1		
3	1	Increase in mercury in Pacific vellowfin tuna
4	2	increase in mercury in racine yenowini tuna
5	2	Paul F. Drewnick ^{1,*} Carl H. Lamborg ² and Martin I. Horgan ³
7	4	Taur E. Dreviner ', Garrin Lamborg', and Martin J. Horgan
8	5	¹ University of Michigan Biological Station and School of Natural Resources and
9	6	Environment Ann Arbor MI 48109 IISA
10	7	² Woods Hole Oceanographic Institution Department of Marine Chemistry & Geochemistry
11 12	, 8	Woods Hole MA 02543 USA
12	g	³ 230 Northridge Dr. Oxford OH 45056
14	10	230 Northinuge D1., 0x1010, 011 +3050
15	10	*Corresponding Author: *F-mail: drevnick@umich.edu_Phone (734) 763-6280
16	12	corresponding ration. I mail arevinek@umen.edu. r none (751)705 0200.
17 18	12	Abstract
19	13 14	Mercury is a toxic trace metal that can accumulate to levels that threaten human and
20	15	environmental health Models and empirical data suggest that humans are responsible for
21	16	a great deal of the mercury actively cycling in the environment at present. Thus, we would
22	10	nredict that the concentration of mercury in fish should have increased dramatically since
23 24	18	the Industrial Revolution - Evidence in support of this hypothesis has been hard to find
25	10	however, and some studies have suggested that analyses of fish show no change in mercury
26	20	concentration By compiling and re-analyzing published reports on vellow fin tuna
27	20	(<i>Thunnus albacares</i>) caught near Hawai'i over the nast half century we find that the
28	21	concentration of mercury in these fish is currently increasing at a rate > 3.8 % per year
29 30	23	This rate of increase is consistent with a model of anthronogenic forcing on the mercury
31	23	cycle in the North Pacific and suggests fish mercury concentrations are keeping nace with
32	25	current loadings increases to the ocean. Future increases in mercury in vellow fin tuna and
33	26	other fishes can be avoided by reductions in atmospheric mercury emissions from point
34 25	27	sources
36	28	
37	29	Introduction
38	30	Mercury is a notent toxin that can accumulate to high concentrations in fish nosing a health
39	31	risk to humans who eat fish Methylmercury the predominant form of mercury in fish is
40 41	32	formed from mercuric ions (Hg[II]) by microbes and perhaps abiotically in waters and
41	00	is the second seco

sediments, then enters the base of the food web and increases in concentration with each 42 33 43 34 successive trophic level. Consumption of mercury-contaminated fish from gross pollution 44 events, e.g., in Minamata, Japan [1] where fish with concentrations as high as 36 ppm could 35 45 36 "easily be captured by hand", has resulted in severe neurologic damage in humans, most 46 37 acutely in children exposed prenatally via maternal fish consumption. In waters not 47 directly affected by local pollution, mercury concentrations in fish are typically < 1 ppm. 48 38 49 39 However, even at this lower level, prenatal exposure is associated with developmental 50 40 deficits [2]. In the U.S. annually, Trasande et al. [3] found that *c*. 300,000-600,000 children 51 are born with mercury concentrations in cord blood that exceed 5.8 µg/L, a value 41 52 associated with significant loss of IQ (intelligence quotient); the economic consequence of 42 53 54 which is estimated to be US\$8.7 billion annually in lost income. 43 55

Humans are exposed to methylmercury primarily via consumption of ocean fish [4],
and while models (see review in [5]) and empirical data [6] suggest an increase in mercury
content of the global ocean since preindustrial times, a corresponding increase in

methylmercury has not been shown for ocean fish. Local pollution of nearshore environments occurs where there are point-source water discharges, but for open ocean

the principal source of mercury is atmospheric deposition [7]. Atmospheric mercury is

- now dominated by human contributions (primarily from fossil fuel combustion and
- artisanal gold mining), and present-day rates of mercury deposition are 3-5x greater than
- natural (pre-anthropogenic) rates [8]. It had been thought that mercury pollution could
- only result in a negligible increase in mercury concentration in open ocean waters [9], but a
- recent synthesis of data from water column profiles of total mercury points to a 2.6x increase (since the 1500s) in waters shallower than 1000 m, globally [6].

