

# Physico-chemical data from samples collected along the coast of Louisiana, USA during 2018.

Website: <https://www.bco-dmo.org/dataset/745449>

Data Type: Other Field Results

Version: 1

Version Date: 2018-08-30

## Project

» [Collaborative Research: EAGER: Salinity-based selection between sister clades of abundant coastal bacterioplankton](#) (CoastalSAR11)

Contributors	Affiliation	Role
<a href="#">Thrash, J. Cameron</a>	Louisiana State University (LSU)	Principal Investigator, Contact
<a href="#">Kujawinski, Elizabeth</a>	Woods Hole Oceanographic Institution (WHOI)	Co-Principal Investigator
<a href="#">Ake, Hannah</a>	Woods Hole Oceanographic Institution (WHOI BCO-DMO)	BCO-DMO Data Manager

## Abstract

Physico-chemical data from samples collected along the coast of Louisiana, USA during 2018.

---

## Table of Contents

- [Coverage](#)
- [Dataset Description](#)
  - [Acquisition Description](#)
  - [Processing Description](#)
- [Parameters](#)
- [Instruments](#)
- [Deployments](#)
- [Project Information](#)
- [Funding](#)

---

## Coverage

**Spatial Extent:** N:29.9213 E:-89.5905 S:29.0799 W:-93.5676

**Temporal Extent:** 2018-01-27 - 2018-07-24

---

## Dataset Description

Physico-chemical data from samples collected along the coast of Louisiana in the Gulf of Mexico.

## Acquisition Description

Samples were collected manually by filling an acid-washed and autoclaved 20L carboy after three rinses. Temperature, pH, and salinity were taken using a handheld YSI. Cell counts were obtained by filtering water through a 2.7  $\mu\text{m}$  Whatman GF/D filter, fixing with 10% formaldehyde, placing on ice, and then counting using flow cytometry (Thrash et al., 2015, Hydrocarbon and Lipid Microbiology Protocols). Inorganic nutrients were measured at the University of Washington Marine Chemistry Laboratory after sequential filtration through 2.7 Whatman GF/D and 0.22  $\mu\text{m}$  Sterivex filters. Samples were initially placed on ice in the field, and then refrigerated until shipment with ice packs.

## Processing Description

## **Data Problem Report:**

pH is not reported for the first time points because a crack was noticed in the housing of the YSI cable that seemed to be associated with calibration. This was subsequently replaced and all values were also checked in the lab with a Fisherbrand pH meter.

## **BCO-DMO Data Processing Notes:**

- Reformatted columns to comply with BCO-DMO standards
- Replaced blank cells with nd
- Reformatted dates
- Added ISO DateTime column

[ [table of contents](#) | [back to top](#) ]

---

## **Parameters**

Parameter	Description	Units
Site	Site code	unitless
Lat	Latitude	decimal degrees
Lon	Longitude	decimal degrees
Date	Date; yyyy-mm-dd	unitless
Time_Central	Time Central USA; hh:mm	unitless
Offset	Time difference	unitless
Time_UTC	Time UTC; hh:mm	unitless
Temp	Temperature	Celsius
Cond	Conductivity	ms/cm
Salinity	Salinity	PSU
pH	pH	pH units
Cell_counts	Cell counts	count
PO4	Phosphate	uM
SiOH4	Silicate	uM
NO3	Nitrate	uM
NO2	Nitrite	uM
NH4	Ammonium	uM
ISO_DateTime_UTC	DateTime ISO UTC formatted	unitless

[ [table of contents](#) | [back to top](#) ]

---

## Instruments

<b>Dataset-specific Instrument Name</b>	Millipore Guava 5HT HPL benchtop flow cytometer
<b>Generic Instrument Name</b>	Flow Cytometer
<b>Dataset-specific Description</b>	Used for sampling
<b>Generic Instrument Description</b>	Flow cytometers (FC or FCM) are automated instruments that quantitate properties of single cells, one cell at a time. They can measure cell size, cell granularity, the amounts of cell components such as total DNA, newly synthesized DNA, gene expression as the amount messenger RNA for a particular gene, amounts of specific surface receptors, amounts of intracellular proteins, or transient signalling events in living cells. (from: <a href="http://www.bio.umass.edu/micro/immunology/facs542/facswhat.htm">http://www.bio.umass.edu/micro/immunology/facs542/facswhat.htm</a> )

<b>Dataset-specific Instrument Name</b>	YSI 556 MPS handheld meter
<b>Generic Instrument Name</b>	Salinity Sensor
<b>Dataset-specific Description</b>	Calibrated with salinity and pH standards immediately prior to use
<b>Generic Instrument Description</b>	Category of instrument that simultaneously measures electrical conductivity and temperature in the water column to provide temperature and salinity data.

