koehn 1985) and temperature (Rand et al. 2002) gradients. More recently, Planes and Romans (2004) detected selection against individuals in a wild cohort of fish that carried an allele linked with lower growth of fish in a laboratory experiment. Our study provides direct evidence of selection against slow growing fish but, in contrast to these previous studies, we detected this effect from comparisons of haplotypes in a non-coding region of the mitochondrial genome that is assumed to be selectively neutral.

Because the mitochondrial genome is inherited as a unit (Randi 2000), the control region is linked to other coding regions that could be under selection. As the mitochondria produces 90% of the energy for the cell, changes in mitochondrial efficiency will alter energy available for growth, creating different phenotypes to be acted upon by selection. Although the bulk of metabolic genes in the mitochondria are produced in the nucleus, mitochondrial performance can be affected by genetic variation at both nuclear and mitochondrial loci (Rawson and Burton 2002), suggesting a proximal linkage for selection of mtDNA variation and the phenotypic traits measured in this study.

Although \(\phi_{ST}\) responded to both size at day 12 and growth rate, results were equivocal for size at hatching. There are two potential explanations for this result. First, it may be that the back calculations of larval size become more error prone the further back they are extended, resulting in less reliable estimates. However, otolith analysis suggests otherwise, with consistent selective patterns obtained for all three traits. Hence, a more plausible explanation is that size at hatching is influenced by maternal effects (McCormick 1998, 1999, Jones and McCormick 2002). Therefore, this character may be more reflective of the reproductive condition of the female than the physiological abilities of the larvae, resulting in the absence of any discernable pattern with respect to genetic selection.

**Selection and the evolution of life history traits**

Life history strategies represent an ensemble of coadapted traits that collectively determine individual fitness through testing by natural selection. If, as our results indicate, post-settlement mortality is not always a chance event and faster growing individuals can have better survival after settlement, why has directional selection not purged slow growing phenotypes from these populations? The most plausible explanation involves balancing selection operating in a heterogeneous selection landscape.

There are well-documented tradeoffs among demographic traits such as growth, reproduction, and longevity (Stearns 1976). The reproduction of many marine species is characterized by high fecundity, small eggs with minimal provisioning, and planktotrophic
larvae, which is a “risk-spreading” tactic selected by unpredictable conditions in the larval environment (Einum and Fleming 2000a, b). Total mortality is extremely high in the early life histories of such organisms and small variations in the larval environment (Houde 1987) can produce large differences in recruitment between years (Cushing 1975). When all other things are equal, maternal provisioning can enhance larval survival (Einum and Fleming 1999, Marshall and Keough 2004) but population genetics studies have shown that there is a huge variance in reproductive success among spawners that may result in genetic sweepstakes (Hedgecock 1994). While some cohorts have narrower genetic variation than their source populations, indicating very small effective population size (Turner et al. 2002), others do not, indicating that there can be many winners of the recruitment sweepstakes (Flowers et al. 2002). Some species invest in propagules of different size and quality to combine different “bet-hedging” tactics (Einum and Fleming 2004). In an unpredictable adaptive landscape, directional selection may be limited and environmental variability may reward many phenotypes sufficiently to prevent their elimination.

Hixon and Carr (1997) showed that the most intense predation on juvenile reef fishes occurs when both resident and transient piscivores target the same patch of prey. Since mobile optimal foragers should concentrate on patches offering greatest reward within their home ranges, it follows that predation pressure will not be uniform (Brunton and Booth 2003). Slow-growing individuals in a shoaling species like Neopomacentrus filamentosus face double jeopardy when they join a high-density school of conspecifics because of the neighborhood attraction of predators and the density-dependent interactions with conspecifics that can result in growth stunting of subordinate individuals. Meanwhile, fast-growing individuals should gain some relief from predation through the presence of more vulnerable individuals in the same school. These are benefits that cannot be easily predicted during the process of settlement and will depend upon the initial composition and densities of groups within a local neighborhood. For some species, predation can also be reduced when easier prey, like cardinalfishes, settle abundantly at the same time (Webster and Almany 2002). While fast growth is the best protection against predation, there is evidence that it comes with other costs (see review by Mangel and Stamps 2001). For example, oversize, fast-growing individuals can have reduced resistance to pathogens or parasites (e.g., Desclaux et al. 2004). These and other tradeoffs will favor the retention of phenotypic plasticity and genetic diversity at the population level.

Applications to marine fisheries

Our observation of temporal genetic change in a reef fish cohort that can be linked to natural size-based selection provides two cautionary lessons. First, our discovery of linked selection on a non-neutral molecular marker, the control region of mtDNA, has significance for studies that use this marker to discriminate genetic structure arising from reproductive isolation (Ferguson and Danzmann 1998). While this becomes less of an issue when sampling mixed year classes, there is potential for spatial and/or temporal differences in the intensity of size-based selection to compromise inferences about genetic stock structure in short-lived species, samples with very different year class composition, and spatial comparisons of recruitment. The use of multiple molecular markers will avoid this issue, although this is rarely done for reasons of extra cost. Under these circumstances, otolith back calculations can provide a useful cross validation that the samples are not likely to have these problems.

Beyond adding another example of non-neutral evolution of mtDNA (Ballard and Whitlock 2004), our results have implications for the management of marine fisheries if we assume that demographic variety is the evolved solution to unpredictable and variable environments. In contrast, modern fisheries are currently imposing directional selection on fish stocks through intense and consistent size-selective harvesting (Ricker 1981, Rijnsdorp 1993, Law 2000) that has the potential to eliminate the genotypes for fast growth (Heino and Godo 2002). Conover and Munch (2002) demonstrated measurable and heritable shifts in size and growth in Atlantic silversides, Menidia menidia, after just four generations of selection.

In a recent terrestrial example, Coltman et al. (2003) showed that targeted shooting of mountain sheep with the largest horns by trophy hunters had purged the population of these genotypes in a few generations, so that the original phenotypic range could not be recovered even with complete protection. While these closed remnant populations are the opposite of the open extensive populations typical of most marine organisms, there is growing evidence that modern fishing methods, which inherently target large individuals, are driving genetic structure towards genotypes for short-lived, early-maturing individuals (Heino and Godo 2002). Since the individuals removed from the gene pool (genotypes for long-lived, late-maturing individuals with high fecundity) are also those that provide most compensatory reserve in these populations (Rose et al. 2001), there is an urgent need to reconsider management regimes that encourage such selective depletion.

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GENETIC IDENTITY DRIVES FISH MORTALITY

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**APPENDIX A**

Sampling dates and sample sizes (Ecological Archives E088-079-A1).

**APPENDIX B**

Results of nonlinear mixed effects (NLME) model analyses of growth back-calculated from otoliths of four cohorts of *Neopomacentrus filamentosus* (Ecological Archives E088-079-A2).

**APPENDIX C**

Difference in size and genetic structure between settlers and recruits based on size at hatching for both natural and augmented selection regimes (Ecological Archives E088-079-A3).

**APPENDIX D**

Difference in size and genetic structure between settlers and recruits based on size at day 12 for both natural and augmented selection regimes (Ecological Archives E088-079-A4).

**APPENDIX E**

Difference in size and genetic structure between settlers and recruits based on growth rate for both natural and augmented selection regimes (Ecological Archives E088-079-A5).