Here we show for a commercially important species, the yellowfin tuna (Thunnus albacares), a temporal increase in mercury concentration. Mercury data for Pacific vellowfin tuna from waters near Hawai'i present a unique record in that the same population/location was sampled three different time periods over 37 years, muscle tissue samples were analyzed for Hg, and data were reported in peer-reviewed literature. A network of fish aggregation devices in Hawai'i have documented the high site fidelity of yellowfin tuna [10]. The following is a report of our compilation, re-analysis, and interpretation of the mercury data.

Methods

We compiled published reports on mercury in yellowfin tuna caught from waters of the North Pacific Ocean near Hawai'i during 1971 [11, 12], 1998 [13], and 2008 [14]. From each specimen, muscle tissue was subsampled and measured for total mercury. Data are comparable across studies because of strict adherence to quality assurance/quality control (OA/OC) that ensured accuracy of data. Muscle tissue samples from 1971 [11, 12] and 1998 [13] were acid digested and analyzed by cold-yapor atomic absorption spectrophotometry. Rivers et al. [11] reported the mean recovery of spiked samples was 101% and that the result for each sample was validated by separate analysis of methylmercury, indeed confirming that nearly all of total mercury is methylmercury [15]. Thieleke [12; see also 16] had each sample analyzed by two independent laboratories, with excellent agreement between laboratories, as the maximum deviation among samples was 6%. Kraepiel et al. [13] reported detailed OA/OC procedures, including sample handling and use of duplicates (mean relative percent difference of 4.7%), spikes (mean recovery of 96.7%), and a reference sample (mean concentration of 0.128 ppm with standard deviation of 0.0098 ppm on 6 samples) for analyses. Muscle tissue samples from 2008 [14] were analyzed by thermal decomposition, amalgamation, and atomic absorption spectrophotometry. Choy et al. [14; see also 17] also reported detailed QA/QC procedures, including analysis of duplicates (data only accepted if relative percent difference between duplicates was <5%) and certified reference materials (mean recoveries of DORM-3 and High Purity Standard Trace Metal Fish were 100.3% and 96.1%, respectively).

To compare Hg concentration among the three data sets (1971, 1998, and 2008), we used the analysis-of-covariance (ANCOVA) model, with body size as the covariate. ANCOVA is the standard parametric test for comparing a characteristic of groups of subjects while controlling for the effect of another variable on that characteristic. Controlling for the effect of body size when assessing mercury concentration among groups of fish is thus a classic use of ANCOVA. Fish from 22–76 kg were included in the analysis as this size range $(\pm 5 \text{ kg})$ was common to all three data sets. It was necessary to remove the fish < 22 kg,

28

29

93 because these fish did not adhere to the assumption of linearity. Mercury concentrations in

young tuna tend to be low, but highly variable [18]. A diet shift occurs in young tuna when

95 a critical body mass is developed that enables endothermic capability to allow access to

96 prey in deeper, colder water [19]. At a certain size (depending on species), likely due to

97 this ontogenetic diet shift, the mercury concentration versus size relationship conforms to 98 expectations (i.e. a linear relationship). Outliers, identified with Tukey box plots and

 $^{10}_{11}$ 99 confirmed by one-sided Grubbs' tests, were also removed from the data sets.

12 100 13 101 **Results**

The ANCOVA revealed that slopes of the relationships between mercury concentration and fish size were not statistically different among the three time periods ($F_{2,223} = 1.17$, P = 0.31; Fig. 1), but mercury concentrations were higher in 2008 than either 1971 or 1998

18 105 ($F_{2,225} = 11.6$, P < 0.0001; Tukey's HSD; Fig. 1). Accordingly, the average mercury

 9 106 concentration (least square mean ± std. error) was considerably higher in 2008 (0.336

107 ± 0.023) than in 1971 (0.229 ± 0.008), or 1998 (0.218 ± 0.008). Sample size (for 1971,

108 1998, and 2008, n = 111, 104, and 14, respectively) is incorporated in the ANCOVA, and
23 109 with the significant result, it can be ruled out – with 95% probability – that the effect of

sample year was due to chance. The fourteen data points from 2008 are elevated relative
 to the two other data sets. Note that we conducted statistical diagnostics and found five
 points among the three datasets with potentially high leverage or high influence. We

performed the ANCOVA without these points and found no change in the qualitative results
 of the ANCOVA (no difference in slopes, significant difference in intercepts).