<b>Dataset-specific Instrument Name</b>	Masterflex I/P peristaltic pump
<b>Generic Instrument Name</b>	Pump
<b>Dataset-specific Description</b>	Used for sampling
<b>Generic Instrument Description</b>	A pump is a device that moves fluids (liquids or gases), or sometimes slurries, by mechanical action. Pumps can be classified into three major groups according to the method they use to move the fluid: direct lift, displacement, and gravity pumps
<b>Deployments</b>	

## Coastal\_SAR11

<b>Website</b>	<a href="https://www.bco-dmo.org/deployment/745476">https://www.bco-dmo.org/deployment/745476</a>
<b>Platform</b>	shoreside Gulf of Mexico
<b>Start Date</b>	2018-01-27
<b>End Date</b>	2018-07-24

[ [table of contents](#) | [back to top](#) ]

---

## Project Information

**Collaborative Research: EAGER: Salinity-based selection between sister clades of abundant coastal bacterioplankton (CoastalSAR11)**

**Coverage:** Coastal Louisiana, northern Gulf of Mexico

NSF award abstract: Adaptation to new environments is a fundamental challenge for organisms, including microbes, in expanding their habitat range. It is important to investigate the cellular mechanisms underlying salinity tolerance in coastal bacterioplankton and their different responses to salinity in nature because (i) it will provide fundamental understanding

for how microorganisms evolve to inhabit environments with different salinities, and (ii) alterations in coastal salinity are connected to climate change, so the way these alterations affect abundant coastal microorganisms also alters the biogeochemical cycling of, e.g., carbon. The project will examine microbial adaptations to salinity and determine how changes in salinity affect microbial metabolism using two closely related groups of abundant coastal bacterioplankton as model taxa. In addition, the research will continue and expand microbiology Course-based Undergraduate Research Experiences (mCUREs) in high-throughput cultivation and microbial characterization at the Louisiana State University. Sections of freshman biology laboratories will learn how to isolate, characterize, and molecularly identify microorganisms from local aquatic systems. mCURE sections will lead to newly isolated strains, genome sequences, and physiological data, these results will be published with the contributing students as co-authors. The relative success of mCURE sections will be assessed compared to traditional freshman biology sections. mCURE sections will offer unique opportunities for LSU students by creating excitement about research through discovery of new organisms and generating knowledge of the coastal habitats that are essential to the livelihood of the Gulf Coast. The evolutionary transition between salt- and freshwater environments occurs rarely in microorganisms. In one of the most abundant aquatic groups, SAR11, the transition between salt- and freshwater environments has happened only once: all freshwater SAR11 belong to subclade IIIb/LD12, which has also been found to inhabit coastal environments where salinity varies widely. The first reported isolates of the SAR11 freshwater clade LD12 and a member of the sister clade IIIa from the same region are now available. These pure culture representatives provide a powerful model for experimentally investigating adaptations to new environments in microorganisms, specifically (i) the genomic pathway and regulatory distinctions that arise during the evolutionary transition from marine to freshwater environments, and (ii) the physiological mechanisms that underlie the ecological restrictions imposed on microorganisms by ionic strength in coastal and freshwater environments. Furthermore, because these organisms have distinct differences in metabolic potential, the isolates facilitate testing (iii) the effects of changing coastal salinity on microbial contributions to other biogeochemical cycles, such as that for carbon. The project will test the hypothesis that the relative ionic strength tolerances between the sister lineages (LD12, IIIa) result from fundamental differences in metabolic flexibility at a genomic and regulatory level. To do so it will assess transcriptional and metabolic responses to varied ionic strength for both taxa and measure the distribution and activity of both groups in nature to translate laboratory findings to the field. The research will provide new understanding of LD12 habitat range and insights into how the "freshwater" lineage evolved from a SAR11 common ancestor. The project will also more generally provide important information on microbial responses to salinity changes in coastal systems and the evolutionary paths separating freshwater and marine microorganisms. This award is co-funded by Biological Oceanography, Division of Ocean Sciences in the Directorate for Geosciences and by Systems and Synthetic Biology, Division

of Molecular and Cellular Biosciences in the Directorate for Biological Sciences.

[ [table of contents](#) | [back to top](#) ]

---

## Funding

Funding Source	Award
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1747681</a>
<a href="#">NSF Division of Ocean Sciences (NSF OCE)</a>	<a href="#">OCE-1747722</a>

[ [table of contents](#) | [back to top](#) ]