

# Geochemistry, Geophysics, Geosystems

## RESEARCH ARTICLE

10.1029/2020GC009481

†Deceased.

### Key Points:

- Unique stable carbon isotope of suspended particulate organic carbon is indicative of unique carbon fixation pathway by vent biomes
- We estimated first dissolved organic nitrogen for hydrothermal vent fluids and observed enhanced nitrogen fixation and/or denitrification
- Total dark organic carbon production within the plume matches thermodynamic prediction based on available, reducing chemical substances

### Supporting Information:

Supporting Information may be found in the online version of this article.

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### Citation:

Lin, H.-T., Butterfield, D. A., Baker, E. T., Resing, J. A., Huber, J. A., & Cowen, J. P. (2021). Organic biogeochemistry in West Mata, NE Lau hydrothermal vent fields. *Geochemistry, Geophysics, Geosystems*, 22, e2020GC009481. <https://doi.org/10.1029/2020GC009481>

Received 15 OCT 2020

Accepted 7 FEB 2021

## Organic Biogeochemistry in West Mata, NE Lau Hydrothermal Vent Fields

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**Abstract** The impact of submarine hydrothermal systems on organic carbon in the ocean—one of the largest fixed carbon reservoirs on Earth—could be profound. Yet, different vent sites show diverse fluid chemical compositions and the subsequent biological responses. Observations from various vent sites are to evaluate hydrothermal systems' impact on the ocean carbon cycle. A response cruise in May 2009 to an on-going submarine eruption at West Mata Volcano, northeast Lau Basin, provided an opportunity to quantify the organic matter production in a back-arc spreading hydrothermal system. Hydrothermal vent fluids contained elevated dissolved organic carbon, particulate organic carbon (POC), and particulate nitrogen (PN) relative to background seawater. The  $\delta^{13}\text{C}$ -POC values for suspended particles in the diffuse vent fluids ( $-15.5\text{‰}$  and  $-12.3\text{‰}$ ) are distinct from those in background seawater ( $-23 \pm 1\text{‰}$ ), indicative of unique carbon synthesis pathways of the vent microbes from the seawater counterparts. The first dissolved organic nitrogen concentrations reported for diffuse vents were similar to or higher than those for background seawater. Enhanced nitrogen fixation and denitrification removed 37%–89% of the total dissolved nitrogen in the recharging background seawater in the hydrothermal vent flow paths. The hydrothermal plume samples were enriched in POC and PN, indicating enhanced biological production. The total “dark” organic carbon production within the plume matches the thermodynamic prediction based on available reducing chemical substances supplied to the plume. This research combines the measured organic carbon contents with thermodynamic modeled results and demonstrates the importance of hydrothermal activities on the water column carbon production in the deep ocean.

**Plain Language Summary** External energies fuel the production of organic carbon and nitrogen by living organisms. Exploring the organic compounds' concentrations and characteristics help reveal the energy flow and biogeochemical processes. Here, we investigate the marine organic matter in the submarine hot springs (hydrothermal fluids) generated by deep seawater interacting with uplifting magma caused by spreading seafloor. We surveyed the organic carbon and nitrogen contents with samples collected during a cruise in May 2009, in response to an on-going submarine eruption at West Mata Volcano, northeast Lau Basin. The submarine hot spring water, relative to background seawater, had elevated organic carbon and nitrogen with unique characters. The amount of elevated organic carbon meets our calculated values based on the energy that the hot spring brings to marine life. We conclude that the underwater hot springs fuel “dark” organic carbon production via unique carbon fixation pathways in the deep ocean.

## 1. Introduction

The dynamics and characteristics of organic carbon in the deep ocean potentially have profound impacts on the biological activities and the global carbon cycle (Hansell et al., 2009; Jiao et al., 2010). Marine dissolved and particulate organic carbon (DOC and POC), though operationally defined size-fractions of organic matter, play essential but distinct roles (e.g., Carlson & Hansell, 2015; Druffel et al., 1992; Griffith et al., 2012; Seymour et al., 2017). POC, composed of planktons and detritus, acts as products of biological

activities (Cauwet et al., 1997; Seymour et al., 2017), whereas the DOC, produced by excreted from organisms, lysis, POC solubility, and grazing activities, act as fuels for heterotrophic bacterial activities (e.g., Azam et al., 1983; Carlson, 2002; Ducklow & Carlson, 1992; Williams, 1970). Net productions of DOC and POC often result from temporal and spatial decoupling of its in situ biological production and consumption (Carlson, 2002; Hansell & Carlson, 2001; Honjo et al., 2008). Elevated concentrations of DOC and POC occur in areas of, or at the time of, vigorous biological activities (e.g., Carlson et al., 2000; Engel et al., 2012; Hansell & Carlson, 2001; Smith et al., 1992; Tanoue & Handa, 1979). Low DOC and low POC, such as those observed in the deep ocean, results from significantly reduced biological production in the dark ocean, accompanied by abiotic and biotic removal of DOC during thermohaline circulation (Aristegui et al., 2002; Druffel et al., 1992).

In the deep ocean, submarine hydrothermal activities also fuel biotic and abiotic organic carbon production, evidenced by elevated DOC in hydrothermal vent systems. Lang et al. (2006) reported the first systematic survey of hydrothermal vent fluid DOC. The diffuse vents off the sedimented Endeavor and the unsedimented Axial Volcano vent fields contained DOC with concentrations of 42–69  $\mu\text{M}$  and 34–71  $\mu\text{M}$ , respectively (Lang et al., 2006). As a comparison, the non-vent influenced background seawater had DOC of 36  $\mu\text{M}$  (Lang et al., 2006). Recently, Hawkes et al. (2015) observed significantly elevated DOC concentration up to 607  $\mu\text{M}$  for a diffuse vent in the unsedimented basalt-hosted Axial Volcano with relatively low hydrogen concentration (<1 mM). They also reported DOC as high as 750  $\mu\text{M}$  for a sedimented Gabbro/Peridotite-hosted focused vent (Chimlet 2) in Von Damm, Caribbean, where the hydrogen concentration was high (>10 mM). These high DOC concentrations have passed the quality check by Hawkes et al. (2015), who attributed the variable vent fluid DOC caused by the natural dynamics or random contamination from sediments, macro-, micro-fauna, or ROV operations.

Elevated suspended organic particles in hydrothermal vents and plumes also indicate production in hydrothermal systems. Comita et al. (1984) first reported that the POC and particulate nitrogen (PN) in the vent fluids and the non-vent water varied by a factor of 2–5, the higher values being in the vent fluids of the East Pacific Rise 21°N vent field. Maruyama et al. (1993) reported 5.8 to 9.2  $\mu\text{mol-C/L}$  of suspended organic particles in the black smoking vent fluids off Izena, Okinawa Trough. Bennett, Statham, et al. (2011) observed increased POC concentrations (0.87–3.81  $\mu\text{mol/L}$ ) in the near-field diffuse vents from East Pacific Rise 9°50'N.

Nevertheless, some hydrothermal systems do not favor organic matter production. The heats, the porous rocks, and the heterotrophic organisms residing in the subsurface may indeed remove deep-ocean dissolved organic matter (DOM) effectively (Hawkes et al., 2015; Lang et al., 2006; Lin et al., 2012; Walter et al., 2018). Sediment-buried ridge flank oceanic crustal fluids contained significantly depleted DOC on the Juan de Fuca (~12  $\mu\text{M}$ ) and the North Pond (18–33  $\mu\text{M}$ ) relative to recharging background seawater (Lang et al., 2006; Lin et al., 2012, 2019; McCarthy et al., 2010; Walter et al., 2018). High-temperature focused vent fluids from mid-ocean ridges at Mean Endeavor Field, Axial Volcano, East Pacific Rise 9°50'N (Bio 9) contained 14–17  $\mu\text{M}$  of DOC (Lang et al., 2006; Longnecker et al., 2018). The fractions of solid-phase extractable DOC in hydrothermal solutions during laboratory heating experiments were much less than those in deep ocean samples (Hawkes et al., 2015). These observations reinforced the importance of possible thermal alteration and removal of DOM in some hydrothermal systems.

Qualitatively, extensive seafloor and subseafloor biological and abiotic processes in hydrothermal regions should alter the composition of the organic matter. The hydrothermal ecosystem may generate organic carbon with unique stable carbon isotope compositions. Lin et al. (2019) and Walter et al. (2018) used large volume samples (1–60 L) from the warm (65°C) and cold (10°–25°C) ridge flanks, respectively, to investigate the DOC carbon isotope biogeochemistry. Lin et al. (2019) reported elevated aromatic compounds in the warm basaltic ridge flank fluids, which is likely resulted from thermal condensation reactions (Lin et al., 2019). However, organic matter characterization research often requires a large sample size (Repeata, 2015) and is thus challenging for hydrothermal studies.