30 115 31 116 **Dis**

116 Discussion 32 117 Our analysis, at least for the early part of the record, is in agreement with that of Kraepiel et 33 al. [13], who found no change in tuna mercury between the same 1971 and 1998 datasets. 118 34 This conclusion led Kraepiel et al. [13] to hypothesize that methylmercury forms from 119 35 36 120 mercury naturally occurring in deep waters, sediments, or possibly hydrothermal vents 37 121 and is therefore largely natural. Subsequently developed independent lines of evidence 38 122 have suggested that vents are not strong enough sources of mercury to supply foodwebs 39 123 [20], that fish do acquire methylated mercury from shallow depths in the ocean [21], and 40 124 that seawater mercury concentrations are increasing ocean-wide [6], including near 41 42 125 Hawai'I [22]. Thus, we should expect to see changes in the concentration of mercury in 43 126 tuna now and in the foreseeable future.

44 127 Such changes are discernable in the recent part of the record, from 1998 to 2008, 45 which shows an increase in mercury concentration in tuna at a rate of \geq 3.8% per year, in 128 46 129 agreement with recent and modeled changes in mercury cycling in the North Pacific (Fig. 2). 47 48 130 Sunderland et al. [22] found a significant rise in mercury concentrations in seawater at all 49 131 depths (0-1000 m) from 2002 to 2006. The largest increases, modeled at 3% per year 50 132 between 1995-2006, are occurring in intermediate waters (150-1000 m), which in addition 51 133 to receiving mercury from atmospheric deposition to surface waters above (0-150 m) also 52 receive inputs associated with lateral flow of mercury-enriched waters from the coast of 53 134 54 135 the Northwest Pacific [22, 23]. Yellowfin tuna near Hawai'i spend most of their time in 55 136 surface waters or immediately below [24]. The agreement between the data/modeling by 56 137 Sunderland et al. [22] and the updated tuna record compiled and re-analyzed here provides 57

58

support for the alternative hypothesis that mercury and methylmercury concentrations in the ocean are increasing due to human activity and that anthropogenic methylmercury accumulates in important commercial fish.

A criticism of the Kraepiel et al. [13] study, that effects of fishing on oceanic food webs may have affected methylmercury accumulation in tuna [25], also applies here. Data for age, growth, and trophic level are not available for the individual fish in our synthesis. In the Pacific Ocean, the largest tunas have become more rare, but no detectable change in trophic level has occurred in any population [26]. Growth statistics of yellowfin tuna appear unchanged over the past half century [27], but the data aren't ideal for determining temporal trends. One could expect an increase in growth rate, as a density-dependent response to over-exploitation of the population [28]. With increased growth, there would be "growth dilution" of mercury [29]. To sum, there are no data to suggest Pacific yellowfin tuna have different growth rates or trophic level for the study period, but if either of those two factors have changed, it would likely be in a direction that would tend to lower the concentration of mercury in tuna, masking the effect of increased mercury loading to the system.

Because fish mercury concentrations appear to be increasing in step with modeled loadings, these data indicate that mercury concentrations in open ocean fish are responsive to anthropogenic mercury releases. Lamborg et al. [6] suggested that if atmospheric mercury emissions continue to increase, the deepwater sink will become 'overwhelmed' in the coming decades. Therefore, it should be expected that the rate of increase in mercury in surface waters (0-1000 m) will be greater than the rate of increase in emissions. Mercury contamination of ocean fish is a serious global health issue, now being addressed by the UN Minamata Convention on Mercury. Current goals of the convention – that aim to reduce atmospheric mercury emissions from point sources - would result in avoided increases in rates of atmospheric mercury deposition [30]. However, even if current deposition rates are maintained, North Pacific intermediate waters are expected to double in mercury concentration by 2050 [22]. Thus, more stringent reductions in emissions are necessary.

Supplemental Data

The data reported were previously published [11-14], but can also be found in Table S1.

Acknowledgments

PED was supported by the University of Michigan and the Fonds de recherche du Québec -Nature et Technologies. CHL was supported by NSF OCE-1129339, 1232760 and the Woods Hole Oceanographic Institution. Bruce Monson, Ed Swain, and anonymous reviewers provided constructive comments on earlier drafts of the manuscript. The authors declare no competing financial interests or any other conflicts of interest.

References

- [1] Harada M. 1995. Minamata disease: methylmercury poisoning in Japan caused by environmental pollution. Crit. Rev. Toxicol. 25:1-24.

