

1 **Optical map of the Genotype A1 WB C6 Giardia lamblia genome isolate**

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27 **Abstract**

28 The *Giardia lamblia* genome consists of 12 Mb divided among 5 chromosomes ranging in size
29 from approximately 1 to 4 Mb. The assembled contigs of the genotype A1 isolate, WB, were
30 previously mapped along the 5 chromosomes on the basis of hybridization of plasmid clones
31 representing the contigs to chromosomes separated by PFGE. In the current report, we have
32 generated an Mlul optical map of the WB genome to improve the accuracy of the physical map.
33 This has allowed us to correct several assembly errors and to better define the extent of the
34 subtelomeric regions that are not included in the genome assembly.

35

36 Key words: optical map, genome, pulsed field gel electrophoresis, subtelomeric variation

37

38 **Introduction**

39 The published sequence of the *Giardia lamblia* genotype A1 isolate, WB, consists of 11.7 Mb
40 divided among 306 contigs. Some of these contigs were joined into larger scaffolds, primarily by
41 “contig-joining” clones that linked these contigs even in the absence of continuous sequence
42 [1]. The results were supplemented by the use of multiple BAC clones that were end-sequenced
43 and physically mapped to specific chromosomes using pulsed field gel electrophoresis (PFGE).
44 Subsequent physical mapping studies using NotI-digested chromosomes of the genotype A1
45 isolate, BRIS/83/HEP/106, [2, 3] have made additional contributions to a complete physical
46 map. The current manuscript describes the use of optical mapping to refine and extend the
47 physical map of the WB isolate.

48

49 **Methods**

50 WB-C6 *Giardia* trophozoites were used to generate the optical map. The WB isolate was
51 originally axenized from a patient who most likely acquired his giardiasis in Afghanistan [4] and
52 subsequently has been cloned a number of times. The C6 clone from the laboratory of Dr. Fran
53 Gillin, UC San Diego, was used for the genome project and was also used for the optical
54 mapping described here. However, the WB isolate has been subjected to multiple rounds of
55 replication in the laboratory, so any changes that occur rapidly, such as changes in the
56 subtelomeric regions (STRs) may have resulted in differences between the organisms used for
57 the genome project and those used for the optical mapping.

58 Trophozoites were grown to confluence, pelleted and embedded in soft agarose as previously
59 described [5], followed by digestion with proteinase K in the presence of 1% Sarkosyl. The
60 optical mapping performed by OpGen (Gaithersburg, Maryland) [6, 7] consisted of melting the
61 agarose blocks followed by digestion with B-agarase. The DNA was mounted on a glass optical
62 mapping surface and digested in situ with Mlul so that the order of the individual restriction
63 fragments was maintained. The DNA was labeled with fluorescent YOYO-1 and imaged by
64 fluorescent microscopy, allowing the sizes of the fragments to be estimated by the intensity of
65 the fluorescent labeling. OpGen software was used to generate an Mlul restriction map and
66 then to compare that map with the available genomic sequence data. The map generated 150-
67 fold coverage. An algorithm that incorporates the length of the alignment and the quality of the
68 individual restriction fragments was used to overlay the sequence contigs (and secondarily the
69 scaffolds) onto the optical map. Individual sequence contigs could be flagged as problematic if
70 regions of match were followed by complete mismatch, suggesting an assembly error in the
71 individual sequence contigs.

72 Contigs that matched the optical map over their entire sequence were left intact. Those that
73 matched the optical map for only a portion of the map were split at the point of discrepancy
74 (c13, c27 and c29; Table 1). Conversely, if two contigs overlapped on the contig map and had
75 areas of sequence identity consistent with their positions on the optical map, these contigs
76 were joined. (17a and 53, 61 and 29a; Table 1).

77 **Results and Discussion**

78 The Mlul optical map yielded a genome size of 12.1 Mb divided among five chromosomes
79 ranging in size from 1.46 to 4.43 Mb. There were 1463 Mlul sites with an average restriction
80 fragment size of 8.29 kb. These chromosome sizes compare with PFGE estimates of 1.6 Mb to
81 3.5 Mb (Table 1). The total genome size estimated by the optical map is remarkably similar to
82 the 12 Mb estimated by PFGE [5] and 11.7 Mb by the published genome, which did not include
83 the rDNA repeats [1]. Although the total size was nearly identical to that estimated by PFGE,
84 the sizes of the individual chromosome estimates differed in that the chromosome 5 size had
85 been underestimated by PFGE (assuming that the optical map is indeed more accurate) and the
86 estimates for other chromosomes were smaller for the optical map than for PFGE.