Nevertheless, the organic carbon to nitrogen ratios (C/N ratios) and the stable carbon isotopic compositions ( $\delta^{13}\text{C}$ ) with small samples (few liters) provide insights into the quality of the organic matter. Comita et al. (1984) concluded that the suspended particulate organic matter (POM) in the vent fluids with low

C/N (hereafter C/N<sub>POM</sub>) of 12–18 was distinct from the high C/N<sub>POM</sub> (~24) for the presumably resistant and remineralized POM in non-vent water. The low C/N in vents may reflect the accumulation of fresh biomass, such as marine planktons, on average, having a C/N ratio of ~6 (Redfield, 1934, 1958). Low C/N ratios ( $4.1 \pm 0.6$ ) of the quantifiable suspended particles in the seafloor ridge-flank-fluid samples (Lin et al., 2019) suggest bacterial biomass as the dominant source of POC (Fukuda et al., 1998). The carbon isotopic compositions of POC also reflect the potential carbon source and carbon assimilation pathways (e.g., Hayes, 2001; Hofmann et al., 2000; Popp et al., 1998; Preuss et al., 1989; Sirevåg et al., 1977). Autotrophs preferentially incorporate <sup>12</sup>C-depleted inorganic carbon into their biomass, leading to a low  $\delta^{13}\text{C}$ -POC value in the biomass than the  $\delta^{13}\text{C}$  of inorganic carbon (House et al., 2003). Microbial metabolism using isotopically light methane (Lin et al., 2014) also results in depleted  $\delta^{13}\text{C}$ -POC value ( $-34 \pm 0.1\%$ ) in a ridge flank hydrothermal system (Lin et al., 2019).

The amounts of dissolved organic nitrogen (DON), nitrogen-containing DOC, though rarely reported, also characterize the dissolved organic matter in an aquatic environment. For example, surface seawater has elevated DON ( $5.1 \pm 1.7 \mu\text{M}$ ) than the deep sea ( $2\text{--}5 \mu\text{M}$ ; with an average of  $3.2 \pm 2.2 \mu\text{M}$ , Sipler & Bronk, 2015). In productive coastal regions, the DON can be as high as  $16 \mu\text{M}$  (Bronk et al., 2014; Sipler & Bronk, 2015). The C/N of DOM (hereafter, C/N<sub>DOM</sub>) also provide qualitative information. For instance, DOM enriched in aromatic over aliphatic compounds would have higher C/N<sub>DOM</sub> (Sipler & Bronk, 2015). DOM containing elevated humic or fulvic acids would also be high in C/N<sub>DOM</sub> (Bronk et al., 2007; See & Bronk, 2005). However, DON measurements are not yet available for hydrothermal fluids, likely due to methodological constraints. Some hydrothermal vent fluids, such as the ridge flank basaltic fluids from the eastern Juan de Fuca, contained ammonium as high as  $100 \mu\text{M}$  (Lin et al., 2012). The DON concentrations, typically obtained by subtracting inorganic nitrogen from the total dissolved nitrogen (TDN), is as large as the analytical uncertainty.

This study overcame the sampling and analytical challenges and presented the first hydrothermal vent fluid DON measurements. We report and discuss the DOC, POC,  $\delta^{13}\text{C}$ -POC, PN for diffuse vent fluids in a vent field where there was an on-going seafloor eruption. We elucidate the biogeochemical processes leading to our observation. Lastly, we investigate the amount of potential organic carbon production in the plume from the energy available by thermodynamic modeling.

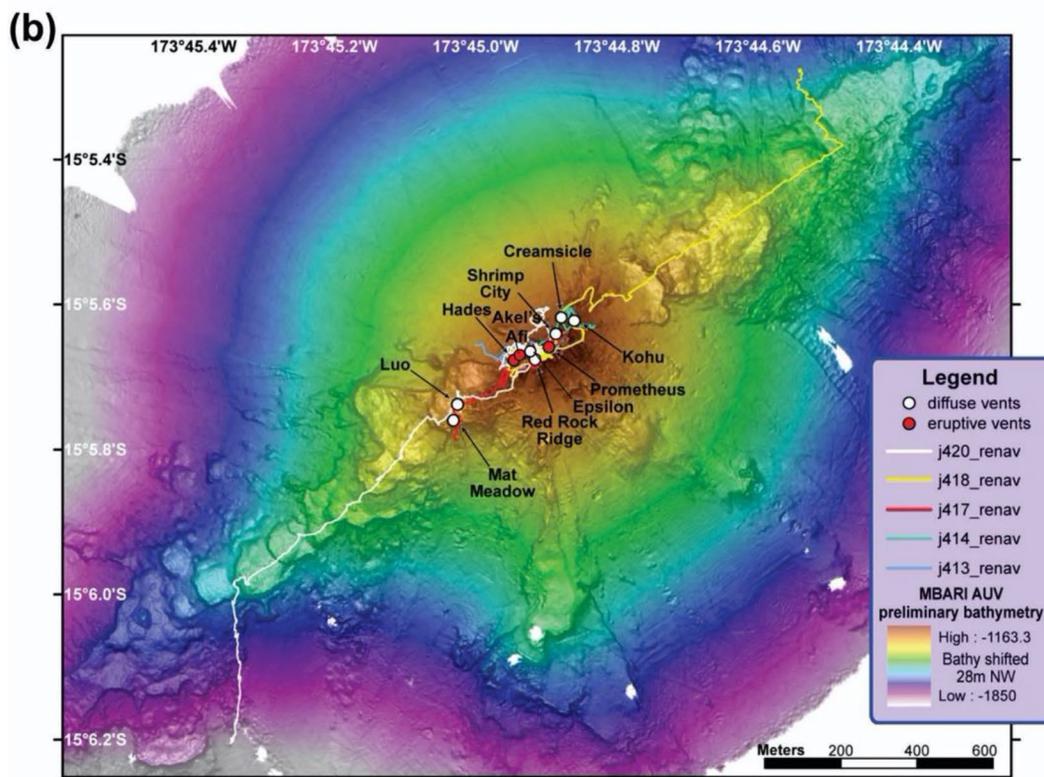
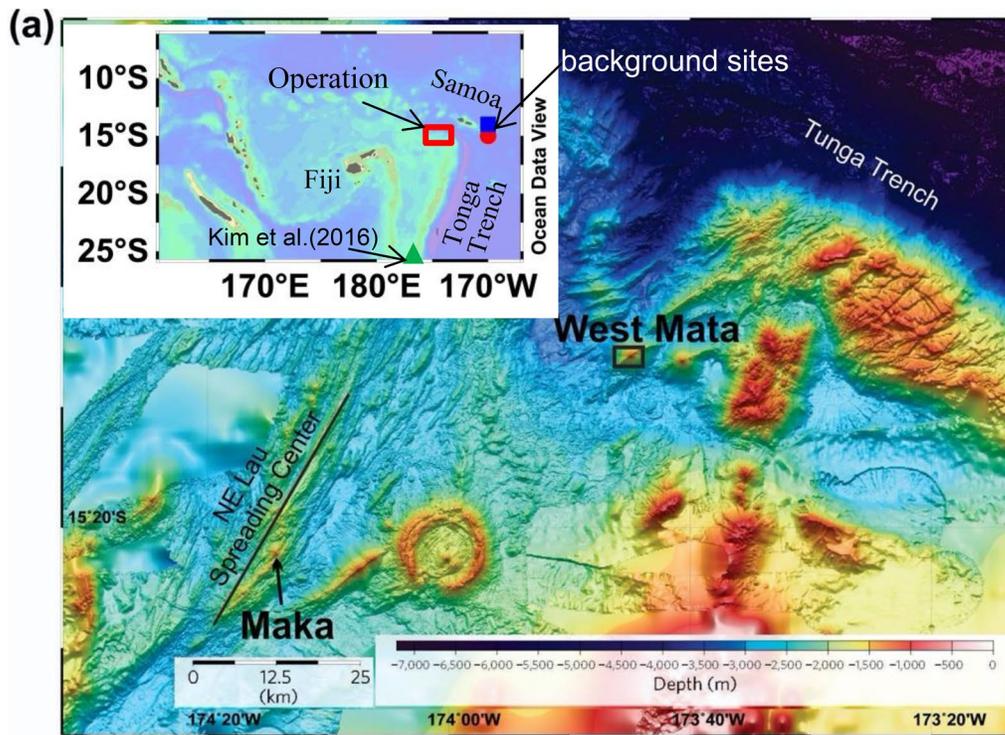
## 2. Geological Settings and Methods

### 2.1. Lau Basin

The Lau Basin is a triangular-shaped active back-arc basin associated with the Tonga-Kermadec subduction zone along the western Pacific margin (Figure 1a). There are multiple rifting and spreading centers in the Lau Back-arc Basin (Hawkins, 1995; Taylor et al., 1996); the most northeastern of them is the Northeast Lau Spreading Center (NELSC). NELSC's maximum full spreading rate is estimated to be about 94 mm/year based on the value obtained from a three-plate kinematic model for Lau Basin opening (German et al., 2006; Zellmer & Taylor, 2001). Between the NELSC and the frontal magmatic arc of the Tonga-Kermadec subduction zone lie several submarine volcanoes, including the West Mata volcano (Baker et al., 2011; Embley et al., 2014). Slab subduction induced eruptions from an active proto-arc volcano (Resing et al., 2011). This entire area is hydrothermally very active (Baker et al., 2011; German et al., 2006; Resing et al., 2011). National Oceanic and Atmospheric Administration Pacific Marine Environmental Laboratory (NOAA-PMEL) detected active eruption events on both the NELSC and at West Mata in November 2008 (Baker et al., 2011; Baumberger et al., 2014; Clague et al., 2011; Embley et al., 2014; Resing et al., 2011; Rubin et al., 2012).

