Sediment geochemical and microbial activity data collected on R/V Oden along the East Siberian Arctic Shelf from 2014 (ESAS Water Column Methane project)

Website: https://www.bco-dmo.org/dataset/660527

Data Type: Cruise Results

Version: 1

Version Date: 2016-10-04

Project

» The East Siberian Arctic Shelf as a Source of Atmospheric Methane: First Approach to Quantitative Assessment (ESAS Water Column Methane)

Contributors	Affiliation	Role
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Abstract

Sediment geochemical and microbial activity data collected on R/V Oden along the East Siberian Arctic Shelf from 2014 (ESAS Water Column Methane project)

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Coverage

Spatial Extent: **N**:78.942 **E**:172.361 **S**:74.44 **W**:125.243

Temporal Extent: 2014 - 2014

Dataset Description

These data describe sediment geochemical and microbial activity from the Lappet Sea, East Siberian Arctic Shelf.

All of the methods used to determine concentrations and calculate rates of activity are given in the following papers: Joye S.B. et al. 2010; Joye, S. B. et al. 2010 and Orcutt, B. N. et al. 2005.

Acquisition Description

Acquisition methods are described in the following publication: Orcutt, B.N. et al. 2005

Core sectioning, porewater collection and analysis

At each sampling site, sediment sub-samples were collected for porewater analyses and at selected depths for microbial rate assays (AOM, anaerobic oxidation of methane oxidation; methanogenesis (MOG) from bicarbonate and acetate). Sediment was expelled from core

liner using a hydraulic extruder under anoxic conditions. The depth intervals for extrusion varied. At each depth interval, a sub-sample was collected into a cut-off syringe for dissolved methane concentration quantification. Another 5 mL sub-sample was collected into pre-weighed and pre-combusted glass vial for determination of porosity (determined by the change in weight after drying at 80 degrees celsius to a constant weight). The remaining material was used for porewater extraction. Sample fixation and analyses for dissolved constituents followed the methods of Joye et al. (2010).

Microbial Activity Measurements

To determine AOM and MOG rates, 8 to 12 sub-samples (5 cm3) were collected from a core by manual insertion of a glass tube. For AOM, 100 uL of dissolved 14CH4 tracer (about 2,000,000 DPM as gas) was injected into each core. Samples were incubated for 36 to 48 hours at in situ temperature. Following incubation, samples were transferred to 20 mL glass vials containing 2 mL of 2M NaOH (which served to arrest biological activity and fix 14CO2 as 14C-HCO3-). Each vial was sealed with a teflon-lined screw cap, vortexed to mix the sample and base, and immediately frozen. Time zero samples were fixed immediately after radiotracer injection. The specific activity of the tracer substrate (14CH4) was determined by injecting 50 uL directly into scintillation cocktail (Scintiverse BD) followed by liquid scintillation counting. The accumulation of 14C product (14CO2) was determined by acid digestion following the method of Joye et al. (2010). The AOM rate was calculated using equation 1:

AOM Rate = [CH4] x alphaCH4 /t x (a-14CO2/a-14CH4) (Eq. 1)

Here, the AOM Rate is expressed as nmol CH4 oxidized per cm3 sediment per day (nmol cm-3 d-1), [CH4] is the methane concentration (uM), alphaCH4 is the isotope fractionation factor for AOM (1.06; (ALPERIN and REEBURGH, 1988)), t is the incubation time (d), a-14CO2 is the activity of the product pool, and a-14CH4 is the activity of the substrate pool. If methane concentration was not available, the turnover time of the 14CH4 tracer is presented.

Rates of bicarbonate-based-methanogenesis and acetoclastic methanogenesis were determined by incubating samples in gas-tight, closed-tube vessels without headspace, to prevent the loss of gaseous 14CH4 product during sample manipulation. These sample tubes were sealed using custom-designed plungers (black Hungate stoppers with the lip removed containing a plastic “tail” that was run through the stopper) were inserted at the base of the tube; the sediment was then pushed via the plunger to the top of the tube until a small amount protruded through the tube opening. A butyl rubber septa was then eased into the tube opening to displace sediment in contact with the atmosphere and close the tube, which was then sealed with a open-top screw cap. The rubber materials used in these assays were boiled in 1N NaOH for 1 hour, followed by several rinses in boiling milliQ, to

leach potentially toxic substances.