2		
3 ⊿	182	[2] McKelvey W, Oken E. 2012. Mercury and Public Health: An Assessment of Human
4 5	183	Exposure. In Banks MS, Ed, Mercury in the Environment: Pattern and Process, University
6	184	of California Press, Oakland, CA, pp 267-287.
7	185	[3] Trasande L, Landrigan PJ, Schechter C. 2005. Public health and economic consequences
8	186	of methyl mercury toxicity to the developing brain. <i>Environ. Health Perspect.</i> 113:590-
9	187	596.
10	188	[4] Sunderland EM. 2007. Mercury exposure from domestic and imported estuarine and
12	189	marine fish in the U.S. seafood market. <i>Environ. Health Perspect.</i> 115:235-242.
13	190	[5] Black FI, Conaway CH, Flegal AR, 2012, Mercury in the marine environment. In Banks
14	191	MS. Ed. <i>Mercurv in the Environment: Pattern and Process</i> . University of California Press.
15	192	Oakland, CA, pp. 167-219.
16 17	193	[6] Lamborg CH, Hammerschmidt CR, Bowman KL, Swarr GL, Munson KM, Ohnemus DC,
18	194	Lam PI Heimhürger L-E Rijkenberg MIA Saito MA 2014 A global ocean inventory of
19	195	anthronogenic mercury based on water column measurements <i>Nature</i> 512:65-68
20	196	[7] Mason RP Fitzgerald WF Morel FMM 1994 The biogeochemical cycling of elemental
21	197	mercury: anthronogenic influences <i>Geochim Cosmochim Acta</i> 58:3191-3198
22	198	[8] Engstrom DR Fitzgerald WF Cooke CA Lamborg CH Drevnick PF Swain FB Balogh SL
23 24	199	Ralcom PH 2014 Atmospheric Hg emissions from preindustrial gold and silver
25	200	extraction in the Americas: a regulation from lake sediment archives. <i>Environ Sci</i>
26	200	Tochnol A8:6533-6543
27	201	[0] Hammond AI 1971 Morcury in the environment: natural and human factors. Science
28	202	171.788-789
29	203	[10] Itano DC, Holland KN, 2000, Movement and vulnerability of bigove (Thunnus abosus)
31	204	and vellow fin tuna (<i>Thunnus albacares</i>) in relation to FADs and natural aggregation
32	205	nointe Aquat Living Pasour 12:213-222
33	200	[11] Rivers IB Dearson IF Shultz CD 1972 Total and organic mercury in marine fish Rull
34	207	Environ Contam Toyicol 8:257-266
30 36	200	[12] Thieleke IR 1973 Mercury Levels in Five Species of Commercially Important Pelagic
37	205	Fish Takon from the Pacific Ocean Near Hawaii Dh D. Dissortation University of
38	210	Wisconsin-Madison
39	211	[12] Kraoniol AMI, Kollor K, Chin HB, Malcolm FC, Moral FMM, 2003, Sources and
40	212	variations of mercury in tuna <i>Environ Sci Tachnol</i> 37:5551-5558
41 12	213	[14] Choy CA, Donn RN, Kanaka H, Drazon JC, 2000. The influence of depth on moreury
43	214	lovels in polagic fishes and their prov. Proc. Natl. Acad. Sci. II.S.A. 106:13865-13869
44	215	[15] Ploom NS 1002 On the chamical form of more unit in adible fich and marine
45	210	invertebrate tissue Can I Fish Aquat Sci 40:1010 1017
46	217	[16] Douch CM. Thioloke ID. 1002. Total monouni content in vellouifin and higove tune. Dull
47 19	210	[10] DOUSH GM, THEERE JK. 1965. Total mercury content in yenowini and bigeye tuna. <i>Bull.</i>
40 49	219	Environ. Contain. Toxicol. 30:291-297.
50	220	[17] Choy CA. 2013. Pelagic Food web Connectivity in the North Pacific Subtropical Gyre: A
51	221	Combined Perspective from Multiple Biochemical Tracers and Diet. Ph.D. Dissertation,
52	222	University of Hawai I at Manoa.
53 54	223	[18] Lumont G, Viallex G, Lelievre H, Bobenrieth P. 1975. Mercury Contamination in Sea
54 55	224	Fish. Fisheries and Marine Service Lanada, Translation Series No. 33/3, Halifax, NS.
56	225	[19] Granam BS, Grubbs D, Holland K, Popp BN. 2006. A rapid ontogenetic shift in the diet
57	226	of juvenile yellowfin tuna from Hawai I. Mar. Biol. 150:647-658.
58		
59 60		
00		