### 2.2. Response Cruise and Sampling Sites

In response to the two eruptions discovered in November 2008 (Baker et al., 2011; Clague et al., 2011; Resing et al., 2011; Rubin et al., 2012), we staged a research cruise to Northeast Lau over May 5–13, 2009 on the research vessel Thomas Thompson (Resing et al., 2011). We conducted seven dives with the remotely operated vehicle (ROV) JASON II (operated by Woods Hole Oceanographic Institution). Five dives were at the West Mata area, and two dives at the NELSC (Figure 1a). We observed active eruptions at West Mata during each dive and identified three eruption sites—Hades, Akel's Afi, and Prometheus (Resing et al., 2011). Near the



**Table 1**

*Sampling Sites*

Sampling site	JASON dive#	CTD cast #	Latitude (°S)	Longitude (°W)	Sampling depth (m)
Submergence vehicle operation					
W. Mata diffuse vents					
Creamsicle	J418	–	15.093645	173.747971	1,206
Red Rock	J413	–	15.094602	173.748589	1,183
Kohu	J414	–	15.093718	173.747666	1,188
Shrimp City	J414	–	15.094023	173.748098	1,187
Luo	J417	–	15.095635	173.750425	1,277
Epsilon	J417	–	15.094530	173.748575	1,179
W. Mata Eruptive sites					
Akel's Afi	J418	–	15.094446	173.749047	1,215
Hades	J413	–	15.094707	173.749115	1,206
CTD operation					
Vertical cast					
Above W. Mata	–	V09C01	15.094278	173.748523	949–1,148
Above Maka	–	V09C02	15.423150	174.284933	1,195–1,556
Tow-yo					
NE Lau Spreading Center	–	T09C01	15.381515	174.242577	996
NE Lau Spreading Center	–	T09C01	15.390883	174.251013	1598

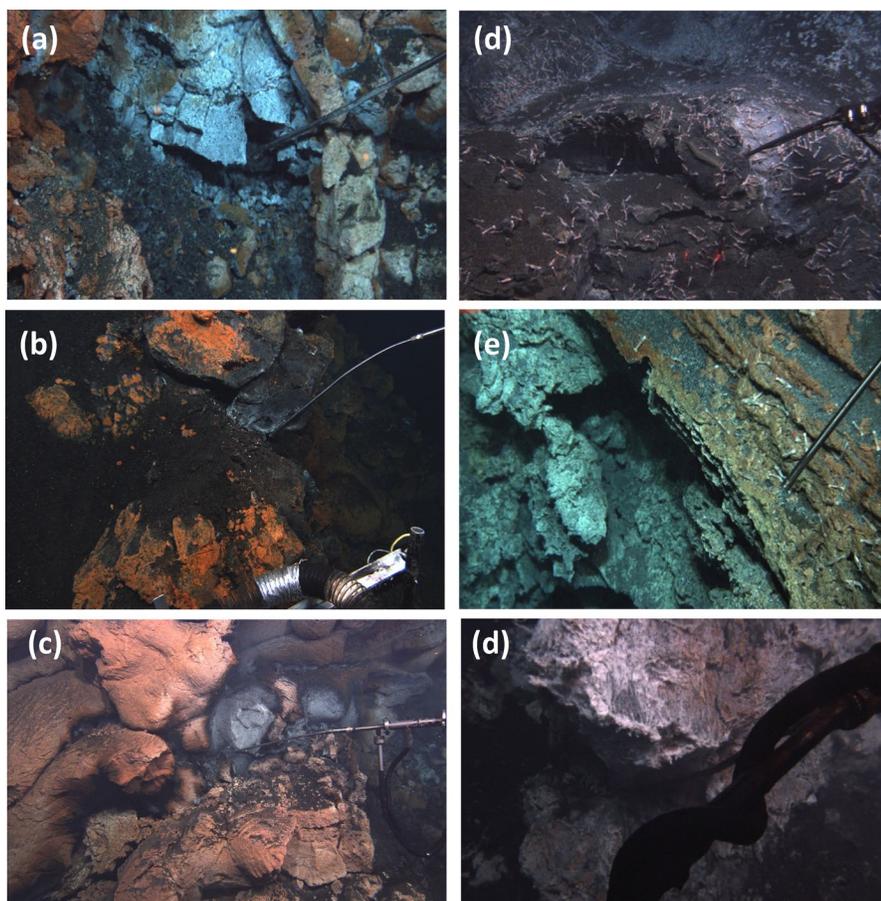
eruption sites, we discovered several low-temperature diffuse vents—Creamsicle, Red Rock Ridge, Kohu, Shrimp City, Luo, and Epsilon (Table 1 and Figures 1 and 2). In contrast, we found no eruptive activity along with the NELSC (Figure 1).

We conducted several CTD-rosette operations from the research vessel, including two vertical casts (V09C01 and V09C02, Table 1) over the West Mata eruptive site and the NELSC, respectively. We made two horizontal tows during which the CTD was continuously lowered and raised in a saw-toothed pattern (i.e., tow-yos; Baker & Massoth, 1987) as water samples were collected. Sensors on the CTD rosette recorded conductivity, temperature, pressure, oxygen, optical backscatter, fluorescence, altimetry, and oxidation-reduction potential (ORP). Anomalies in ORP indicate the presence of reduced chemical species such as H<sub>2</sub>S and Fe<sup>2+</sup>.

### 2.3. Sampling and Analytical Methods

Plume and background seawater samples were collected with Niskin bottles during CTD-rosette vertical and tow-yo casts following the guide to ocean carbon research (Dickson et al., 2007). There is not yet a consensus protocol to collect hydrothermal fluid samples for organic carbon research. Hydrothermal fluids are usually directly collected into a bag, polyvinyl chloride (PVC), or titanium sampler (Bennett, Hansman, et al., 2011; Bennett, Statham, et al., 2011; Hawkes et al., 2015; Lang et al., 2006, 2010; Lin et al., 2012, 2019). There was no opportunity to rinse the sample storage devices during seafloor operation. Previous research pointed out that hydrothermal samples are prone to receive random DOC contamination (Hawkes et al., 2015). Thus, contamination controls on the sampling systems are crucial to using the samples for DOC research.

**Figure 1.** Map of the study site. (a) Regional map of NE Lau basin. The insert shows the location of the NE Lau basin relative to Fiji, Samoa, and Tonga Trench (Ocean Data View; Schlitzer et al., 2018). The red box indicates the operation area for this study and is enlarged in Figure 1a. The red circle and blue square indicate the locations of two background water column casts (15°S, 170°W; 14°S, 170°W; transect P15S of WOCE/CLIVAR <http://yyy.rsmas.miami.edu/groups/biogeochem/Data.html>), used for comparison to data obtained by this study. The green triangle denotes the sites for the sinking particle research by Kim et al. (2016). A black box denoted the area of West Mata enlarged in Figure 1b. (b) Detailed bathymetric map of W. Mata. Map credit: Susan Merle OSU CIMRS/NOAA EOI.



**Figure 2.** Sampling photos: Diffuse vents (a) Creamsicle, (b) Red Rock, (c) Kohu, (d) Shrimp City, (e) Luo, and (f) Epsilon.

For this study, hydrothermal vent fluid samples were collected via ROV JASON II using the PMEL Hydrothermal Fluid and Particulate Sampler (HFPS; Butterfield et al., 2004; Huber et al., 2006) into 800 mL Tedlar bags. The HFPS lids for the bags were cleaned and sterilized with ethanol, rinsed vigorously with deionized water, and finally rinsed with 0.2  $\mu\text{m}$  filtered seawater. The DOC blank for an HFPS bag was  $\sim 1 \mu\text{M}$ , obtained by measuring the deionized water sat in the Tedlar bag for several hours. The TDN blank value for the bag was  $< 0.5 \mu\text{M}$ . The DOC blank of the HFPS bag increased to  $5.2 \mu\text{M}$  in extremely acidic conditions (0.24N HCl,  $\text{pH} < 1$ ) for a prolonged period while the TDN remained undetected ( $< 0.5 \mu\text{M}$ ).

All fluid samples for DOC, TDN, and nutrient analyses were filtered shipboard through GF/F filters (25 mm; precombusted at  $550^\circ\text{C}$  for 5 h) in acid-cleaned Whatman<sup>®</sup> polypropylene 25 mm filter holders (Whatman International Ltd, UK). Before use, we passed at least 50 mL of deionized water through each filter to lower a DOC blank to below the detection limit ( $1 \mu\text{M}$ ) based on our laboratory test results. The filtered samples were stored in acid-washed high-density polyethylene Nalgene bottles and kept frozen until thawed for analysis (Tupas et al., 1994).