A volume of radiotracer solution (100 uL of 14C-HCO3- tracer (~1 x 107 dpm in slightly alkaline milliQ water) or 1,2-14C-CH3COO- tracer (~5 x 107 dpm in slightly alkaline milliQ water)) was injected into each sample. Samples were incubated as described above and then 2 ml of 2N NaOH was injected through the top stopper into each sample to terminate biological activity (time zero samples were fixed prior to tracer injection). Samples were mixed to evenly distribute NaOH through the sample. Production of 14CH4 was quantified by stripping methane from the tubes with an air carrier, converting the 14CH4 to 14CO2 in a combustion furnace, and subsequent trapping of the 14CO2 in NaOH as carbonate (CRAGG et al., 1990; CRILL and MARTENS, 1986). Activity of 14CO2 was measured subsequently by liquid scintillation counting.

The rates of Bi-MOG and Ac-MOG rates were calculated using equations 2 and 3, respectively:

Bi-MOG Rate = [HCO3-] x alphaHCO3/t x (a-14CH4/a-H14CO3-) (Eq. 2)

Ac-MOG Rate = [CH3COO-] x alphaCH3COO-/t x (a-14CH4/a-14CH314COO-) (Eq. 3)

Both rates are expressed as nmol HCO3- or CH3COO-, respectively, reduced cm-3 d-1, alphaHCO3 and alphaCH3COO- are the isotope fractionation factors for MOG (assumed to be 1.06). [HCO3-] and [CH3COO-] are the pore water bicarbonate (mM) and acetate (uM) concentrations, respectively, t is incubation time (d), a-14CH4 is the activity of the product pool, and a-H14CO3 and a-14CH314COO are the activities of the substrate pools. If samples for substrate concentration determination were not available, the substrate turnover constant instead of the rate is presented.

For water column methane oxidation rate assays, triplicate 20 mL of live water (in addition to one 20 mL sample which was killed with ethanol (750 uL of pure EtOH) before tracer addition) were transferred from the CTD into serum vials. Samples were amended with 2 x 10^6 DPM of 3H-labeled-methane tracer and incubated for 24 to 72 hours (linearity of activity was tested and confirmed). After incubation, samples were fixed with ethanol, as above, and a sub-sample to determine total sample activity (3H-methane + 3H-water) was collected. Next, the sample was purged with nitrogen to remove the 3H-methane tracer and a sub-sample was amended with scintillation fluid and counted on a shipboard scintillation counter to determine the activity of tracer in the product of 3H-methane oxidation, 3H-water. The methane oxidation rate was calculated as:

MOX Rate = [methane concentration in nM] x alphaCH4/t x (a-3H-H2O/a-3H-CH4-) (Eq. 3)

Processing Description

BCO-DMO Data Processing Notes:

- filled in blank cells with "nd"
- separated month and year into two columns
- converted lat/lons to decimal degrees
- replaced the code "MUC" with it's complete definition "multiple core"
- replaced the code "BDL" with it's complete definition "below defined level"

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Related Publications

Joye, S. B., Bowles, M. W., Samarkin, V. A., Hunter, K. S., & Niemann, H. (2010). Biogeochemical signatures and microbial activity of different cold-seep habitats along the Gulf of Mexico deep slope. Deep Sea Research Part II: Topical Studies in Oceanography, 57(21-23), 1990–2001. doi:10.1016/j.dsr2.2010.06.001

Joye, S. B., MacDonald, I. R., Leifer, I., & Asper, V. (2011). Magnitude and oxidation potential of hydrocarbon gases released from the BP oil well blowout. Nature Geoscience, 4(3), 160–164. doi:10.1038/ngeo1067