2		
3	227	[20] Lamborg CH, Von Damm KL, Fitzgerald WF, Hammerschmidt CR, Zierenberg R, 2006
4	227	Marcury and monomethylmercury in fluids from Sea Cliff submarine hydrothermal field
5	220	Corda Pidgo Caonhus Pas Latt 23:L17606 doi:10.1020/2006CL026321
6 7	229	[21] Dum ID Down DN Drogon IC Chow CA Johnson MW 2012 Methylmoreuwy production
/ 8	230	[21] Blum JD, Popp BN, Drazen JC, Choy CA, Johnson MW. 2013. Methylmercury production
9	231	below the mixed layer in the North Pacific Ocean. <i>Nat. Geosci.</i> 6:879-884.
10	232	[22] Sunderland EM, Krabbenhoft DP, Moreau JW, Strode SA, Landing WM. 2009. Mercury
11	233	sources, distribution, and bioavailability in the North Pacific Ocean: insights from data
12	234	and models. <i>Glob. Biogeochem. Cycles</i> 23:GB2010, doi:10.1029/2008GB003425.
13	235	[23] Hammerschmidt CR, Bowman KL. 2012. Vertical methylmercury distribution in the
14	236	subtropical North Pacific Ocean. Mar. Chem. 132-133:77-82.
15	237	[24] Brill RW, Block BA, Boggs CH, Bigelow KA, Freund EV, Marcinek DJ. 1999. Horizontal
10	238	movements and depth distribution of large adult vellow fin tuna (<i>Thunnus albacares</i>)
18	239	near the Hawaijan Islands, recorded using ultrasonic telemetry. Implications for the
19	240	neur the nuwanan islands, recorded using all asome telemetry. Impleations for the
20	240	[2E] Donnor D. 2004. Whore is the more unit Environ. Sci. Technol. 29:124
21	241	[25] Keinier K. 2004. Where is the inercury? Environ. Sci. Technol. So.12A.
22	242	[26] Sibert J, Hampton J, Kleiber P, Maunder M. 2006. Biomass, size, and trophic status of
23	243	top predators in the Pacific Ocean. Science 314:1//3-1//6.
24	244	[27] Zhu G, Xu L, Dai X, Liu W. 2011. Growth and mortality rates of yellowin tuna, <i>Thunnus</i>
25	245	albacares (Perciformes: Scombridae), in the eastern and central Pacific Ocean. Zoologia
20 27	246	28:199-206.
28	247	[28] Polacheck T, Eveson JP, Laslett GF. 2004. Increase in growth rates of southern bluefin
29	248	tuna (<i>Thunnus maccoyii</i>) over four decades: 1960 to 2000. <i>Can. J. Fish. Aquat. Sci.</i>
30	249	61:307-322.
31	250	[29] Newman MC, Unger MA. 2003. <i>Fundamentals of Ecotoxicology</i> , 2 nd ed., CRC Press, Boca
32	251	Raton, FL.
33	252	[30] Selin NE, 2014, Global change and mercury cycling: challenges for implementing a
34	253	global mercury treaty <i>Environ Toxicol Chem</i> 33:1202-1210
36	254	Gibbar mereary dealy. Environ. Toxicol. Onem. 55.1262 1210.
37	254	Figure Captions
38	255	Figure captions
39	250	Fig. 1. Linear regressions of fish size (kg) versus mercury concentration (parts-per-infinion)
40	257	for three datasets for yellowin tuna from North Pacific waters near Hawai i; 19/1 (ref. 11,
41	258	12; black): Hg = $-0.0799 + 0.0068 \text{ x mass}, n = 111, r^2 = 0.413, P < 0.0001; 1998 (ref. 13;$
42	259	red): Hg = $-0.1619 + 0.0083 \text{ x mass}$, $n = 104$, $r^2 = 0.375$, $P < 0.0001$; 2008 (ref. 14; green):
43	260	Hg = $-0.0718 + 0.0093$ x mass, $n = 14$, $r^2 = 0.656$, $P = 0.0004$.
44 15	261	
46	262	Fig. 2. Least square mean (± std err) mercury concentration in yellowfin tuna from 1971
47	263	(black), 1998 (red), and 2008 (green) from waters near Hawai'i. Overlaid on the fish data
48	264	are mercury concentrations in seawater; gray squares (± std dev) represent point
49	265	estimates from integrated 1000-m profiles in the eastern North Pacific, and the solid grav
50	266	line represents modeled trends for intermediate waters (150-1000 m) basin wide (dashed
51	267	lines represent 95% confidence interval). Segurater data and model output are from
52	////	mes represent 7570 connuctice intervalis seawater uata and model output at C II OII
52	207	Sunderland et al [22]
53 54	268	Sunderland et al. [22].
53 54 55	268	Sunderland et al. [22].