Particulates were collected by pumping 2–5 L of hydrothermal vent fluids through combusted GF/F filters using the HFPS in situ. About 10 L of seawater collected with CTD-rosette Niskin bottles were filtered on board ship through combusted GF/F filters. Individual filters were wrapped in combusted foils and stored frozen until thawed for analysis.

DOC and TDN concentrations were measured by high-temperature ( $720^\circ\text{C}$ ) combustion using a Shimadzu TOC- $V_{\text{CSH}}$  analyzer. Samples were acidified to  $\text{pH} < 2$  within the autosampler syringe by adding  $45 \mu\text{L}$  of 2N HCl to 3 mL samples. No acid contamination was observed throughout the analysis. The acidified

samples were purged with purified air (Shimadzu compressed air and carrier gas purification kit) to remove inorganic carbon. Five to six replicate analyses were performed using an injected sample volume of 150  $\mu\text{L}$ . DOC and TDN concentrations were calibrated with gravimetrically prepared potassium hydrogen phthalate (KHP) and sodium nitrate standards. The reproducibility between replicate injections was  $<2 \mu\text{M}$ , that is,  $<5\%$  at 40  $\mu\text{M}$ . Two consensus reference materials (CRM, from University of Miami, Dickson et al., 2007), Florida Strait 700 m-deep seawater (DSW) and low carbon water (LCW), were used extensively before, between, and after sample analysis. At least one CRM was measured every five samples. The measured concentration of CRM-DSW was  $42 \pm 0.2 \mu\text{M}$  ( $n = 23$ ), consistent with the certified range of DOC concentration (41–43  $\mu\text{M}$ ). The average measured CRM-LCW value was  $1.3 \pm 0.4 \mu\text{M}$  ( $n = 7$ ), which also fell within the certified range of 1–2  $\mu\text{M}$ . The detection limit was about 1  $\mu\text{M}$ . TDN was measured with a chemiluminescence detector in-line with the described Shimadzu TOC-V<sub>CSH</sub> analyzer (Sharp et al., 2002). Measured CRM-DSW was  $33 \pm 0.1 \mu\text{M}$  ( $n = 23$ ), the same as the certified value of 33  $\mu\text{M}$ . The analytical precision was 0.1  $\mu\text{M}$ , and the detection limit was about 0.5  $\mu\text{M}$ .

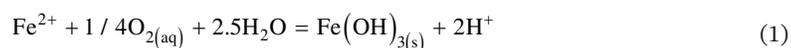
Nitrate-plus-nitrite (N + N) analysis was carried out with a custom-build colorimetric flow injection analyzer (Grasshoff et al., 1999). Sample nitrate was first reduced to nitrite with a Cu-Cd column and then analyzed as nitrite. The Cu-Cd column's reduction efficiency, the absorbance ratio of nitrate and nitrite standard at an equal concentration (e.g., 5 or 20  $\mu\text{M}$ ; gravimetrically prepared), was maintained at  $100 \pm 1\%$ . We regenerated the Cu-Cd column with ammonium chloride between each sample to minimize possible interferences by the reducing compounds in the hydrothermal fluids. The detection limit for N + N analysis was 0.1  $\mu\text{M}$ , and analytical precision was  $<0.05 \mu\text{M}$ . Ammonium concentrations were measured at the University of Hawaii with a flow injection-fluorometric method (Jones, 1991) and calibrated against gravimetrically prepared ammonium chloride ( $\text{NH}_4\text{Cl}$ ) standards. The detection limit was 0.1  $\mu\text{M}$  at the setting for measuring low concentrations of ammonium in the vent fluids. The analytical uncertainty was 0.03  $\mu\text{M}$ .

Silicate was measured by colorimetry (Grasshoff et al., 1999). Samples were first diluted 100 times, reacted with acidic molybdenum, and reduced with ascorbic acid. The analytical uncertainty for the diluted samples was about 0.1  $\mu\text{M}$ . Total dissolved iron (1-sigma precision 4%) in vent fluids was measured by atomic absorption at PMEL/Seattle on samples preserved shipboard with ultra-pure HCl (TraceMetal Grade). The total iron of acidified plume samples was measured by a Ferrozine colorimetry method (Gibbs, 1976; Stookey, 1970). The ferrozine method had an analytical uncertainty of 0.03  $\mu\text{M}$  and a detection limit of 0.1  $\mu\text{M}$ . The pH was measured shipboard at laboratory temperature in a closed vessel using a glass pH electrode (Ross Sure-Flow) calibrated with NBS-certified buffers (Fisher Scientific). The precision of the measurement is 0.02 pH units.

The POC and PN and isotopic compositions of acidified (with 1 N HCl) GF/F samples were measured with a CN elemental analyzer (Costech, ECS 4010) connected in-line with an isotope ratio mass spectrometer (IRMS; Thermo Finnigan Delta XP) interfaced with a ConFloIV, at the University of Hawaii. Isotope values are reported in conventional  $\delta$  notation relative to the international standards atmospheric  $\text{N}_2$  and V-PDB, for N and C, respectively. The detection limit for carbon and nitrogen content is 10  $\mu\text{g-C}$  and 0.3  $\mu\text{g-N}$ , respectively. The limit for reliable carbon and nitrogen isotopic determination is 10  $\mu\text{g-C}$  and 10  $\mu\text{g-N}$ , respectively. Glycine was used as the reference material for both the quantification of C and N and the isotopic correction. A homogenized tuna white muscle tissue reference was also analyzed at least triplicate as an external standard to verify instrument accuracy and precision. Our in-house reference materials were extensively characterized with certified reference materials, including NBS 18, NBS 19, NIST 1547, NIST 3, USGS 32, USGS 34, and USGS 35.

#### 2.4. Chemoautotrophic Production Energetics

The oxidation reactions of reduced species, using  $\text{Fe}^{2+}$  as an example, can be expressed as:



The amount of energy released or required for the oxidation reactions is expressed as Gibbs free energy of reaction ( $\Delta G_r$ ) and is calculated as:

$$\Delta G_r = \Delta G_r^\circ + RT \ln Q \quad (2)$$

where  $\Delta G_r^\circ$  is the standard state Gibbs energy of reaction at the temperature and pressure of interest,  $R$  is the gas constant, and  $T$  is the temperature in Kelvin.  $Q$  is the reaction quotient and can be calculated as:

$$Q = \Pi \left( a_i^{v_{i,r}} \right) \quad (3)$$

where  $a_i$  denotes the activity of chemical species  $i$ , and  $v_{i,r}$  is its stoichiometric coefficient in reaction  $r$ , which is negative for reactants and positive for products.

$\Delta G_r^\circ$  is derived from the respective equilibrium constant ( $K$ ) near in situ pressure (120 bar). The equilibrium constants at 4°C were computed using the revised H.K.F. equations of state (Helgeson et al., 1981; Shock et al., 1992; Tanger & Helgeson, 1988), the SUPCRT 92 software package (Johnson et al., 1992):

$$\Delta G_r^\circ = -2.3RT \log K \quad (4)$$

The  $Q$  values were calculated based on the maximum values reported for the 2008 plume (Resing et al., 2011) or the values derived from the electronic atlas of WOCE hydrographic and tracer data (Schlitzer, 2000). The activity coefficients of neutral species such as dissolved oxygen and hydrogen were assumed to be unity (McDermott et al., 2015).

### 3. Results

Directly measured values are reported due to the difficulty in obtaining a suitable factor for seawater entrainment correction (details in Section 4.1).

#### 3.1. DOC, TDN, $\text{NO}_3^-$ , $\text{NH}_4^+$ , DON, and POC in Vent Fluid Samples

The DOC concentrations (39–521  $\mu\text{M}$ ) in the diffuse vent fluids from West Mata vary from close to bottom seawater values ( $\sim 39 \mu\text{M}$ ) at Creamsicle and Red Rock to greatly elevated values at Luo and Epsilon (Table 2). The agreement of the replicate bag samples at Creamsicle (Table 2) documents the consistency of our sampling and analytical procedures.

The elevated DOC of vent fluids from Luo at NE Lau was within the range of porewater extracted from microbial mats recovered from the Loihi vent field (68–179  $\mu\text{M}$ , Bennett, Hansman, et al., 2011). The diffuse vents contained TDN concentrations of 4.3–24.5  $\mu\text{M}$ , much lower than bottom seawater (39  $\mu\text{M}$ ). The vent fluids had  $\text{NO}_3^-$  between 1.8 and 21  $\mu\text{M}$ , but background seawater had  $\text{NO}_3^-$  up to 37  $\mu\text{M}$ . The vent fluids had 0.3–3.4  $\mu\text{M}$   $\text{NH}_4^+$  whereas seawater  $\text{NH}_4^+$  was undetectable (Table 2). The calculated DON was between 1.6 and 3.4  $\mu\text{M}$  with cumulative errors of between 0.1 and 0.5  $\mu\text{M}$ . The POC and PN concentrations in Red Rock and Luo diffuse vent fluid samples are an order of magnitude higher than that for background seawater (Table 3). The  $\delta^{13}\text{C}$  values for the POC from Red Rock and Luo vents are  $-15.5\text{‰}$  and  $-12.3\text{‰}$ , respectively, which is significantly enriched in  $^{13}\text{C}$  relative to the POC in nearby background seawater ( $-23 \pm 1\text{‰}$ ) (Table 3).