87 The assembly of the published WBC6 genome consisted of 306 contigs in descending sizes by
88 increasing ID number. These contigs are identified in Genbank and in the Giardia genome
89 database as AACB02000001-AACB02000306. Many of the contigs were joined into scaffolds,
90 most frequently because of longer contig-joining clones. Contigs 1 through 70 with the
91 exception of 66 were placed onto the optical map (Fig 1). (A more detailed demonstration of
92 the placing of the contigs can be seen in Supplementary Figure 1). However, contigs 13, 27 and
93 29 were each split into two fragments. Contig 13a was placed onto chromosome 5, but contig
94 13b was not placed. The two fragments of contig 27 were placed on chromosomes 5 and 1,
95 respectively. Contig 29a was also placed onto chromosome 5, but contig 29b (39.8 kb) was not
96 placed on the map. There were nine places on the optical map with “negative gaps”, meaning
97 that there was an overlap between two contigs. In each case, we used BLAST comparisons of
98 the adjacent contig sequences to look for regions of sequence identity near the contig ends that
99 would allow them to be joined. We identified regions of sequence overlap for two of the nine

100 contig pairs. The two pairs of contigs with overlapping sequences were joined and then
101 analyzed using the OpGen software. This analysis confirmed that the joined contigs were
102 compatible with the optical map. For the remaining seven pairs of overlapping but unjoined
103 contigs, it is possible that misassembled sequences are present at one or both of the adjacent
104 ends; this remains to be determined.

105 The contigs smaller than contig 70 (34.2 kb) had too few Mlul sites to allow direct placement
106 onto the optical map. However, several were contained in a scaffold of the published genome.
107 These were left in the same positions if they did not contradict the optical map.

108 With the exception of the end gaps, 95% of the genome is covered by the optical map. The
109 genome assembly omitted the STRs entirely. A subsequent report [8] described the sequences
110 at most of the STRs, but the optical map provides the first accurate assessment of the extent of
111 the STRs not covered by the sequence assembly. The 10 end gaps ranged in size from 2 to 819
112 kb, with all but one being less than 45 kb in size. The exception is the 819 kb gap from one end
113 of chromosome 5, much of which consists of a repetitive region with Mlul fragments 4400-4600
114 bp in size. We believe this most likely represents the rDNA repeat region. Although the rDNA
115 repeat is 5566 bp in length and has only one Mlul site, this is the only repeat region in the
116 optical map compatible with prior data regarding the location of the rDNA sequence in
117 subtelomeric repeats. Prior data indicated that three genotype A1 isolates (Portland, ISR, and
118 CAT) varied greatly in the locations of the rDNA repeats [9]. These repeats are located in the
119 STRs of different chromosomes in different isolates. Even within different cloned lines of the ISR
120 isolate, the sizes of the rDNA-containing subtelomeric regions varied substantially [10]. This is

121 particularly remarkable since the chromosome-internal regions demonstrate very little
122 sequence variability.

123 A map placing the contigs and supercontigs onto a physical map that was derived by PFGE
124 hybridization studies has recently been published [11]. Many sections of the map in the current
125 study are identical to those obtained by PFGE, but there are a few notable differences. Some of
126 these differences resulted from the fact that the optical map split some of the contigs and
127 supercontigs (Sc) or allowed the placement of additional Sc between two existing Sc. For
128 example, Sc 1764 and 1761 were adjacent to each other at the right end of chromosome 4 on
129 the PFGE-based map, while Sc 1801 was placed between them on the optical map. The
130 differences not explained by splitting the contigs or supercontigs are found primarily in the
131 subtelomeric regions. For example, Sc 1769 and 1767 were located at the left ends of
132 chromosomes 1 and 2, respectively, in the PFGE-based map, but in the optical map, Sc 1769
133 was at the end of chromosome 2, while Sc 1767 was at the end of chromosome 1. We suggest
134 that these subtelomeric differences may be the result of using different isolates in the two
135 studies.

136 The optical map has provided independent verification for the majority of the contigs and
137 supercontigs of the WB Giardia genome as originally published [1]. In addition, it has corrected
138 several errors that resulted from misassembly. We believe the increased accuracy of the
139 current map will facilitate improved analysis of recombination and of gene expression that
140 depends on the local context.