Orcutt, B., Boetius, A., Elvert, M., Samarkin, V., & Joye, S. B. (2005). Molecular biogeochemistry of sulfate reduction, methanogenesis and the anaerobic oxidation of methane at Gulf of Mexico cold seeps. Geochimica et Cosmochimica Acta, 69(17), 4267–4281. doi:10.1016/j.gca.2005.04.012

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Parameters

Parameter	Description	Units
station	Station where sampling occurred	unitless
collection_type	Method used to collect samples	unitless
year	Year of sampling; YYYY	unitless
month	Month of sampling; mm	unitless
lat	Latitude	decimal degrees
lon	Longitude	decimal degrees
sediment_depth	Depth of sediment; negative depth values represent overlying water samples	centimeters
sample_ID	PI issued sample ID number	unitless
sed_CH4	Methane concentration in sediment	microns (uM)
AOM_rate	Anaerobic Oxidation of Methane; CH4 oxidized in sediment per day; Rates were measured at stations 13 and 23 only.	picomole per centimeter per day (pmol/cm/day)
turnover_14_CH3COO_MOG	14-CH3COO methanogenesis turnover; Rates were measured at stations 13 and 23 only.	percent
turnover_H14_CO3_MOG	H14-CO3 methanogenesis turnover; Rates were measured at stations 13 and 23 only.	percent
turnover_SRR	Sulfate reduction methanogenesis turnover; Rates were measured at stations 13 and 23 only.	percent

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Instruments

Dataset-specific Instrument Name	CTD
Generic Instrument Name	CTD profiler
Dataset-specific Description	Used to collect water column samples
Generic Instrument Description	The Conductivity, Temperature, Depth (CTD) unit is an integrated instrument package designed to measure the conductivity, temperature, and pressure (depth) of the water column. The instrument is lowered via cable through the water column and permits scientists observe the physical properties in real time via a conducting cable connecting the CTD to a deck unit and computer on the ship. The CTD is often configured with additional optional sensors including fluorometers, transmissometers and/or radiometers. It is often combined with a Rosette of water sampling bottles (e.g. Niskin, GO-FLO) for collecting discrete water samples during the cast. This instrument designation is used when specific make and model are not known.

Dataset-specific Instrument Name	Multiple core
Generic Instrument Name	Multi Corer
Dataset-specific Description	Core used in sampling
Generic Instrument Description	The Multi Corer is a benthic coring device used to collect multiple, simultaneous, undisturbed sediment/water samples from the seafloor. Multiple coring tubes with varying sampling capacity depending on tube dimensions are mounted in a frame designed to sample the deep ocean seafloor. For more information, see Barnett et al. (1984) in Oceanologica Acta, 7, pp. 399-408.

Dataset-specific Instrument Name	Liquid scintillation counter
Generic Instrument Name	Liquid Scintillation Counter
Dataset-specific Description	Used to determine activity of tracer substrate
Generic Instrument Description	Liquid scintillation counting is an analytical technique which is defined by the incorporation of the radiolabeled analyte into uniform distribution with a liquid chemical medium capable of converting the kinetic energy of nuclear emissions into light energy. Although the liquid scintillation counter is a sophisticated laboratory counting system used the quantify the activity of particulate emitting (β and a) radioactive samples, it can also detect the auger electrons emitted from 51Cr and 125I samples.

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Deployments

SWERUS-C3

Website	https://www.bco-dmo.org/deployment/660539
Platform	R/V Oden
Report	http://www.swerus-c3.geo.su.se/index.php/expedition
Start Date	2014-07-01

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Project Information

The East Siberian Arctic Shelf as a Source of Atmospheric Methane: First Approach to Quantitative Assessment (ESAS Water Column Methane)

Coverage: East Siberian Arctic Shelf

We propose to study methane (CH4) release over the East Siberian Arctic shelf (ESAS), the largest (~10% of the world ocean shelf area) and the shallowest shelf (mean depth

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Funding

Funding Source	Award
NSF Division of Polar Programs (NSF PLR)	PLR-1023444

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