#### 3.2. DOC, TDN, POC, and PN in Hydrothermal Plume Samples

We identified a pronounced particle-rich layer  $\sim 80$ – $100$  m above the West Mata eruption sites as a neutrally buoyant hydrothermal plume, accompanied by anomalously high turbidity and low ORP (Table 4; Figure 3). The DOC concentrations in the neutrally buoyant plume samples above West Mata ( $45 \pm 1 \mu\text{M}$ ) are significantly higher than those of background seawater ( $39 \pm 1 \mu\text{M}$ ). TDN concentrations in plume samples above both West Mata are uniformly  $39 \pm 1 \mu\text{M}$ , not significantly different from those in background seawater. POC and PN concentrations,  $0.45 \pm 0.12 \mu\text{mol-C/L}$  and  $0.043 \pm 0.08 \mu\text{mol-N/L}$  ( $n = 4$ ), respectively, in West Mata plume samples are higher than those in background seawater samples ( $0.18 \pm 0.06 \mu\text{mol-C/L}$  and  $0.022 \pm 0.09 \mu\text{mol-N/L}$ ; Table 4; Figure 4).

**Table 2**

*Diffusive Vent Fluid and Background Seawater Biogeochemical Compositions: Maximum Temperature During Sampling ( $T_{max}$ ), Equilibrated pH at 25°C, Concentrations of Magnesium (Mg), Dissolved Organic Carbon (DOC), Total Dissolved Nitrogen (TDN), Ammonium ( $NH_4^+$ ), Nitrite ( $NO_2^-$ ), Nitrate ( $NO_3^-$ ), Calculated Dissolved Organic Nitrogen ( $DON = TDN - NH_4^+ - NO_2^- - NO_3^-$ ), silicate (Si), Total Dissolved Iron (TdFe), and Phosphate ( $PO_4^{3-}$ )*

Vent type & name	Sampler type	$T_{max}$ (°C)	pH	Mg (mM)	DOC ( $\mu$ M)	TDN ( $\mu$ M)	$NH_4^+$ ( $\mu$ M)	$NO_2^-$ ( $\mu$ M)	$NO_3^-$ ( $\mu$ M)	Cal. DON ( $\mu$ M)	C/N. DOM	DON: TDN	Si ( $\mu$ M)	TdFe ( $\mu$ M)	$PO_4^{3-}$ ( $\mu$ M)
Diffusive vent															
Creamsicle	HFPS-Bag	29.3	3.0	58	$39 \pm 2$	$4.3 \pm 0.1$	0.3	<0.05	1.8	2.1	18	50%	2986	812	6.3
Creamsicle	HFPS-Bag	29.3	2.9	58	$41 \pm 5$	$5.8 \pm 0.4$	0.3	<0.05	-	-	-	-	3,104	840	5.6
Red Rock	HFPS-Bag	13.8	3.2	57	$43 \pm 1$	$24.5 \pm 0.5$	0.4	<0.05	21	3.0	14	12%	1,173	854	6.2
Kohu	HFPS-Bag	30.8	3.0	58	$86 \pm 3$	$13.9 \pm 0.3$	0.5	<0.05	10	3.4	25	25%	3,319	962	6.8
Shrimp City	HFPS-Bag	14.8	5.8	57	$84 \pm 2$	$10.8 \pm 0.4$	0.8	<0.05	8	2.1	40	19%	1,084	21	6.1
Luo	HFPS-Bag	22.4	5.3	55	$126 \pm 7$	$12.7 \pm 0.3$	3.4	<0.05	6	3.3	38	26%	1,570	124	9.3
Epsilon	HFPS-Bag	29.6	2.8	59	$521 \pm 5$	$21.5 \pm 0.4$	0.9	<0.05	19	1.6	322	8%	2235	1,078	8.8
Background seawater															
T09C-01, 996 m	Niskin-CTD	4.5	7.78	53	$40 \pm 0.3$	$38 \pm 1.0$	<0.1	<0.05	37	1.6	25	4%	69	<0.1	2.1
T09C-01, 1,598 m	Niskin-CTD	3.0	7.77	53	$38 \pm 0.7$	$39 \pm 1.0$	<0.1	<0.05	37	2.9	13	7%	100	<0.1	2.1
V09C-02, 1,557 m	Niskin-CTD	3.2	7.76	53	$38 \pm 0.4$	$39 \pm 0.7$	<0.1	<0.05	37	2.5	16	6%	103	<0.1	2.1

*Note.* Temperature not measured during sampling; or no aliquot was available for nitrate analysis and thus dissolved organic nitrogen could not be calculated. Abbreviation: N.A., not available.

## 4. Discussions

### 4.1. Sample Overview

The end-member of a high-temperature focused vent is typically obtained using Mg contents to correct seawater entrainment during seafloor sampling (Bischoff & Seyfried, 1978; Mottl, 1983; Seyfried et al., 2003). However, there is no suitable proxy to derive end-member of diffuse vents (Butterfield et al., 2011; Gamo et al., 1997). Nevertheless, the high temperatures (13–50°C), low pH (1.5–5.8), high silicate (>1,000  $\mu$ M), high dissolved iron (21–1,078  $\mu$ M) were all indicative of vigorous water-rock reactions. The fluids were likely to have been minimally mixed with seawater. Thus, the measured values without further correction were reported as the end-member values and discussed.

**Table 3**

*Concentrations and Isotopic Compositions of Particulate Organic Carbon (POC) and Particulate Nitrogen (PN) for Suspended Particles in Hydrothermal Fluids from Three Diffuse Vents*

Sample name	POC ( $\mu$ mol/L)	PN ( $\mu$ mol/L)	C/N-POM	$\delta^{13}C$ -POC (‰)	$\delta^{15}N$ -PN (‰)	Cell/L ( $\pm 95\%$ C.I.)	Dive #
Background seawater	0.23	0.028	8.1	-23.3	N.D.	$2.5(\pm 0.2) \times 10^4$	CTD
Diffusive vent							
Creamsicle	-	-	-	-	-	$1.3(\pm 0.1) \times 10^5$	J2-418
Red Rock	2.45	0.398	6.1	-15.5	N.D.	-	J2-418
Kohu	0.48	0.042	11.4	N.D.	N.D.	$1.3(\pm 0.2) \times 10^5$	J2-414
Shrimp City	-	-	-	-	-	$2.4(\pm 0.3) \times 10^5$	J2-414
Luo	3.08	0.605	5.1	-12.3	4.3	$2.2(\pm 0.2) \times 10^5$	J2-417
Epsilon	-	-	-	-	-	$2.2(\pm 0.2) \times 10^5$	J2-417

*Notes.* N.D.: Not detectable due to insufficient C and N materials collected for isotope analysis. The detection limit for POC is 8  $\mu$ g and for PN is 2  $\mu$ g. The isotopic compositions of carbon and nitrogen cannot be determined when POC and PN are lower than 10 and 8  $\mu$ g, respectively.

**Table 4**

Concentrations of Dissolved Organic Carbon (DOC), Total Dissolved Nitrogen (TDN), Particulate Organic Carbon (POC), Particulate Nitrogen (PN), Turbidity (dNTU), and Oxidation and Reduction Potential (ORP) in the W

Station (cast ID)	Depth (m)	DOC ( $\mu\text{M}$ )	TDN ( $\mu\text{M}$ )	POC ( $\mu\text{mol/L}$ )	PN ( $\mu\text{mol/L}$ )	C/N-POM	$\delta^{13}\text{C-POC}$ (‰)	Si ( $\mu\text{M}$ )	Fe ( $\mu\text{M}$ )	dNTU	ORP-02	ORP-07	Niskin bottle #	UH ID
Plume above W. Mata summit (1176m. 15.09°S, 173. 75°W; above active eruption sites)														
V09C01	1,092	45 ± 1.0	40 ± 0.8	0.28	0.031	8.8	−25.3	–	<0.1	0.003	−27	15	24	7
V09C01	1,096	45 ± 0.9	39 ± 0.3	0.46	0.044	10.5	−21.5	–	<0.1	1.459	−91	−141	16	6
V09C01	1,099	44 ± 0.1	39 ± 0.6	0.50	0.045	11.0	−22.4	78	<0.1	1.262	−82	−115	20	5
V09C01	1,134	45 ± 1.1	39 ± 0.8	0.56	0.050	11.2	−24.8	78	<0.1	0.062	53	102	45	4
Above Maka (1569m. 15.42°S, 174. 28°W)														
V09C02	1557	38 ± 0.4	39 ± 0.7	0.28	0.029	9.6	−22.1	–	–	0.045	26	137	17	34
V09C02	1297	41 ± 0.8	39 ± 0.7	–	–	–	–	91	<0.1	0.001	35	123	21	7
V09C02	1370	40 ± 2.0	39 ± 0.5	0.23	0.028	8.0	−24.4	90	<0.1	0.001	25	116	20	43
Background seawater (NELSC)														
T09C-01	996	40 ± 0.3	38 ± 1.0	0.14	0.015	9.1	–	69	<0.1	0.001	32	230	18	16
T09C-01	1,598	38 ± 0.7	39 ± 1.0	0.23	0.028	8.1	−23.3	82	<0.1	0.004	130	226	52	15

Notes. Mata vent sites. Seawater samples were collected with Niskin bottles on a CTD rosette deployed from the research vessel.