141

142 **Table 1: Contig changes and chromosome sizes and coverage**

Joined Sequences¹	Nucleotide Sequence	Overlap Region	Sequence Length	Chromosome		
53_17a	53: 62321-1 17a: 124018-1	53: 2139-1 17: 124628 - 121880	184,200	2		
61_29a	61: 1- 46797 29a: 124018-1	61: 45748 - 46797 29: 124018 - 122599	169,395	5		
Split Sequences	Nucleotide Sequence	Sequence Length		Chromosome		
13a	1-209,610	209,610		5		
13b	209,611-266,103	56,493		N/A		
17a	1-124,018	124,018		2		
17b	124,019-203,025	85,161		1		
27a	1-108,674	108,674		5		
27b	108,675-148,504	39,830		1		
29a	1-123,648	123,648		5		
29b	123649-143,621	19,972		N/A		
Chrom	Size by PFGE(Mb)	Size by Optical Map	Total coverage	Total coverage excluding end gaps	Total internal gaps	Total end gaps
1	1.6	1.487	90.21%	93.02%	103,720	41,886
2	1.6	1.504	96.64%	99.29%	10,818	40,123
3	2.3	1.944	94.18%	96.68%	64,935	49,032
4	3.0	2.788	94.73%	95.94%	112,140	33,349
5	3.5	4.429	73.97%	92.84%	319,957	842,792
Total	12.0	12.096	87.05%	94.98%	611,570	1,007,182

143

144 The sequence numbers refer to the numbers of the 306 contigs in the assembly, matching the
 145 final three digits of the GenBank/GiardiaDB entries. Thus, sequence 17 would be
 146 AACB02000017. The a or b letter suffix is used for contigs that were split into a and b sections
 147 by the optical map.

148

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152 Environmental Health Sciences (P50 ES012742).

153

154 **Figure Legends**

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156 Fig 1. Optical maps of the five chromosomes are overlaid with the contigs from the WB genome
157 assembly. The upper number indicates the contig number, which matches the last three digits
158 of the GenBank/GiardiaDB entry. Thus, the GenBank entry for contig 1 is AACB02000001 The
159 number in parentheses indicates the supercontig/scaffold (sc) number from the published
160 assembly (www.giardiadb.org). The number shown consists of the last four digits of the
161 GiardiaDB entry (eg. Sc 1767 is identified as [CH991767](http://www.giardiadb.org/entry/CH991767) in the GiardiaDB web site). Chromosome
162 5 is shown on two lines and the long repeat region on the right represents what may be rDNA
163 repeats. The three scaffolds that were split by the optical map (1763, 1767, 1769) are shown in
164 unique colors to display the new locations.

165

166 Supplementary Fig 1: The placement of the contigs along each of the chromosomes is shown.
167 Sequence ID is the full name of the contig sequence in the GiardiaDB. “Contig” is the shortened
168 name which corresponds to the unique last three digits of the full name. The “length” column
169 gives the lengths of the individual contig sequence, while the gap gives the number of bp
170 between contigs. A negative gap indicates overlap between adjacent contigs. “Along the
171 chromosome” indicates the cumulative distance across the chromosome as determined by the
172 optical data.

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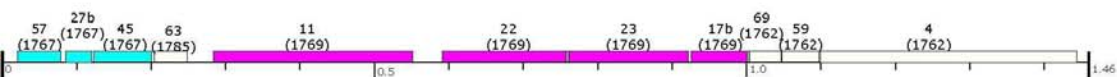
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200

Chromosome 1

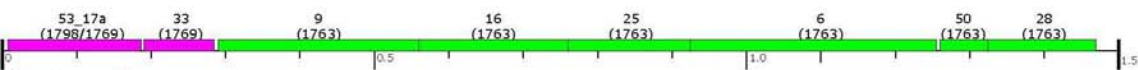


Scaffold 1763

Scaffold 1767

Scaffold 1769

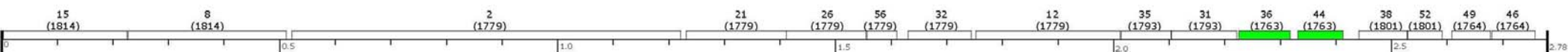
Chromosome 2



Chromosome 3



Chromosome 4



Chromosome 5