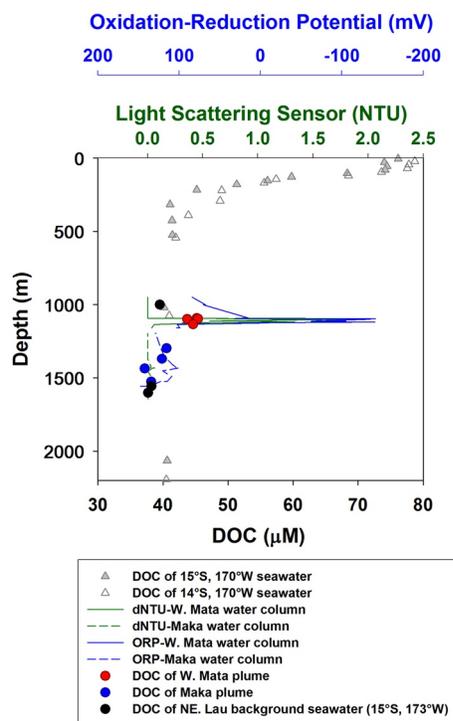
## 4.2. Biological Production of Organic Carbon

### 4.2.1. Carbon-Fixation Pathways

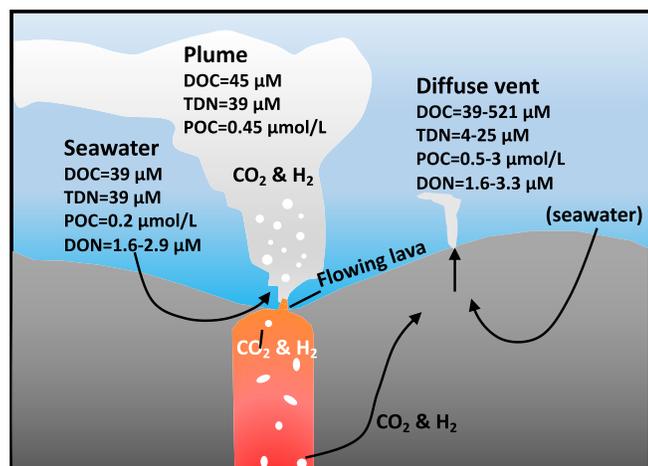
To explore the organic biogeochemistry of the W. Mata vent field in a global context, we summarized literature values of DOC, POC, C/N<sub>POM</sub>, and  $\delta^{13}\text{C-POC}$  for various deep-sea hydrothermal vents and plumes in Table 5. Biological organic carbon production in the W. Mata back-arc spreading hydrothermal system is evidenced by the elevated DOC and POC, elevated cell abundances (Table 3), Redfield-like C/N<sub>POM</sub>, distinct  $\delta^{13}\text{C-POC}$  values, and the depleted TDN. Additional evidence comes from the abundant filamentous microbial mats observed at Epsilon and Creamsicle (Figure 2). Scattered microbial mats were also observed at Shrimp City and Luo, where high abundances of indigenous shrimp were also present (Figure 2).

The cell abundances ( $1.3 \times 10^5$  to  $2.4 \times 10^5$  cells/mL) within the diffuse vent fluids were 5-to-10 folds higher than those in background seawater ( $2.5 \pm 0.2 \times 10^4$  cells/mL). The high cell counts were consistent with elevated POC and PN concentrations (Table 3). The POC in the Red Rock Ridge and Luo vent fluids had C/N ratios similar to the Redfield ratio (6.6), indicating that the POC was freshly made biomass. The C/N of POC from Kohu vent is much higher than those from the Red Rock and Luo vents, suggesting that the POC from Kohu might have undergone some degree of degradation (Table 3). Degradation of POC preferentially removes N over C, resulting in elevated C/N ratios (Martin et al., 1987). Alternatively, the production of carbon-rich POM such as polysaccharides.

The POC in Red Rock and Luo vents were significantly more enriched in  $^{13}\text{C}$  than that in nearby background seawater ( $-23 \pm 1\%$ , Table 3) or in the hydrothermal-vent-influenced sinking particles from the Tonga arc region ( $-24.3\%$  to  $-21.4\%$ ; Kim et al., 2016) (Table 5). The majority of the W. Mata POC must have been produced within the subsurface or on the seafloor, and only to a small extent accumulated sinking particles



**Figure 3.** Dissolved organic carbon (DOC) concentrations in hydrothermal plume fluids and background seawater. Filled circles are DOC data from this study. Open and filled triangles are DOC data generated and published on-line by Dennis Hansell at the University of Miami to provide additional local background seawater values (15°S, 170°W; 14°S, 170°W, transect P15S of WOCE/CLIVAR <http://yyy.rsmas.miami.edu/groups/biogeochem/Data.html>).



**Figure 4.** Schematic drawing of the production of organic carbon (DOC) and depletion of total dissolved nitrogen (TDN) in West Mata vent field. In this model, high DOC is produced within the subsurface and at the seafloor or on the vent surface habitats. The circulating hydrothermal fluids entrain DOC before and after the fluids are heated. Some DOC may be generated by thermal degradation of particulate organic carbon during the ascending of hydrothermal fluids, resulting in high DOC observed in fluids collected. Drawing modified from Butterfield et al. (2011). POC, particulate organic carbon. DON, dissolved organic nitrogen.

such as the POC from the water column (Table 3). The  $^{13}\text{C}$  enrichment suggests that the resident biomes fixed carbon via a pathway with small carbon isotope fractionation ( $\epsilon$ ) between bicarbonate and POC. From dissolved inorganic carbon (DIC) in a typical hydrothermal vent with a  $\delta^{13}\text{C}$ -DIC value between 0‰ to  $-9$ ‰ (Takai et al., 2008; Walker et al., 2008; Zerkle et al., 2005), vent biome would generate organic carbon with  $\delta^{13}\text{C}$ -DOC values of  $-0.2$ ‰ to  $-14.5$ ‰ via the reductive tricarboxylic acid (rTCA) cycle ( $\epsilon$  of 2–5.5‰) or via the 3-hydroxypropionate/4-hydroxybutyrate (3-HP/4-HB) cycle ( $\epsilon$  of 0.2–3.6‰) (House et al., 2003; Zerkle et al., 2005; Zhang et al., 2002). In contrast, the biomass would have  $\delta^{13}\text{C}$ -DOC values of between  $-22$ ‰ and  $-44.5$ ‰ via the Calvin-Benson cycle ( $\epsilon$  of 22–35.5‰) (e.g., Wong & Sackett, 1978).

The use of the rTCA cycle for biomass synthesis requires much less energy than the Calvin-Benson cycle (White et al., 2012). Thus, chemolithotrophic micro-organisms residing in hydrothermal environments may have adapted to utilize such metabolic pathways to thrive in places where available energy is constrained by the total amounts of available chemical compounds (McCollom & Amend, 2005). Based on genomic sequences, rTCA cycles have been suggested to be the carbon fixation pathways for some deep-sea hydrothermal isolates or closely related strains (Hügler & Sievert, 2011), for example, *phylum Aquificae* (Hügler et al., 2007), *Epsilonproteobacteria* (Hügler et al., 2005), and *thermophilic archaeon Pyrococcus sp. Strain NA2* (Lee et al., 2011). Organisms utilizing the 3-HP cycle are not yet isolated but have been suggested to be important contributors at deep-sea hydrothermal vents (Nakagawa & Takai, 2008; Nakagawa et al., 2006).

#### 4.2.2. Enhanced Nitrogen Fixation and Denitrification

The TDN concentrations in the West Mata low-temperature diffuse vents were much lower than those in bottom seawater (Table 2), indicating available nitrogen was consumed likely by organisms in a prosperous subsurface and surface vent habitat. Similar to photosynthesis, chemosynthesis converts DIC, and dissolved inorganic nitrogen (DIN) into POM (Gruber, 2008). In an anoxic environment, denitrifiers reduce nitrate, ultimately into nitrogen gas, decreasing the nitrate concentration in the fluids (Bourbonnais et al., 2012; Butterfield et al., 2004; Devol, 2008). All of the surveyed W. Mata low-temperature diffuse vents contained ammonium, demonstrates nitrate reduction along the fluid flow paths.

The dissolved and particulate nitrogen was not mass-balanced between the recharging background seawater and the discharging diffuse vents. The total decrease in the TDN was 14 and 25  $\mu\text{M}$ , exceeding the combined increase in PN and DON in the Red Rock and Luo vents (1.87 and 2.28  $\mu\text{mol/L}$ ) (Tables 2 and 4). The PN only measured the suspended organic matter (living cells and detritus) in the vent fluids retained on the filters. In contrast, the DON measures the organic nitrogen dissolved in the water. The rest of the lost TDN was likely converted into rock-attached biomass and biofilms within the subsurface. Alternatively, denitrification ultimately to gaseous nitrogen within the subsurface was a dominant process.

The sources and sinks of the DON in the hydrothermal system are intimately linked to assimilation, nitrification, and denitrification. Microbial, macrofauna, and microfauna activities on the seafloor and the subsurface release the DON into the hydrothermal fluids. Heterotrophic activities consume DON, and heat degradation reduces DON in the hydrothermal systems. These DON input and removal processes might alter the  $\text{C}/\text{N}_{\text{DOM}}$ . The Creamsicle, Red Rock, and Kohu vents had  $\text{C}/\text{N}_{\text{DOM}}$  ranged from 14 to 25, similar to the  $\text{C}/\text{N}_{\text{DOM}}$  of nearby background seawater (13–25; Table 2). The high  $\text{C}/\text{N}_{\text{DOM}}$  in Shrimp City (40) and Luo (38) suggests that the vent-derived DOM might contain more aromatic compounds than the DOM from other vent sites. Shrimps were abundant in Shrimp City (40) and Luo (Figure 2), suggesting possible DOM derived from the shrimp-ectosymbionts and from the metacommunity activities (Zbinden & Cambon-Bonavita, 2020). The elevated  $\text{C}/\text{N}_{\text{DOM}}$  over the  $\text{C}/\text{N}_{\text{POM}}$  (6.1–11.2) observed for the Red Rock, Kohu, and Luo vents suggests preferential removal of N-containing organic matter during the transformation from

**Table 5**  
*Summary of Dissolved Organic Carbon (DOC), Particulate Organic Carbon (POC) Concentrations,  $\delta^{13}\text{C}$ -POC (‰), and The C to N Ratio of The Particulate Organic Matter (C/N<sub>POM</sub>) of This Research and in The Literature*

Study sites	DOC ( $\mu\text{M}$ )	POC ( $\mu\text{mol/L}$ )	$\delta^{13}\text{C}$ -POC (‰)	C/N <sub>POM</sub>	Reference
<b>Background seawater</b>					
West Mata, Lau (956 m & 1,598 m)	38–40	0.14–0.23	–23	8–9	This study
North Central Pacific (Eve 1; 1,800–5,600 m)	34–38	0.049–0.129	–21.7 to –22.6	–	Druffel et al. (1992)
Sargasso Sea (Hydro 6; 852–4,450 m)	40–43	0.040–0.106	–20.2 to –20.5	–	Druffel et al. (1992)
East Pacific Rise 9°50'N	37–39	0.11–0.21	–	–	Bennet, Statham, et al. (2011)
East Pacific Rise 21°N	–	–	–	24	Comita et al. (1984)
Loihi (1,003 m & 1,072 m)	39.2–40.4	0.1	–	–	Bennet, Hansman, et al. (2011)
North Pond (4,000 m)	40–50	0.1–0.3	–	7–8	Meyer et al. (2016); Walter et al. (2018)
<b>Hydrothermal plume</b>					
Above West Mata vent field	44–45	0.28–0.56	–21.5 to –25.3	9–11	This study
Loihi sea mount; dispersing plume	41–60	0.06–0.2	–20 to –28	–	Bennet, Hansman, et al. (2011)
East Pacific Rise 9°50'N	35–43	0.07–0.46	–	–	Bennet, Statham, et al. (2011)
East Pacific Rise 9°50'N; near-field buoyant plume	38–39	0.87–3.81	–	–	Bennet, Statham, et al. (2011)
<b>Low-temperature diffuse vent</b>					
West Mata, Lau	39–521	2.5–4.1	–12.3 to –15.5	5–11	This study
Baby Bare, Juan de Fuca	7–27	–	–	–	Lang et al. (2006)
Endeavor	42–69	–	–	–	Lang et al. (2006)
Axial Volcano	34–71	–	–	–	Lang et al. (2006)
Axial Volcano	61–189	–	–	–	Hawkes et al. (2015), <sup>b</sup>
Loihi sea mount	54–92	–	–	–	Bennet, Hansman, et al. (2011)
East Pacific Rise 9°50'N; near field vent	38–47	0.87–3.81	–	–	Bennet, Statham, et al. (2011)
East Pacific Rise 21°N	–	0.4–1 <sup>a</sup>	–	12–18	Comita et al. (1984)
<b>High-temperature focused vent</b>					
Lost City	101–106	–	–	–	Lang et al. (2010)
Endeavor	11–26	–	–	–	Lang et al. (2006)
Endeavor	15–22	–	–	–	Hawkes et al. (2015), <sup>b</sup>
Axial Volcano	8–24	–	–	–	Lang et al. (2006)
Axial Volcano	24–50	–	–	–	Hawkes et al. (2015), <sup>b</sup>
Izena, Okinawa Trough	–	5.8–9.2	–	–	Myruiyama et al. (1993)
<b>Ridge flank crustal fluids</b>					
Juan de Fuca	9–17	0.05–0.09	–34	–	Lin et al. (2019)
North Pond	18–43	0.4–1.2	–	27–62	Meyer et al. (2016); Walter et al. (2018)
	Porewater DOC ( $\mu\text{M}$ )	POC (%)	$\delta^{13}\text{C}$ -POC (‰)		Reference
<b>Porewater from vent microbial mats</b>					
Loihi sea mount	68–179	0.28–0.63	–23 to –26		Bennet, Hansman, et al. (2011)
<b>Hydrothermal-influenced sinking particles</b>					
Tonga arc	–	>0.7–6	–23 to –24.3		Kim et al. (2016)

*Note.* Dashes indicate that samples were not measured for the particular component.

<sup>a</sup>Comita et al. (1984) reported a factor of 2–5 times more elevated than the non-vent fluid without providing the quantified values. We calculated by assuming a background non-vent fluid at the 21°N same as the 9°50'N by Bennet, Statham, et al. (2011). <sup>b</sup>Only the samples collected with HFPS bags were summarized. Readers are encouraged to see additional DOC data in Hawkes et al. (2015).

POM to DOM. The relatively low  $C/N_{DOM}$  (14) of the Red Rock sample corresponds well with the low  $C/N_{POM}$  (6.1).

The very high DOC in the Epsilon fluid sample may seem suspicious but is not exceptional because Hawkes et al. (2015) reported high DOC concentrations in various vent fields. Large filamentous microbial mats were observed at Epsilon (Figure 2f). The filamentous bacterial microbial mat matrix often consists of carbon-rich polysaccharides (Elshahed et al., 2003; Emerson et al., 2007; Moyer et al., 1995). The solubility of microbial mats may leach DOM with high C/N. The measured dissolved hydrogen concentration was too low (58  $\mu\text{mol/Kg}$ , Baumberger et al., 2014) to permit spontaneous abiotic formic acid production from  $\text{H}_2$  and  $\text{CO}_2$  (Supporting information, Figure S1, Tables S1 and S2). The abiotic formic acid reaction becomes thermodynamically favorable (exergonic, negative Gibbs free energy) when the environment contains a much higher  $\text{H}_2$  concentration (up to 30  $\text{mmol/Kg}$ ) (Figure S1). Therefore, the high DOC at Epsilon cannot be explained by abiotic formic acid production as those observed in the Von Damm vent field (Lang et al., 2010).

### 4.3. Impact on the Water Column

Within the particle-rich neutrally buoyant plume above the W. Mata vent field, POC and PN concentrations are twice those of the background seawater values (Table 4). Because there is no significant increase in Si concentrations in the plume samples relative to background seawater samples, we estimate a minimum of a hundred-time dilution in forming the plume from the volcanic activities and the diffuse vents. Such a dilution factor agrees with typical three-to-five orders of magnitude dilution of the source hydrothermal vent fluids observed elsewhere (e.g., Cowen et al., 2003; Edmonds et al., 2003; Field & Sherrell, 2000; Lupton et al., 1985). Using a dilution factor of a hundred, we estimated that a maximum of 0.03  $\mu\text{mol-C/L}$  of POC could have been injected from the diffuse vent fluids with high POC (e.g., Red Rock Ridge and Luo). Although we did not collect fluid samples for POC analysis from sites adjacent to molten lava, we can assume that POC could not escape from the high-temperature molten lava. Accordingly, the POC from diffuse vents accounts for no more than 13% of the averaged increase in POC (0.23  $\mu\text{mol-C/L}$ ) in the plume sample. Therefore, most of the elevated POC in the plume samples was likely produced within the plume as a result of enhanced biological activities induced by the elevated supply of reducing species—including dissolved iron, hydrogen sulfide, hydrogen, and ammonium—into the overlying water column as indicated by the anomaly in ORP (Table 4).

Earlier thermodynamic calculations (McCollom, 2000) predicted that the potential primary productivity by oxidizing the amount of reducing species in mid-ocean-ridge black smoker derived hydrothermal plume is up to 50 mg dry weight biomass/L-vent fluid, which is approximately 1.9  $\text{mmol-C/L}$  recalculated based on biomass to carbon conversion factor of 0.46 and a carbon molar mass of twelve (Battley, 1998). Our measured POC content ( $0.45 \pm 0.12 \mu\text{mol-C/L}$ ) in the plume is three orders of magnitude lower than that predicted by McCollom (2000). We attribute the low POC to the much lower sulfur, sulfide, and hydrogen concentrations measured in the W. Mata plume samples than the estimated concentrations used for McCollom (2000) model. Hence, we recalculated the potential “dark” carbon production based on the maximum measured concentrations of dissolved hydrogen (14,843 nM), total dissolved iron (6,180 nM), total manganese (359 nM), and elemental sulfur (61,646 nM) in the 2008 plume (Table 6) (Resing et al., 2011). For easy comparison, we directly applied the  $\Delta G^0$  reported in McCollom (2000) because of the differences in values of  $\Delta G^0$  due to the changes in pressure—120 bar at W. Mata versus 250 bar in the mid-ocean-ridge black smoker—is minimum. Our modeled results (Table 7) show that the reducing species in the 2008 plume can potentially support at least 4.2  $\mu\text{mol-C/L}$  primary carbon production or 0.11 mg-dry wet per kilogram of plume water. The thermodynamically estimated primary carbon production is similar to the combined increase in DOC and POC (4.5  $\mu\text{mol-C/L}$ ) observed in the 2009 plume. Our result demonstrates that chemoautotrophs were quite effective in utilizing available reducing compounds to producing organic carbon.

The DOC increase ( $\sim 4 \mu\text{M}$ ) was 11 times higher than the POC increase in the W. Mata plume than background seawater values. In comparison, there was an approximate 10–20  $\mu\text{M}$  DOC increase without a significant POC increase within the Loihi plume (Bennett, Hansman, et al., 2011) (Table 5). In

**Table 6**  
Estimated Chemical Compositions and Activities of W. Mata Plume Used for Thermodynamic Modeling

Chemical species	Unit	Value	Activity coefficient	Activity	log(activity)	Sources
SO <sub>4</sub> <sup>2-</sup>	mM	28	0.2	5.6E-03	-2.3	Average seawater value
O <sub>2(aq)</sub>	μM	150	1	1.5E-04	-3.8	From WOCE P15
H <sub>2(aq)</sub>	nM	14,843	1	1.5E-05	-4.8	Resing et al. (2011); Table S1
Fe <sup>2+</sup>	nM	6,180	0.2	1.2E-06	-5.9	Estimated from the highest PFe = 4,120 nM, and dissolve Fe = 1.5 × PFe and assuming all dissolved Fe is Fe <sup>2+</sup> in Resing et al. (2011); Table S1
Mn <sup>2+</sup>	nM	359	0.2	7.2E-08	-7.1	Assuming all TMn is Mn <sup>2+</sup> in Resing et al. (2011); Table S1
S <sub>0(s)</sub>	nM	41,646	-	-	-	Resing et al. (2011); Table S1
HCO <sub>3</sub> <sup>-</sup>	μM	2640	0.55	1.5E-03	-2.8	Resing et al. (2011); Table S1

Note. Mata plume used for thermodynamic modeling.

contrast, a maximum POC increase of 0.2 μmol/L was observed with a DOC decrease of 2 μM in the 9°15'N East Pacific Rise plumes, which the authors explained as adsorption of DOC by the rich particles in the plume to form POC and heterotrophic biological consumption of DOC (Bennett, Statham, et al., 2011). Further research is needed to unravel mechanisms, such as aging of a hydrothermal plume and microbial community structures, regulating the production and consumption of organic carbon, resulting in the different partitioning between the chemolithoautotrophic DOC and POC among various plumes.

## 5. Conclusion and Future Research Directions

The DOC, TDN, NO<sub>3</sub><sup>-</sup>, NH<sub>4</sub><sup>+</sup>, DON, and POC distribution in the West Mata eruptive vent field was summarized with a schematic drawing (Figure 4). The elevated DOC in the low-temperature diffuse vents is explained by biological production. Elevated POC and cell counts and depleted TDN support vigorous biological production within the seafloor and seafloor habitats associated with diffuse vents. The low C/N<sub>POM</sub>

**Table 7**  
Thermodynamic Calculation of Potential Metabolic Energy Available for A Few Selected Chemolithoautotrophic Reactions Occurring in the West Mata Hydrothermal Plume

Chemolithoautotrophic energy source	Overall reaction	ΔG <sup>0</sup> (Kcal)		ΔG (Kcal/mol reactant)	Limiting reactant	Limiting reactant concentration (M) <sup>a</sup>	Available energy (cal/kg plume fluid) <sup>b</sup>	Primary biomass potential (mg dry wt/kg plume) <sup>c</sup>	Organic carbon production (μmol-C/kg plume) <sup>d</sup>
		ΔG <sup>0</sup>	lnQ						
Iron (II) oxidation	Fe <sup>2+</sup> + 1/4O <sub>2(aq)</sub> + 2.5H <sub>2</sub> O = Fe(OH) <sub>3</sub> + 2H <sup>+</sup>	-92	-32	-110	Fe <sup>2+</sup>	6.2E-06	0.61	0.017	0.64
Mn (II) oxidation	Mn <sup>2+</sup> + 1/2O <sub>2(aq)</sub> + H <sub>2</sub> O = MnO <sub>2(s)</sub> + 2H <sup>+</sup>	-1.8	-11	-8	Mn <sup>2+</sup>	3.6E-07	0.003	0.00007	0.0026
Hydrogen oxidation	H <sub>2(aq)</sub> + 1/2O <sub>2(aq)</sub> = H <sub>2</sub> O	-63	16	-55	H <sub>2(aq)</sub>	1.5E-05	0.73	0.08025	3.07
Sulfur oxidation	S <sub>0(s)</sub> + 3/2O <sub>2(aq)</sub> + H <sub>2</sub> O = SO <sub>4</sub> <sup>2-</sup> + 2H <sup>+</sup>	-129	8	-124	S <sub>0(s)</sub>	4.2E-06	0.47	0.013	0.49
							Sum	0.11	4.2

Note. Primary biomass potential is also calculated. Parameters used for the calculation are listed in Table 6.

<sup>a</sup>Limiting reactant concentrations are derived from the measured concentrations in the 2008 plume reported by Resing et al. (2011). <sup>b</sup>Assuming 90% of the limiting reactant can be utilized. <sup>c</sup>Energy to biomass conversion factor is ~9,100 cal/g for hydrogen oxidizers and ~36,400 cal/g for other microbes (McCollom, 2000).

<sup>d</sup>Biomass to organic carbon conversion factor = 0.46 (Battley, 1998).

in diffuse vent fluids further indicates freshly produced biomass. The DON concentrations in the diffuse vent fluids were similar to or slightly higher than those in recharging seawater. The higher  $C/N_{\text{DOM}}$  than  $C/N_{\text{POM}}$  supports preferential nitrogen removal during DOM formation. The  $^{13}\text{C}$ -enriched POC implies that resident organisms along the fluid flow paths utilized carbon-fixation pathways different from marine phytoplankton.

The potential primary organic carbon production estimated from the energy derived through the oxidation of hydrothermally discharged reducing species into the water column matches the elevated organic carbon concentrations relative to background seawater values. Overall, this study confirms enhanced production of organic matter at northeast Lau hydrothermal vent fields and that unsedimented oceanic crust associated with back-arc spreading volcanism is a net DOC and POC source for the deep ocean.

Future research should include DOC characterization at the molecular level. Large volume—up to several tens of liters—fluid samples are needed for functional group analysis by nuclear magnetic resonance spectroscopy, molecular composition analysis by electrospray ionization (ESI) coupled with a Fourier-transform ion-cyclotron-resonance mass-spectrometry (FT-ICR MS) or with an Orbitrap Fusion™ Tribrid™ Mass Spectrometer. The stable and radiocarbon isotopic composition analysis can be performed by accelerator mass spectrometry (AMS). Large-volume-fluid samplers are available for collecting samples at low-temperature sites (e.g., Cowen et al., 2012; Lin et al., 2020; McCarthy et al., 2010). There is a need to develop a large-volume-fluid sampler that is not only DOC-contamination-free but also allows the collection of extremely high temperature and high gas content fluids near molten lava and high-temperature vents.

### Data Availability Statement

The raw data are archived at BCO-DMO (<https://www.bco-dmo.org/project/836899>).

### Acknowledgments

The authors thank the remotely operated-vehicle Jason II and research vessel Thomas Thompson crews for their skillful support during the field sampling. The authors also thank Chih-Chiang Hsieh, Natalie Hamada, and Kathryn Hu for their fantastic lab support. This project was supported by N.S.F. (OCE0929881, J. P. Cowen and K. H. Rubin), the NOAA PMEL VENTS (now Earth-Ocean Interactions) Program and the Joint Institute for the Study of the Atmosphere and Ocean (JISAO) under NOAA Cooperative Agreement No. NA10OAR4320148, and the UH NASA Astrobiology Institute. The Ministry of Science and Technology of Taiwan award (MOST 107-2611-M-002-002, and MOST 108-2611-M-002-006 to H.-T. Lin). Ministry of Education (M.O.E.) Republic of China (Taiwan) 109L892601 to H.-T. Lin. SOEST contributions no. 11285, C-DEBI contribution no. 563. PMEL contribution no. 3996, JISAO contribution 2183. There are no real or perceived conflicts of interest for any author.

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